



<https://theses.gla.ac.uk/>

Theses Digitisation:

<https://www.gla.ac.uk/myglasgow/research/enlighten/theses/digitisation/>

This is a digitised version of the original print thesis.

Copyright and moral rights for this work are retained by the author

A copy can be downloaded for personal non-commercial research or study,  
without prior permission or charge

This work cannot be reproduced or quoted extensively from without first  
obtaining permission in writing from the author

The content must not be changed in any way or sold commercially in any  
format or medium without the formal permission of the author

When referring to this work, full bibliographic details including the author,  
title, awarding institution and date of the thesis must be given

Enlighten: Theses

<https://theses.gla.ac.uk/>  
[research-enlighten@glasgow.ac.uk](mailto:research-enlighten@glasgow.ac.uk)

A Thesis Entitled

"STRUCTURE AND SYNTHESIS OF  
SOME LIVERWORT METABOLITES"

Submitted to  
The University of Glasgow  
for the Degree of Master of Science  
In the Faculty of Science

by

Ahmid SALAMANI

Chemistry Department      October 1988

ProQuest Number: 10999347

All rights reserved

INFORMATION TO ALL USERS

The quality of this reproduction is dependent upon the quality of the copy submitted.

In the unlikely event that the author did not send a complete manuscript and there are missing pages, these will be noted. Also, if material had to be removed, a note will indicate the deletion.



ProQuest 10999347

Published by ProQuest LLC (2018). Copyright of the Dissertation is held by the Author.

All rights reserved.

This work is protected against unauthorized copying under Title 17, United States Code  
Microform Edition © ProQuest LLC.

ProQuest LLC.  
789 East Eisenhower Parkway  
P.O. Box 1346  
Ann Arbor, MI 48106 – 1346

## ACKNOWLEDGEMENTS

I would like to thank my supervisor, Dr. J.D. Connolly, for his help and his expert guidance during the course of this research and for NMR spectra.

I also wish to thank the technical staff of the Chemistry Department for their assistance.

My thanks are also due to Professor G.W. Kirby for providing the facilities to carry out this work.

Finally, I would like to thank the Algerian Government for the financial support, also my parents, my wife and all members of my family for their forbearance during my research.

## SUMMARY

This thesis consists of four chapters. The first chapter, which is a General Introduction, deals with (i) the nature of secondary metabolites and (ii) the terpenoids and aromatic constituents of the liverworts. Chapter 2 describes several efforts to synthesise a lunularic acid derivative which occurs in the liverwort Plagiochila spinulosa. A Wittig reaction of 2- nitroveratraldehyde and 4- methoxybenzyl<sup>triphenyl</sup>phosphonium chloride afforded a mixture<sup>of</sup> stilbenes which on reduction gave 2- amino- 3,4,4'- trimethoxybibenzyl. This amino-bibenzyl was subjected to a Sandmeyer reaction in the hope of producing 2- cyano- 3,4,4'- trimethoxybibenzyl. Unfortunately, the desired compound was obtained only in very low yield. The major product was 3,5,6- trimethoxy- 9,10- dihydrophenanthrene, the result of a Pschorr cyclisation.

A similar Wittig route was used to prepare 2- bromo- 3,4,4'- trimethoxybibenzyl. Attempts at carboxylation resulted only in debromination.

Chapter 3 concerns a discussion of the constituents of Plagiochila spinulosa. Five dihydrophenanthrene derivatives were isolated together with methyl 2- methyl- 3,4- methylenedioxy- 6- methoxybenzoate and an unusual natural product spinuloplugin B which is a diterpenoid- bibenzyl conjugate. A synthesis of 2- hydroxy- 3,7- dimethoxy- 9,10,- dihydrophenanthrene is also described.

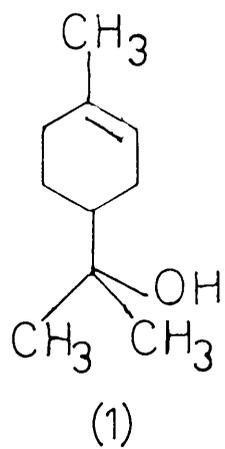
The final chapter considers the metabolites of Frullania tamarisci. In addition to the known compounds (-) frullanolide and tamarisc~~ol~~**ol**, a new dimeric sesquiterpenoid lactone was isolated.

# C O N T E N T S

	page
Summary	
<u>Chapter 1</u> <u>General Introduction</u>	
Introduction	1
Hepaticae	2
Monoterpenoids	3
Sesquiterpenoids	4
Diterpenoids	8
Aromatic compounds	11
<u>Chapter 2</u> <u>Synthesis of a Lunularic Acid Derivative</u>	
Introduction	14
Discussion	18
EXPERIMENTAL	28
<u>Chapter 3</u> <u>Plagiochilaea</u>	
Introduction	40
Discussion	42
EXPERIMENTAL	54
<u>Chapter 4</u> <u>Frullaniaceae</u>	
Introduction	62
Discussion	65
EXPERIMENTAL	68
References	70

C H A P T E R 1

GENERAL INTRODUCTION



The study of natural products has long been a subject of fascination for the organic chemist, foodstuffs, dyestuffs, medicines and stimulants, for example, all falling within its confines. However, it was only in the eighteenth century that chemists began to investigate the extracts obtained from natural sources. The study of natural products provided, in part, the stimulus for the development of modern chromatographic and spectroscopic methods. Nowadays structural elucidation requires only small amounts of material and involves a minimum of chemical transformation.

Natural products traditionally fall into two categories primary and secondary metabolites. Polysaccharides, proteins, fats, lipids, amino acids, nucleosides and nucleic acid are the fundamental building blocks of living matter and are considered to be the primary metabolites. The chemistry of the secondary metabolites, such as phenols, quinones, terpenes, alkaloids and various pigments has been a productive area of research for the organic chemist. Studies of terpenoids, for example, were instrumental in the early development of a synthesis of terpineol (1) in 1904 by Perkin<sup>1</sup>.

Organisms have adapted the production of metabolites, i.e. their enzyme activity to their living conditions, and the production of these compounds cannot be entirely fortuitous. Secondary metabolites, in particular those from plants, are considered to be an important factor in the co-evolution of

plants, animals and insects. The importance of chemical control is apparent in areas of survival e.g. defence chemicals, feedants, and antifeedants, the pheromones and sex attractants.

This thesis is concerned with the isolation of secondary metabolites from the Hepaticae (liverworts) and with the attempted synthesis of two of them.

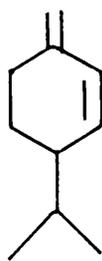
### HEPATICAE

The bryophytes are taxonomically placed between the algae and the pteridophytes and there are about 20,000 species in the world. They are morphologically divided into three main groups: (i) Musci (mosses, 14,000 species); (ii) Hepaticae (liverworts, 300 species) and (iii) Anthocerotae (hornworts, 300 species). Liverworts are the most interesting in chemical terms. This interest is probably associated with the presence of oil bodies in their cells. The Musci and Anthocerotae do not contain oil bodies and have been found to be of limited chemical interest.

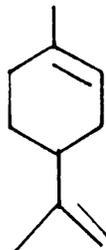
Liverworts are primitive plants, economically unimportant, and are easily overlooked. Some have found biological use e.g. Marchantia polymorpha<sup>2</sup> as a diuretic and Conocephalum conicum against gallstones<sup>2</sup>. Some species induce allergenic contact dermatitis<sup>3</sup> and inhibit the growth of micro-organisms<sup>4</sup>.



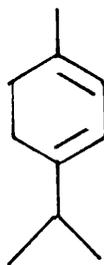
(2)



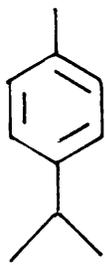
(3)



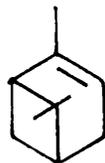
(4)



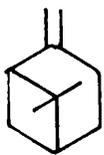
(5)



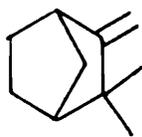
(6)



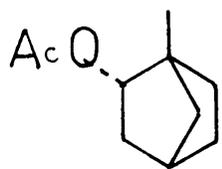
(7)



(8)



(9)



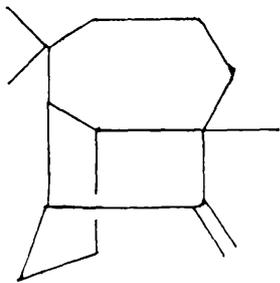
(10)

In 1905 the famous bryologist Karl Muller<sup>5</sup> reported that the oil bodies of the Hepaticae contained sesquiterpenoids. Further studies were delayed until 1956, when Fujita et al<sup>6</sup> reported the presence of sesquiterpenoids in the essential oil of Bazzania pompeana. The presence of chemically and pharmacologically interesting substances in the Hepaticae has made the liverworts the object of considerable attention in the past twenty-five years.

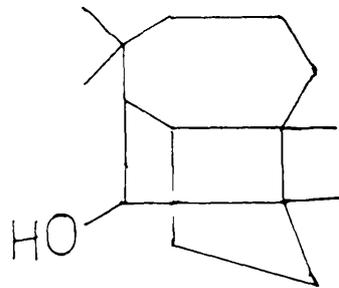
The chemistry of the bryophytes has been the subject of regular reviews. The most recent reviews are those by (i) Markham and Porter<sup>7</sup> reviewing the chemical constituents of the bryophytes, viz, lipids, terpenoids, flavonoids, lignans and bibenzyls published prior to 1978, (ii) Connolly<sup>8</sup> who dealt with terpenoid constituents of some European liverworts; (iii) Asakawa<sup>9</sup> who has produced a comprehensive review of liverwort constituents, (iv) Huneck<sup>10</sup> who dealt with all liverwort metabolites.

### MONOTERPENOIDS

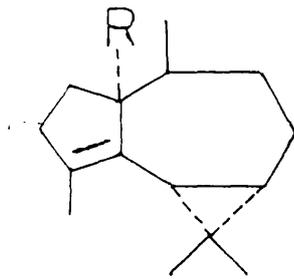
Reports on the occurrence of monoterpenoids in the Hepaticae are limited. Most of monoterpenoids are detected by gc or gcms. Asakawa and co-workers<sup>11</sup> reported the isolation of pure monoterpenoids from Conocephalum conicum species. The most common monoterpenoids are: Myrcene (2),  $\beta$ -phellandrene (3), limonene (4),  $\alpha$ -terpinene (5), p-cymene (6),  $\alpha$ -pinene (7),  $\beta$ -pinene (8) and camphene (9).



(11)



(12)



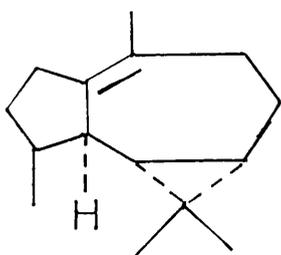
(13)  $R = H$

(14)  $R = OH$

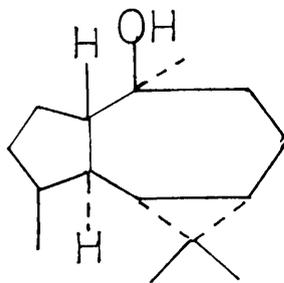
Both optical antipodes of monoterpenoids generally occur in higher plants often as racemates though some species synthesise only one of the two isomers<sup>12</sup>. In general the chiroptical properties of monoterpenoids found in the Hepaticae have not been clarified. Asakawa et al,<sup>11</sup> studied the chirality of monoterpenes from C. conicum which contains (-)-limonene (4) and (+)- bornyl acetate (10), while Jungermannia exsertifolia<sup>13</sup> contains (+)- limonene (ent - 4),  $\alpha$ -pinene (7) and camphene (9). Thus both enantiomers of limonene occur in liverworts as in higher plants but C. conicum and J. exsertifolia each produces only one of the two enantiomers. There remains much scope for further work in this field.

### SESQUITERPENOIDS

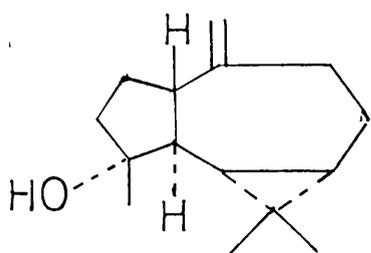
Liverworts are extremely rich in sesquiterpenoids many of which are enantiomeric to those found in higher plants. The first recognition of sesquiterpenes in liverworts was reported by Fujita et al<sup>6</sup>, in a study of the essential oil of Bazzania pompeana. The first positive identification of sesquiterpenes was reported in 1967 by Huneck and Klein<sup>14</sup> who isolated (-)- longifolene (11) and (-)- longiborneol (12) from Scapania undulata. The sesquiterpenoids (11) and (12) are the enantiomers of longifolene and longiborneol derived from Pinus species.



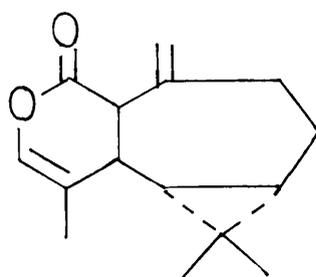
(15)



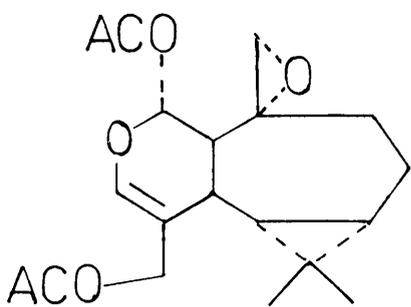
(16)



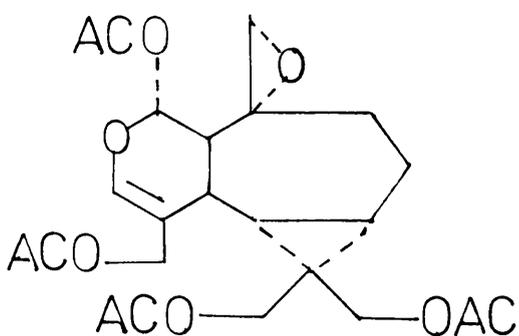
(17)



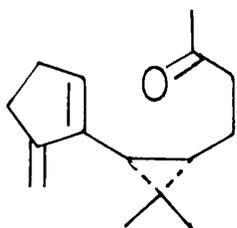
(18)



(19)



(20)



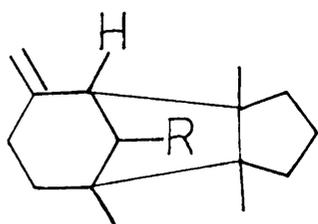
(21)

AROMADENDRANES AND SECOAROMADENDRANES

Aromadendrane-type sesquiterpenoids are mainly distributed in Mylia, Plagiochila and Porella species belonging to Jungermanniales. Cyclocolerenone (13)<sup>15</sup> and 1-hydroxycyclocolerenone (14)<sup>16</sup> from Plagiochila and Porella species are the simplest representatives in this type. Another variation of the aromadendrane skeleton is found amongst the metabolites of Plagiochila<sup>17,18</sup> species e.g. ladene (15), globulol (16) and spathulenol (17). The absolute configuration of these compounds is opposite to that found in higher plants.

Sesquiterpenoids with a secoaromadendrane skeleton are also found in liverworts. The first to be isolated from Plagiochila species was plagiochilide (18)<sup>18</sup>. It is a crystalline compound. Its <sup>1</sup>H n.m.r. showed an exomethylene grouping [ $\delta_{\text{H}}$  4.76, 4.92], two tertiary methyls [ $\delta_{\text{H}}$  1.06 (s, 6H)], a cyclopropane [ $\delta_{\text{H}}$  0.84(m), 0.43(dd, J10Hz.)], a vinyl methyl [ $\delta_{\text{H}}$  1.74(d, J2.0Hz.)] and a vinyl proton [ $\delta_{\text{H}}$  6.24 (d, J2.0Hz.)] and the i.r. indicated the presence of a lactone ( $\bar{\nu}_{\text{max}}$  1760  $\text{cm}^{-1}$ ). These spectroscopic properties indicated structure (18). The Plagiochilae proved to be a rich source of compounds with ent-2,3- secoaromadendrane skeleton<sup>19</sup> e.g. compounds (19, 20).

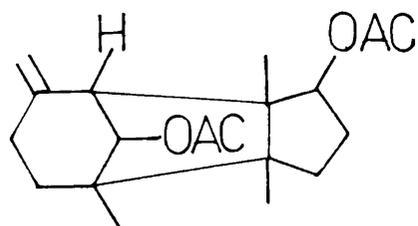
Taylorione (21) from Mylia taylorii<sup>20</sup> was established as an ent-1, 10- secoaromadendrenone. The structure and absolute configuration were determined by extensive chemical degradation and synthesis<sup>21</sup>.



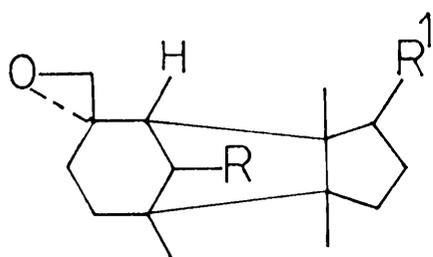
(22)  $R = OH$

(23)  $R = H$

(24)  $R = OAC$

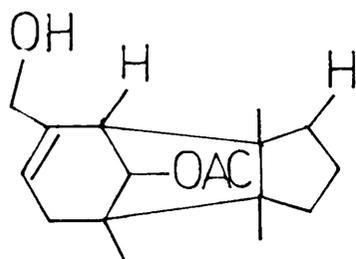


(25)

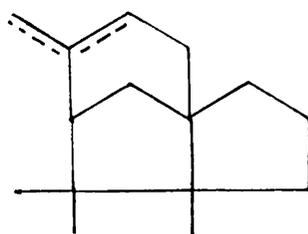


(26)  $R = OAC, R^1 = H$

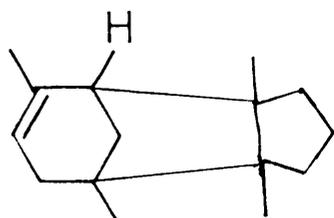
(27)  $R = R^1 = OAC$



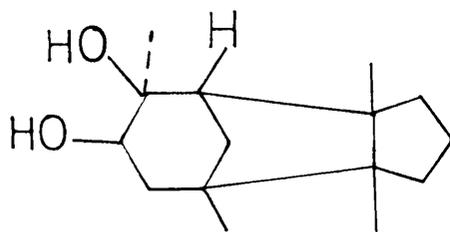
(28)



(29)



(30)



(31)

GYMNOMITRANES (BARBATANES) AND TRICHOTHECANES

Connolly et al<sup>21</sup> isolated a new sesquiterpene alcohol gymnomitrol (22) from Gymnomitrion obtusum together with six other sesquiterpenes with the same skeleton (23) - (28). These structures were determined by a combination of <sup>1</sup>H n.m.r. and Eu(~~fad~~)<sub>3</sub> - induced spectral shifts, decoupling experiments, and extensive chemical transformations.

Matsuo et al<sup>22</sup>, reported the isolation of new tricyclic sesquiterpenes  $\alpha$ - and  $\beta$ - pompene from Bazzania pompeana which were assigned the erroneous structure (29). The spectral data of  $\alpha$ - and  $\beta$ - pompenes were identical with those of  $\alpha$ - barbatene ( $\alpha$ - gymnomitrene) (30) and  $\beta$ - barbatene ( $\beta$ - gymnomitrene) (23) respectively. That  $\alpha$ - pompene is identical with  $\alpha$ - barbatene (30) has been shown by x-ray analysis of the p-bromobenzoate of the derived diol (31).

Connolly et al<sup>21</sup>, suggested that the gymnomitrene skeleton can be derived from an enzyme-bound farnesyl pyrophosphate which leads to a trichodiene-type intermediate and then to the gymnomitrane as shown in Scheme 1 .

Total syntheses<sup>23</sup> have confirmed the structures of ( $\pm$ )- gymnomitrol (22) and  $\alpha$ - and  $\beta$ - barbatenes (30, 23).

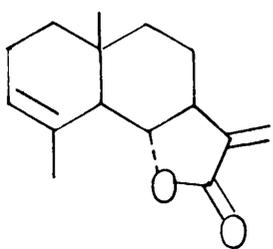


## EUDESMANES

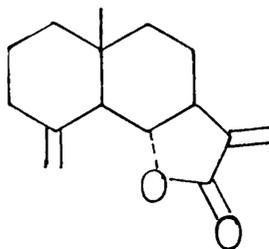
This type of sesquiterpenoid is very widely distributed in the Hepaticae, particularly in the Jungermanniales. The eudesmanolides often have biological activity. Ourisson<sup>24</sup> and co-workers isolated (-)- frullanolide (32) whose structure was confirmed by correlation with 6 $\beta$  11 $\beta$ - santonin (33) and by synthesis<sup>25</sup>. Frullanolide has been shown to be responsible for the high incidence of contact dermatitis among forestry workers.

Connolly and Thornton<sup>26</sup> isolated (-)- frullanolide (32),  $\alpha$ - cyclocostunolide (36), (+) costunolide (35) and  $\gamma$ - cyclocostunolide (34) from Scottish Frullania tamarisci. However,  $\beta$ - cyclocostunolide (37) could not be detected in the extract. It was later isolated<sup>27</sup> from the Japanese Frullania tamarisci subsp obscura together with the new eudesmane- type sesquiterpenoids, the hydroxy aldehyde (38) and 4 $\beta$ - hydroxy-eudesmanolide (39). Similar compounds of the ent- eudesmanolide series have been isolated from Frullania dilatata. It is interesting that F. dilatata biosynthesises ent- sesquiterpenoids while F. tamarisci elaborates compounds belonging to the normal series although these two species are morphologically similar.

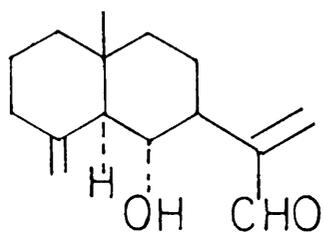
Two ent- eudesmanolides, ent-3- oxodiplophyllin (40) and ent-7- hydroxydiplopyllolide (41) occur in European Chiloscyphus polyanthos<sup>28.29</sup> together with known compounds.



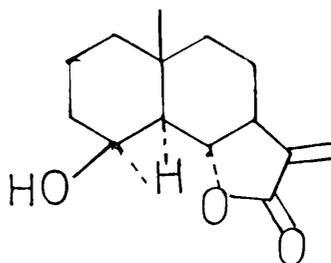
(36)



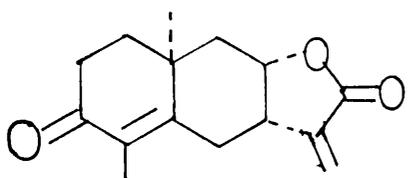
(37)



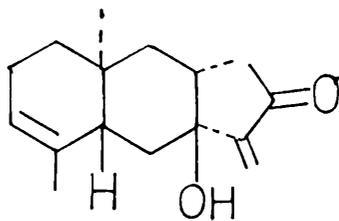
(38)



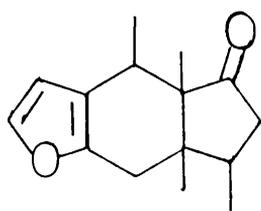
(39)



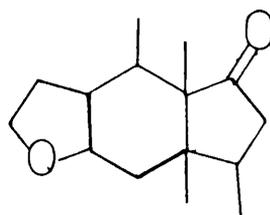
(40)



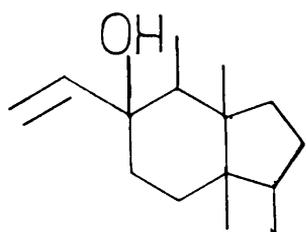
(41)



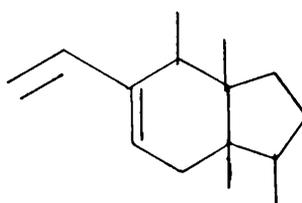
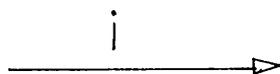
(42)



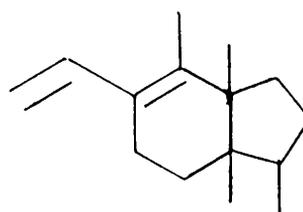
(43)



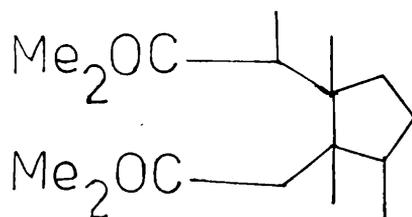
(44)



(45)



(46)



(47)

- (i) p-TsOH/C<sub>5</sub>H<sub>6</sub>
- (ii) O<sub>3</sub>, H<sub>2</sub>O<sub>2</sub>
- (iii) CH<sub>2</sub>N<sub>2</sub>

The structures (40) and (41) were determined on the basis of the  $\text{Eu}(\text{fod})_3$ -induced shifts in the  $^1\text{H}$  n.m.r. spectrum <sup>29</sup>.

### PINGUISANES

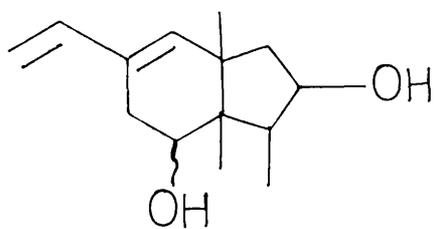
The first representative of this group, pinguisone (42)<sup>30</sup> was found in Aneura pinguis. The basic skeleton was deduced from comparison of the  $^1\text{H}$  n.m.r. of pinguisone (42) with that of its tetrahydroderivative (43). Later an x-ray analysis of its p-bromo-benzylidene derivative established the structure and absolute stereochemistry of (42)<sup>31</sup>.

Recently, further examples of this category have appeared. Pinguisenol (44) and  $\alpha$ -pinguisene (45) have been isolated from Porella vernicosa<sup>32</sup>. A dehydration of (44) with p-TsOH gave two conjugated hydrocarbons (45) and (46), whose ozonolysis without separation followed by methylation afforded dimethyl ester (47). (Scheme 2)

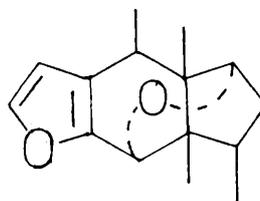
Three pinguisanes (48) - (50) have been isolated from European Porella platyphylla and their structures bases mainly on  $^1\text{H}$  and  $^{13}\text{C}$  n.m.r. spectroscopic evidence and spectroscopic correlation with the co-metabolite deoxypinguisone (52)<sup>33</sup>. Recent work<sup>34</sup> has shown that the structure (49) should be revised to (51).

### DITERPENOIDS

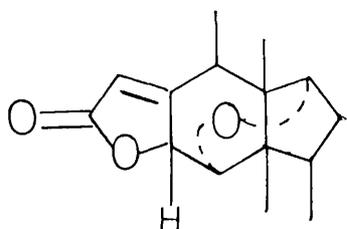
The diterpenoids are the second largest group of terpenoids in bryophytes. The representative types in this category are labdanes, pimaranes, kauranes, together with new types exemplified by sacculatanes, verrucosanes, dolabellanes and fusicoccanes.



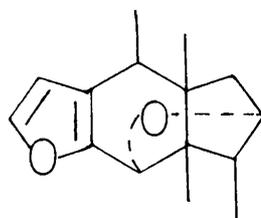
(48)



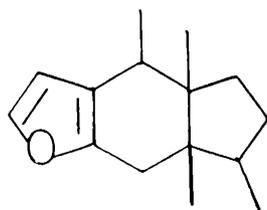
(49)



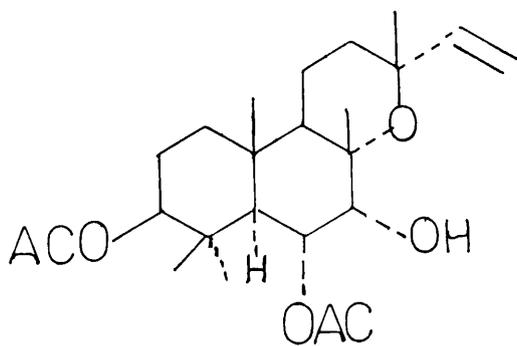
(50)



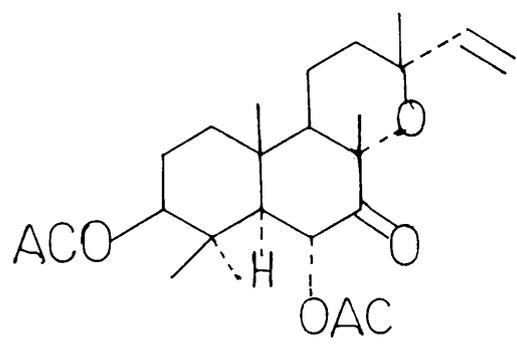
(51)



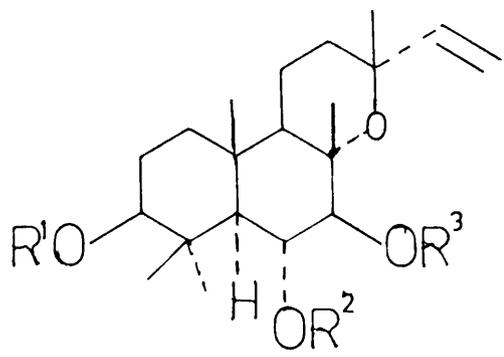
(52)



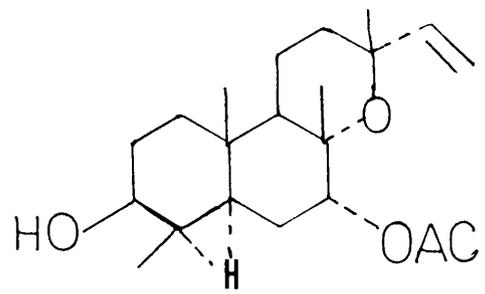
(53)



(54)



(55)  $R^1 = R^2 = AC, R^3 = H$



(57)

(56)  $R^1 = R^3 = AC, R^2 = H$

## LABDANES

The most recent examples of this skeletal type have been reported by Asakawa et al<sup>35</sup>. Five new labdane-type diterpenoids hamachilobenes A - E have been isolated from the liverwort Frullania hamachiloba. They were shown to be the manoyloxy derivatives (53) - (57) by a combination of <sup>1</sup>H and <sup>13</sup>C n.m.r. and CD spectra.

## KAURENES

All representatives from bryophytes belong to the entkaurenes series. Connolly and Thornton<sup>36</sup> isolated four ent-kaurenes from Jungermannia atrovirens

ent - 11 $\alpha$  - hydroxykaur - 16 - en - 15 one (58)

ent - 11 $\alpha$  - hydroxy - (16S) - kaurane - 15 - one (59)

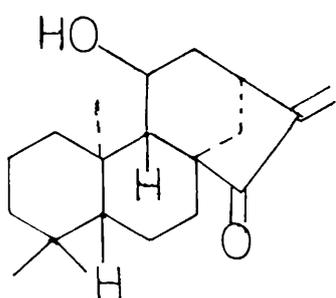
ent - kaur - 16 - ene - 11 $\alpha$ , 15 $\alpha$  - diol (60) and

ent - 15 $\alpha$  - acetoxykaur - 16 - ene - 11 $\alpha$  - ol (61).

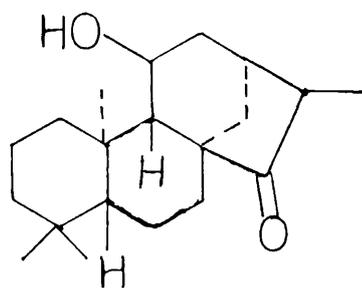
The structure of (60) was established by x-ray crystallographic analysis<sup>37</sup>. The ent - 15 $\alpha$  - hydroxykaura - 9 (11), 16 - diene - 6 - one (62) has recently been isolated from Czechoslovakian Nardia scalaris<sup>38</sup>, as well as being found in N. compressa<sup>39</sup>.

## SACCOLATENES

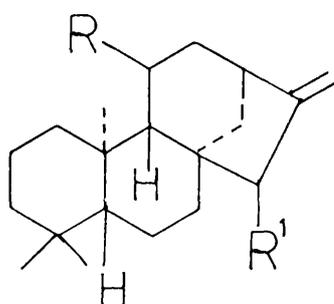
Two unique diterpene dialdehydes, the pungent sacculatal (63) and the non-pungent isosacculatal (64) have been isolated from Trichocoleopsis sacculata<sup>40</sup>. Their structures were determined by spectroscopic and chemical correlation with the sesquiterpene dialdehyde, polygodial (65).



(58)

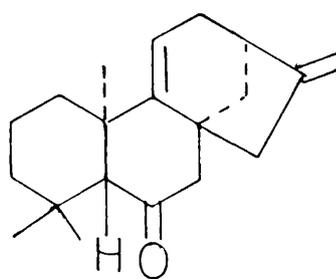


(59)

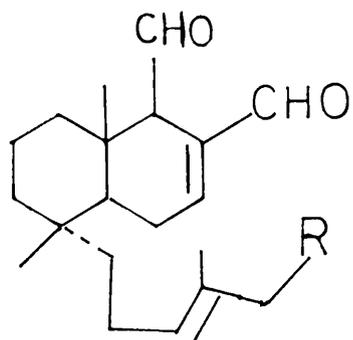


(60)  $R=R'=OH$

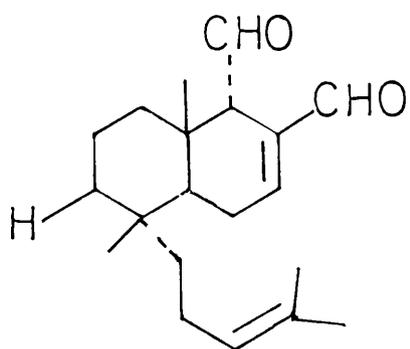
(61)  $R=OH, R'=OAC$



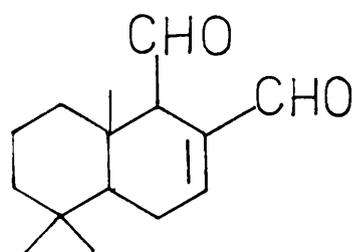
(62)



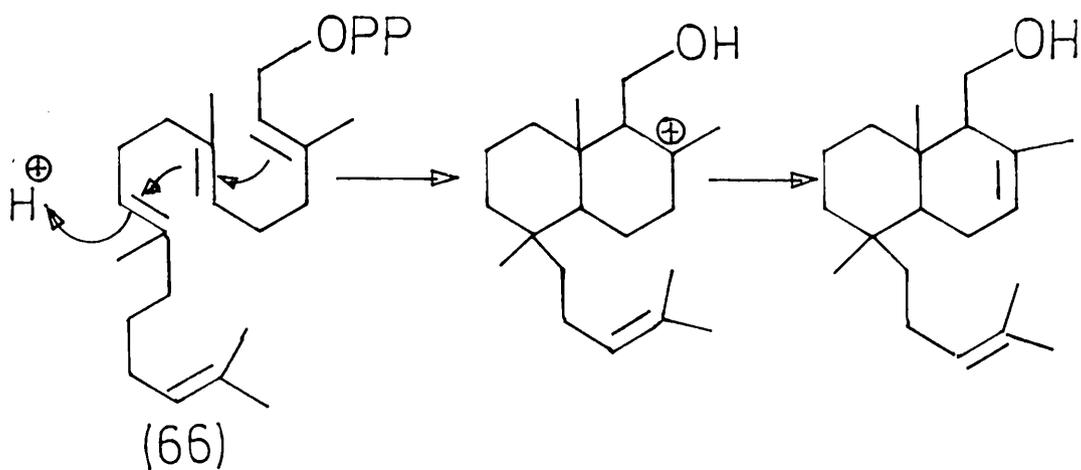
(63)  $R = H$



(64)



(65)



Scheme 3

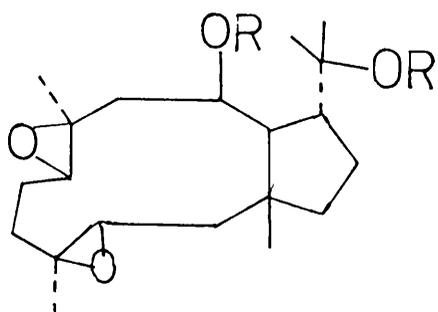
The sacculatanes are interesting from a biogenetic point of view. The ring system is the same as that of the drimene-type sesquiterpenes with an additional isoprene attached to C-15. They may be formed from geranyl-geranyl pyrophosphate (66) by a cyclisation mechanism analogous to that of the drimenes, (scheme 3).

### DOLABELLANES

In 1971, Huneck and Overton<sup>41</sup> isolated diterpenoids from Barbilophozia floerkei, B. lycopodioides, Anastrepta orcadensis and Gymnocolea inflata. However, it is only recently that Connolly<sup>42</sup> reported the occurrence of the dolabellane barbilycopodin (67) in the extract of Barbilophozia floerkei as the major constituent. The structure of (67) was identified by spectroscopic and chemical methods along with an x-ray analysis of the corresponding diol floerkein B (67a). This is the first report of a dolabellane from plants. This class has previously been detected in marine organisms<sup>43</sup>.

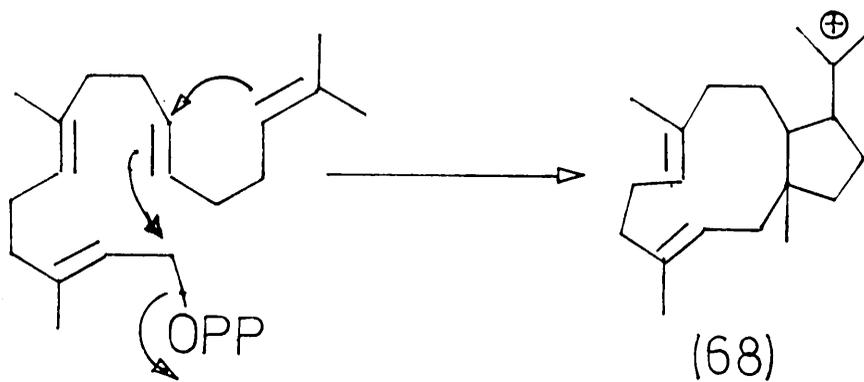
The biosynthesis of barbilycopodin involves cyclisation of all-transgeranyl-geranyl pyrophosphate as shown in scheme 4

Further cyclisation of (68) affords anadensin (69), a diterpenoid isolated from Anastrepta orcadensis<sup>44</sup>. Its structure was assigned on the basis of its <sup>1</sup>H and <sup>13</sup>C n.m.r. spectroscopy and Eu(fod)<sub>3</sub>-induced shifts. The stereochemistry of (69) was confirmed by an x-ray analysis.

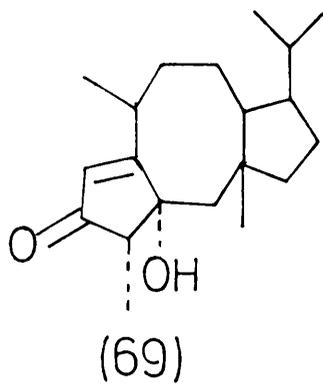


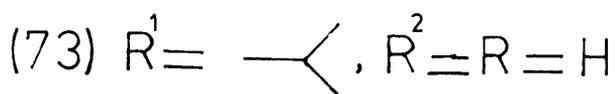
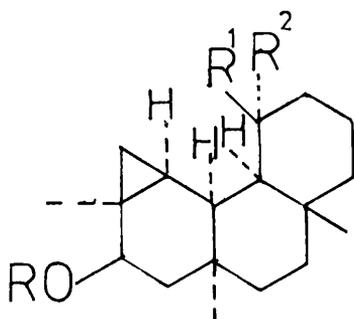
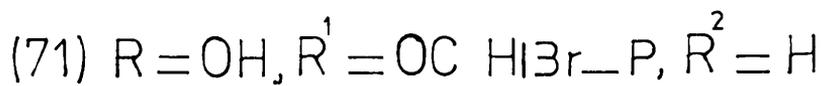
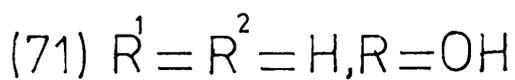
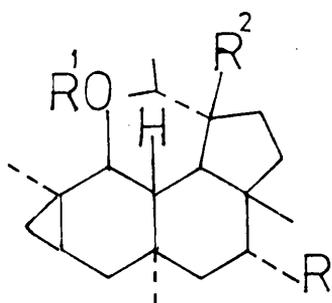
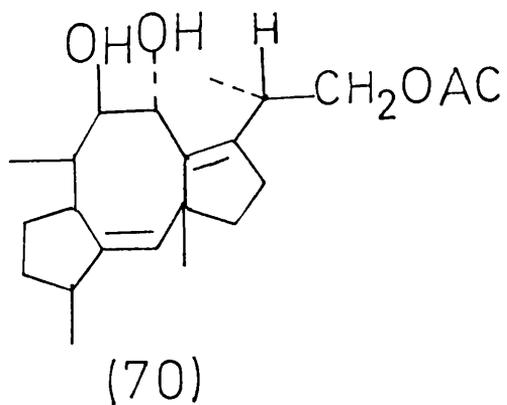
(67)  $R = \text{AC}$

(67a)  $R = \text{H}$



Scheme 4





This skeleton is unknown in higher plants, but has been found in fungal products e.g. fusicoccin (70)<sup>45</sup> and the cotylenins<sup>46</sup>.

### VERRUCOSANES

More deep-seated modification of the dolabellane skeleton leads to the verrucosanes which were isolated from Mylia verrucosa. The (-)- verrucosane - 2 $\beta$ , 9 $\alpha$  - diol (71)<sup>47</sup> was determined by spectroscopic and chemical data. An x-ray of the 2- bromobenzoate (72) established its stereochemistry.

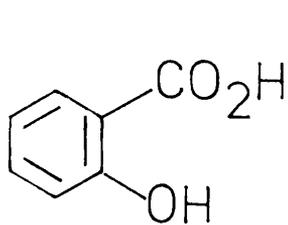
A new verrucosane-type sesquiterpenoid (73) has been isolated from *Plagiochila stephensoniana*<sup>48</sup>. The structure of (73) elucidated by extensive 2 D n.m.r. spectroscopy, was confirmed and its absolute configuration established by x-ray analysis of the p-bromobenzoyl derivative.

### AROMATIC COMPOUNDS

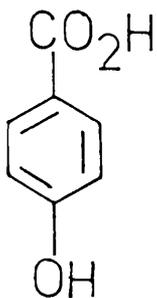
A wide range of aromatic metabolites of the Hepaticae has been reported.

### BENZOIC ACID DERIVATIVES

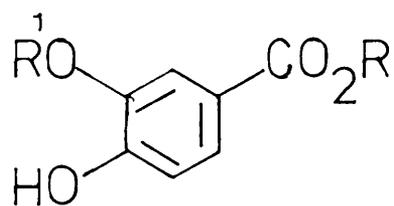
O- and p- hydroxybenzoic acids (74), (75) have been obtained from *Asterella lindenbergiana*<sup>49</sup>. Two methyl esters (76) and (77) of 3,4- disubstituted benzoic acid have been detected in *Trichocolea tomentella*<sup>50</sup> together with 3,4- dihydroxy benzoic acid (78).



(74)



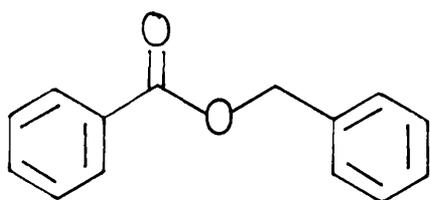
(75)



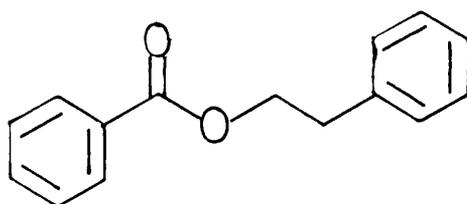
(76)  $R = \text{Me}, R' = \text{H}$

(77)  $R = R' = \text{Me}$

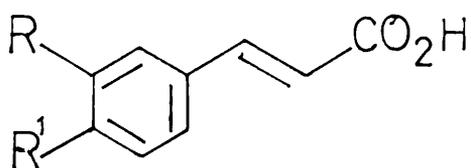
(78)  $R = R' = \text{H}$



(79)

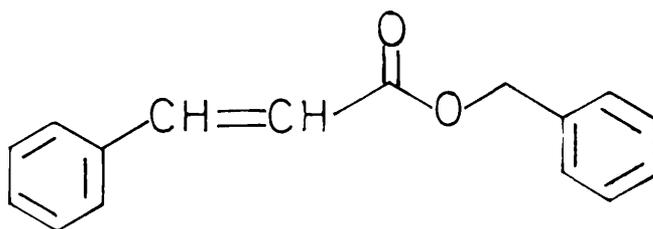


(80)



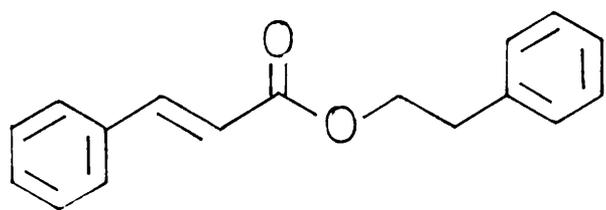
(81)  $R = \text{H}, R' = \text{OH}$

(82)  $R = \text{OH}, R' = \text{H}$

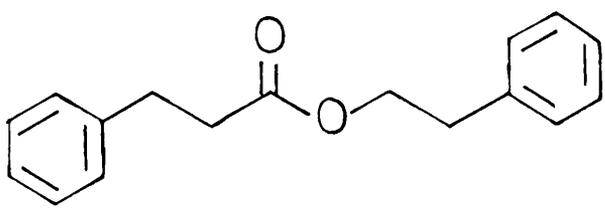


(83) Cis isomer

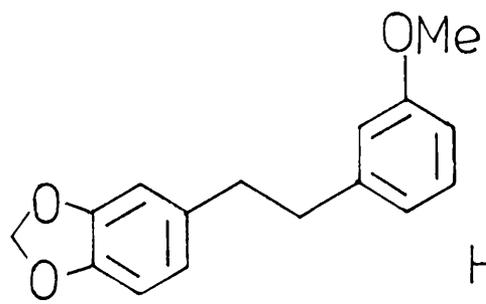
(84) Trans isomer



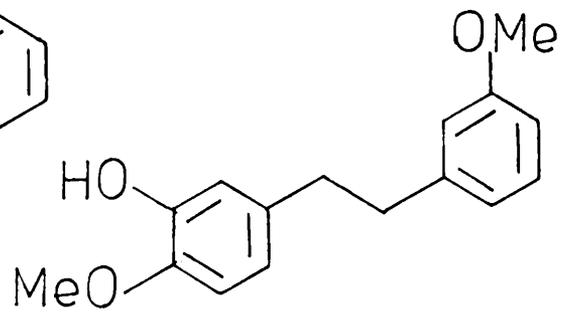
(85)



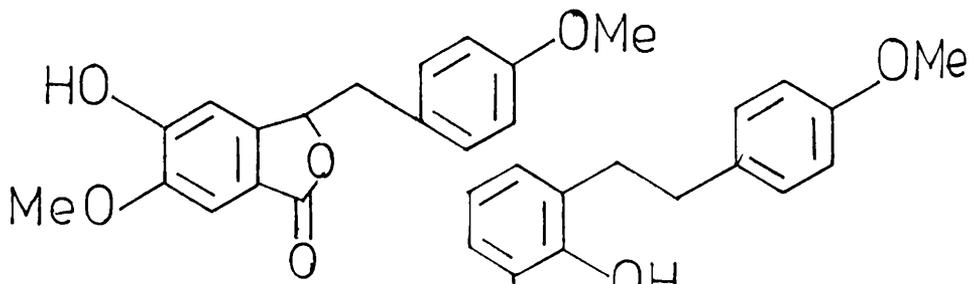
(86)



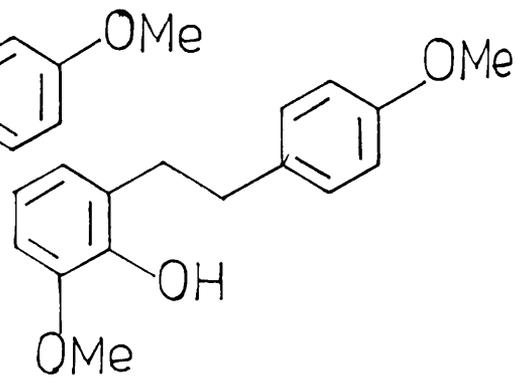
(87)



(88)



(89)



(90)

Benzyl benzoate (79) and  $\beta$ -phenylethylbenzoate (80) have been isolated from the primitive liverwort Isotachis japonica<sup>23</sup>.

#### CINNAMIC ACID DERIVATIVES

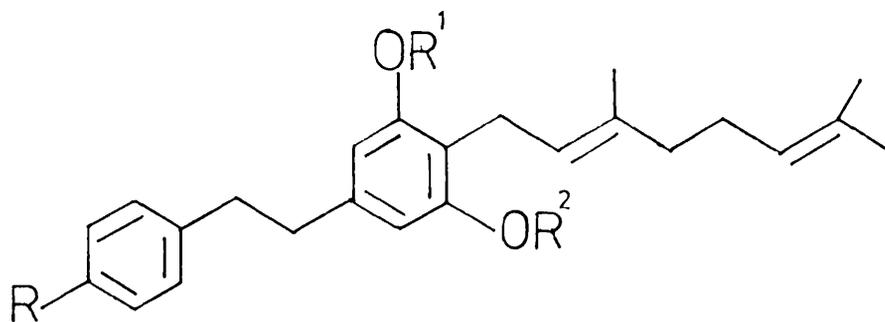
Asterella species<sup>49</sup> contain p- and m- coumaric acids (81), (82) whilst Isotachis japonica<sup>51</sup> contains benzyl trans- and cis- cinnamates (83), (84) together with  $\beta$ - phenylethyl- cinnamate (85) and its dihydro derivative (86).

#### BIBENZYL

Frullania species are rich sources not only of sesquiterpene lactones but also of bibenzyl derivatives. Twenty-five Frullania species have been studied<sup>52</sup> and twelve species contain bibenzyl derivatives. Recently, Asakawa et al<sup>53</sup>, have reported the isolation of three novel bibenzyl derivatives (87) - (89) from the Australian Frullania falciloba. Their structures were established as 3,4- methylene-dioxy- 3'- methoxybibenzyl (87), 3- hydroxy- 4,3'- dimethoxybibenzyl (88) and 3- [4'- methoxybenzyl]- 5,6- dimethoxyphthalide (89) by spectral methods and synthesis.

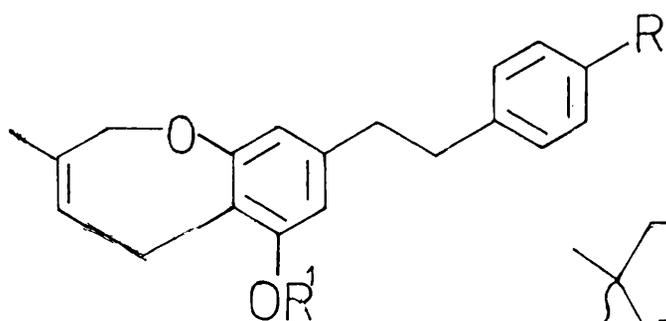
Pellepiphyllin (90) has been isolated from Pellia epiphylla<sup>54</sup>.

Radula species contain many bibenzyls incorporating prenyl groups. Seven bibenzyls (91) - (97) have been isolated from R. variabilis. Their structures were established mainly by detailed analysis of <sup>1</sup>H n.m.r. and mass spectra<sup>55</sup>.



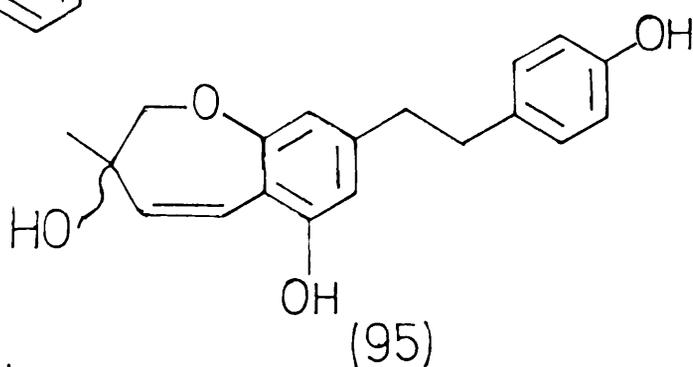
(91)  $R = R^1 = R^2 = H$

(92)  $R = OH, R^1 = R^2 = H$

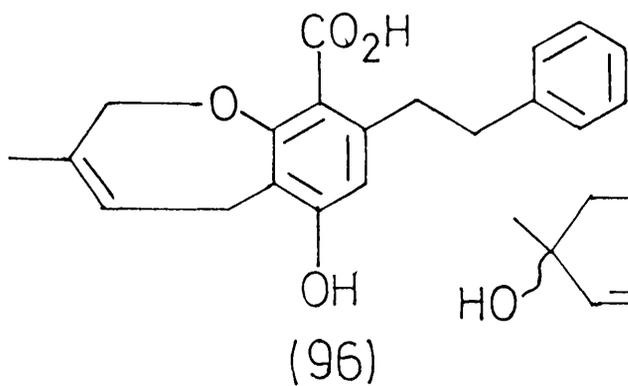


(93)  $R = R^1 = H$

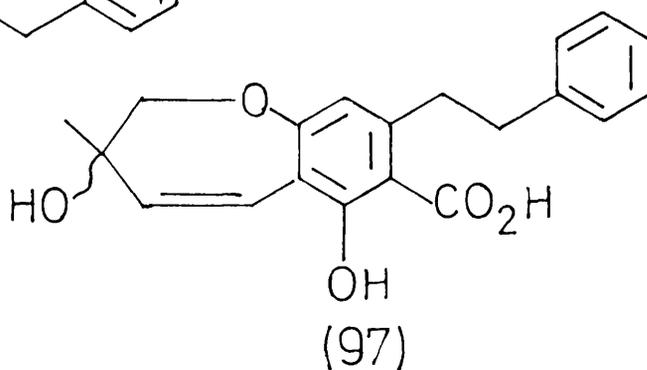
(94)  $R = OH, R^1 = H$



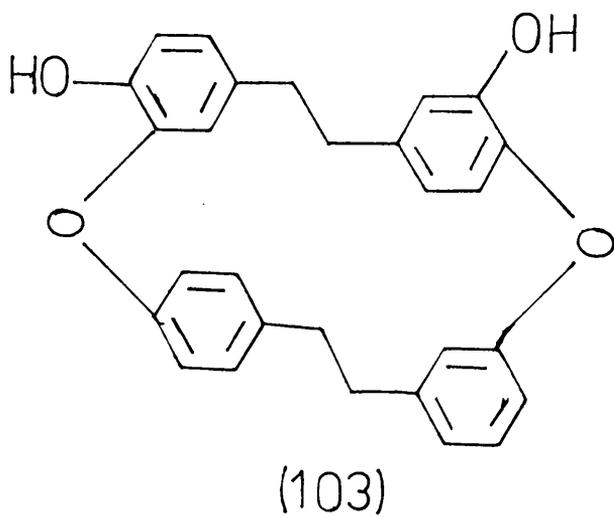
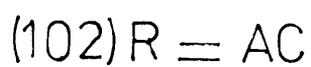
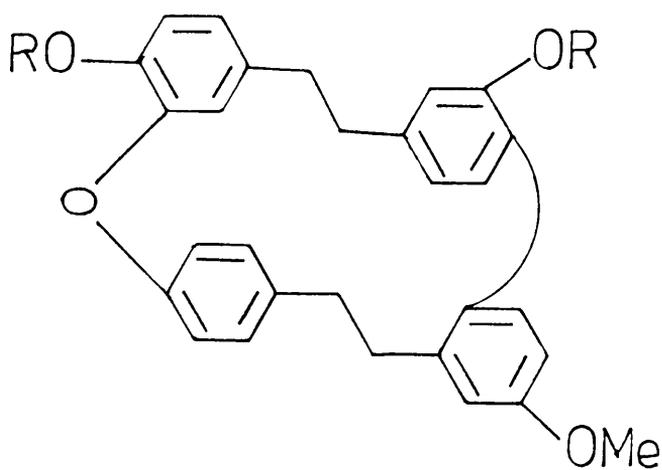
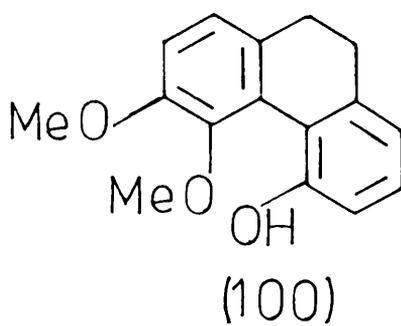
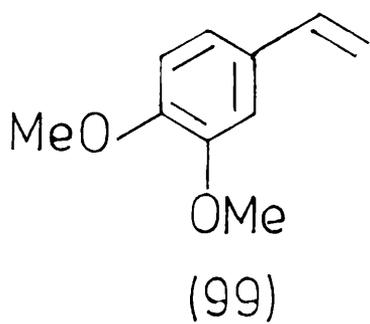
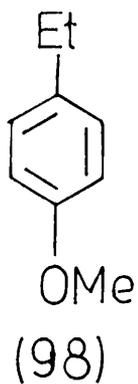
(95)



(96)



(97)

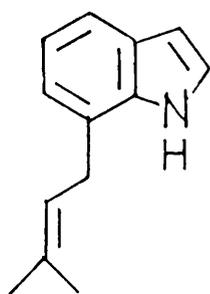


The p- ethylanisole (98) isolated from Loptolejeunes elliptica<sup>56</sup> causes an intense mold-like odour. Conocephalum conicum<sup>57</sup> produces 1- vinyl- 3,4- dimethoxybenzene (99) as a minor component. The 3,4- dimethoxy- 5- hydroxy- 9, 10- dihydrophenanthrene (100) was isolated from Riccardia jackii and its structure established by x-ray crystallographic analysis<sup>58</sup>. The novel aromatic ethers riccardin A (101) and riccardin B (103) have been isolated from Riccardia multifida. The structure of the former was established by x-ray analysis of the diacetate (102)<sup>59</sup>.

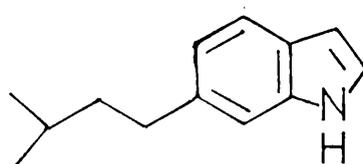
#### INDOLE DERIVATIVES

The unusual indole derivatives (104), (105) have been isolated from Riccardia sinuata<sup>56,57</sup>. These are the first nitrogen-containing compounds in the Hepaticae.

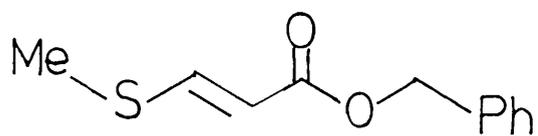
Two new sulphur-containing acrylates, isotachin A (106) and isotachin B (107) were detected in the liverwort Isotachis japonica<sup>60</sup>. Their structures were determined by chemical and spectral methods.



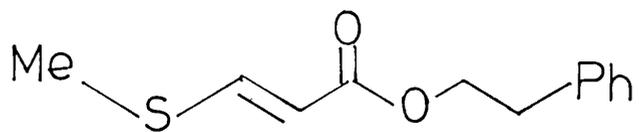
(104)



(105)



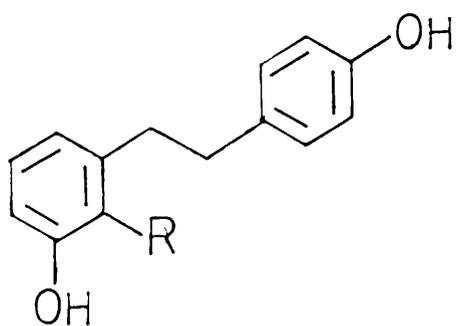
(106)



(107)

C H A P T E R 2

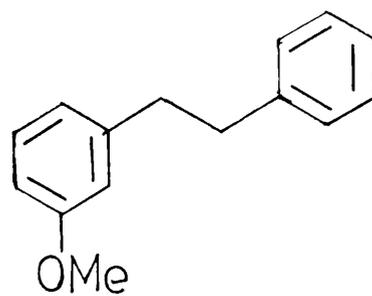
---



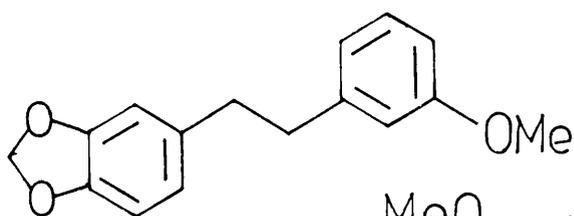
(108)  $R = CO_2H$

(109)  $R = H$

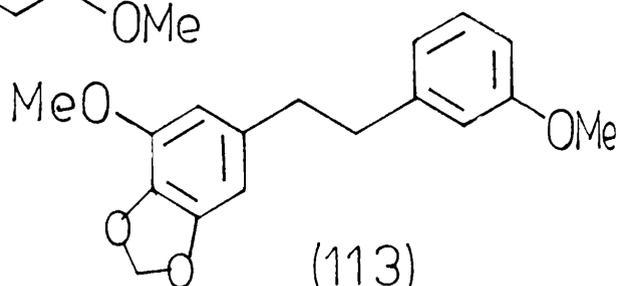
(111)  $R = OH$



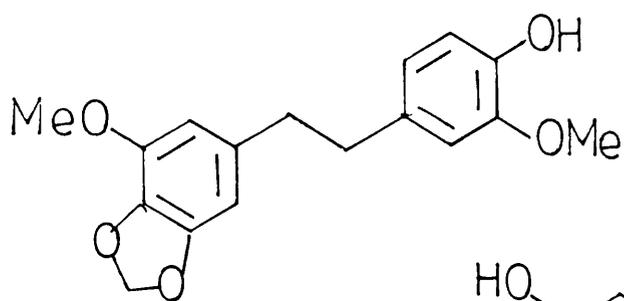
(110)



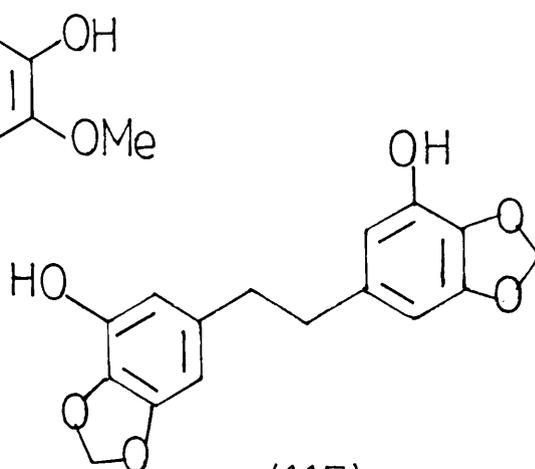
(112)



(113)



(114)



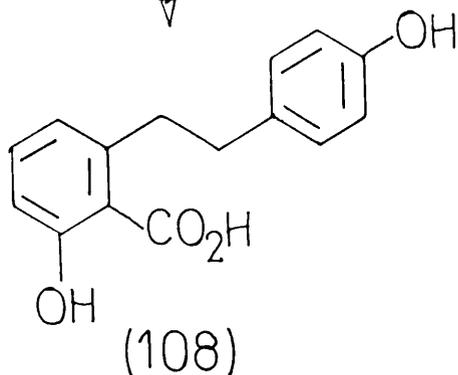
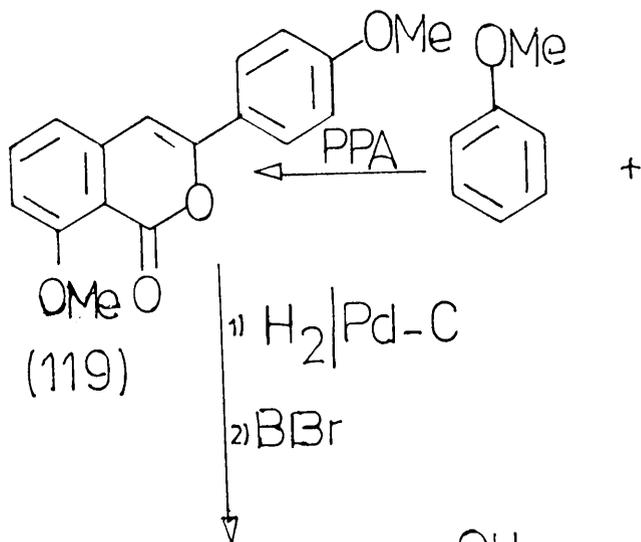
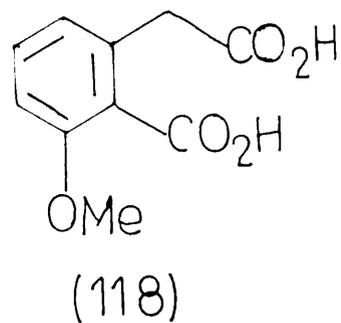
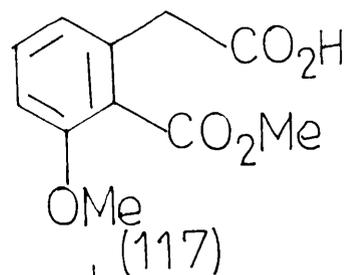
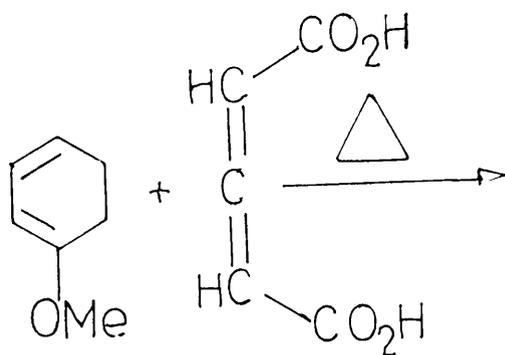
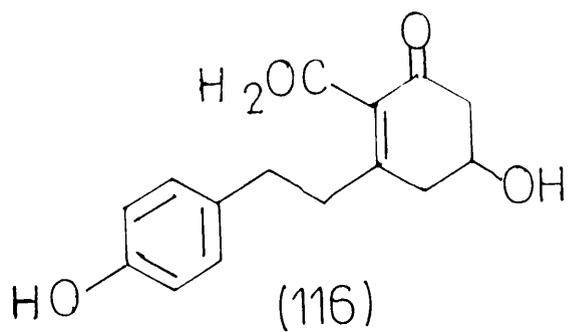
(115)

## INTRODUCTION

Lunularic acid (3,4'- dihydroxybibenzyl-2- carboxylic acid) (108) was first isolated by Valio et al<sup>61</sup>, in 1960 as a growth inhibitor and dormancy factor in the Middle East strain of the liverwort Lunularia cruciata. It has since been found in numerous other liverworts. Compound (108) was tested in a number of higher plant systems by Fries<sup>62</sup> for plant growth inhibitory activities. It was observed that it had an effect at higher concentration, but no effect at low concentration. Lunularic acid (108) and the corresponding decarboxylated derivative lunularin (109) have been detected<sup>63</sup> in 76 species of Hepaticae, but do not occur in the Anthocerotae or algae.

In addition to compound (108) numerous further bibenzyls have been isolated from liverwort species. 3- Methoxybibenzyl (110)<sup>64</sup> was isolated from Frullania dilata var anamola while 2,4,4'- trihydroxybibenzyl (111)<sup>65</sup> was isolated from Pellia endiviifolia and its structure proved by synthesis<sup>66</sup>. Later Asakawa<sup>67</sup> found four new related bibenzyls (112) - (115) from three Frullania species. Compounds (112) and (113) were isolated from F. ericoides. The former compound has been found also in Australian F. falciloba and its structure confirmed by synthesis<sup>53</sup>. Compound (113) has also been synthesised<sup>67</sup>. The bibenzyl (114) was obtained from F. bonincola while compound (115) was isolated from F. paruistipula.

The most significant compound in this series, prelunularic acid (preLNA)(116)<sup>68</sup> was isolated from the suspension

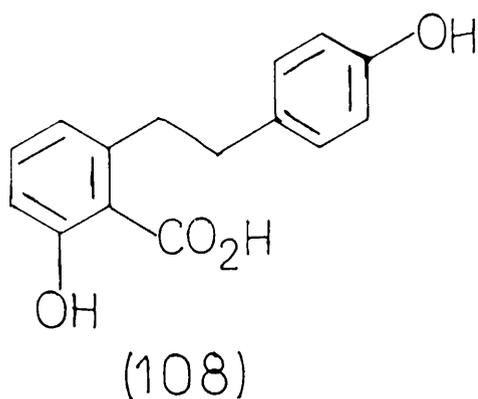
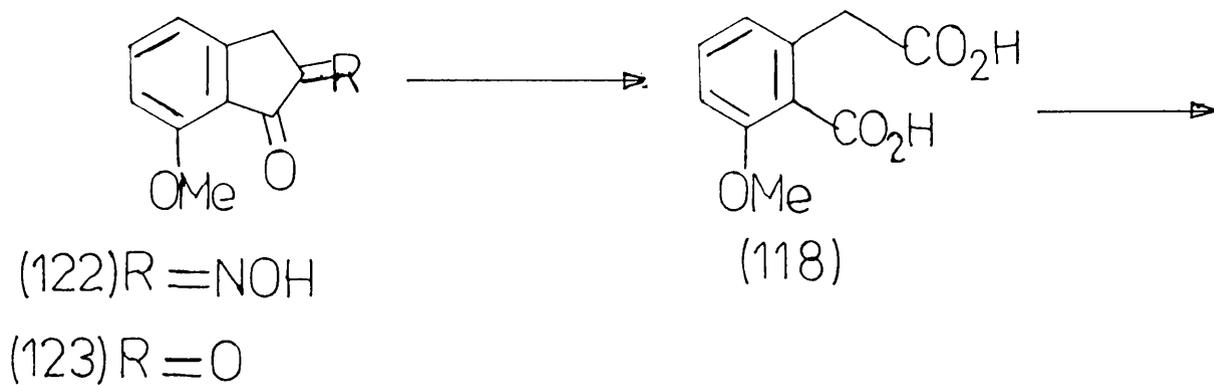
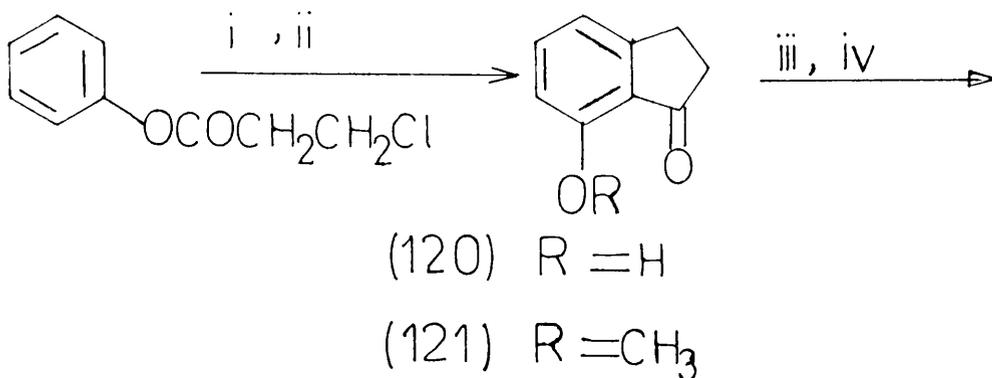


cultured cells of Marchantia polymorpha. Its structure was determined by spectroscopic and chemical means.

Prelunularic acid (116) has been shown to be converted into lunularic acid (108) under acidic or basic conditions. For example approximately 0.3% of preLNA (116) in water was converted in one day at room temperature, and 9% on boiling for 30 minutes. The isolation of (116) is significant for the biosynthesis of (108) and stilbenoids in general.

Two syntheses of lunularic acid have been reported. The first was by Arai et al<sup>69</sup>. Heating a mixture of 1-methoxy cyclo- hexa- 1,3- diene with allene- 1,3- dicarboxylate in a sealed tube gave dimethyl 3- methoxyhomophthalate (117). The hydrolysis of (117) gave 3- methoxyhomophthalic acid (118). Condensation <sup>of</sup>(118) with anisole in polyphosphoric acid gave the isocoumarin (119) in 81% yield. Catalytic hydrogenation of (119) followed by treatment with BBr<sub>3</sub> afforded (108). Its <sup>1</sup>H n.m.r. was identical with that of the natural acid (scheme 5).

In 1977 Huneck and Schreiber<sup>70</sup> devised another synthesis of (108)(scheme 6). Phenyl β-chloropropionate was converted into the corresponding hydrindanone (120) by Friedel-Crafts cyclisation. Methylation with methyl iodide in acetone and K<sub>2</sub>CO<sub>3</sub> afforded (121) which was converted into the oximoketone (122) on treatment with isoamyl nitrite. Compound (122) in 30% formaldehyde-water yielded the diketone (123)



(i)  $AlCl_3$ ; (ii)  $MeI$ ; (iii) isoamyl nitrite; (iv)  $HCHO/HCl$

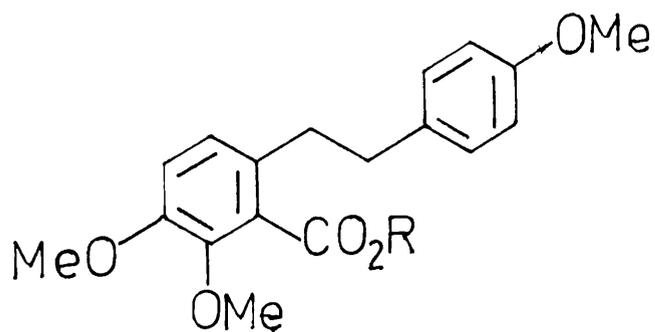


which was transformed into the homophthalic acid (118) by 15%  $H_2O_2$  in glacial acetic acid. The conversion of the homophthalic acid (118) into lunularic acid followed the same route as in (scheme 5).

The biosynthesis of the various aromatic compounds that have been recently isolated from the Hepaticae and Anthocerotae is not always clearly established. Stilbenes, bibenzyls and phenanthrenes are believed to be derived from shikimic acid and acetate units. The biosynthesis of lunularic acid (108) has been investigated and the results show that this bibenzyl derivative (108) is synthesised in vivo<sup>71</sup> via the phenyl propanoid-polymalonate pathway. Reaction of pyruvate (124) with shikimic acid (125) followed by a process not unrelated to the Claisen rearrangement leads to prephenate (126). This is a biological example a symmetry-allowed pericyclic process. Decarboxylation of (126) is followed by aromatisation and reductive transamination to give phenylalanine (127). The stereospecific elimination of ammonia yields trans-p-coumaric acid (128). Condensation of p-coumaroyl-Co-A with a triketide unit, then cyclisation and aromatisation of the polyketide chain followed by the reduction of the olefinic protons in (129) affords lunularic acid (108) (Scheme 7). The main metabolite of lunularic acid in Lunularia cruciata is its decarboxylation product<sup>72</sup> lunularin (109).

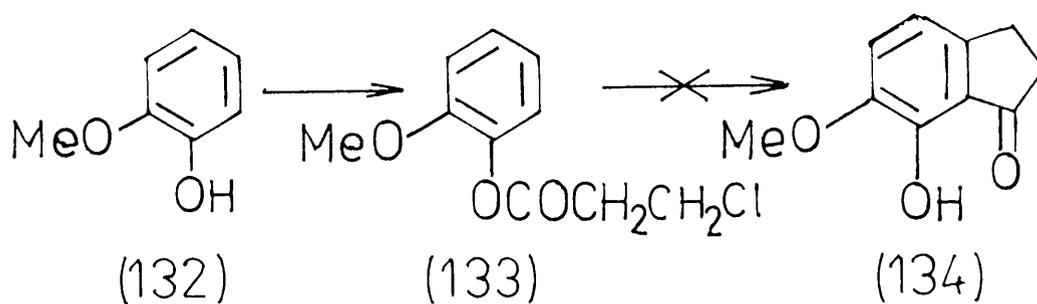
The novel bibenzyl (130) was obtained from Plagiochila spinulosa by Connolly<sup>73</sup>. It is clearly a derivative of lunularic acid. Its structure was determined on the basis of its spectroscopic properties. Its <sup>1</sup>H n.m.r. showed an AA'BB' quartet [ $\delta_{\text{H}}$  7.08(d, J8.8Hz) and 6.83(d, J8.8Hz)] and an AB quartet [6.84(d, J8.5Hz) and 6.81(d, J8.5Hz)] and benzylic methylene protons at  $\delta_{\text{H}}$  2.76. Accurate mass measurement confirmed its molecular formula as C<sub>19</sub>H<sub>22</sub>O<sub>5</sub> [m/z 330.1478] and gave useful fragments at m/z 209 (C<sub>11</sub>H<sub>13</sub>O<sub>4</sub><sup>+</sup>) and the base peak at 121 (C<sub>8</sub>H<sub>9</sub>O<sup>+</sup>).

This chapter deals with the <sup>attempted</sup> synthesis of the lunularic acid derivative (131) to enable further evaluation of its biological activity.

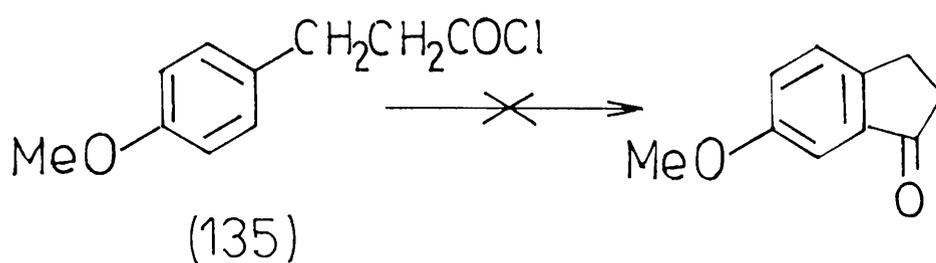


(130) R = Me

(131) R = H



Scheme 8



Scheme 9

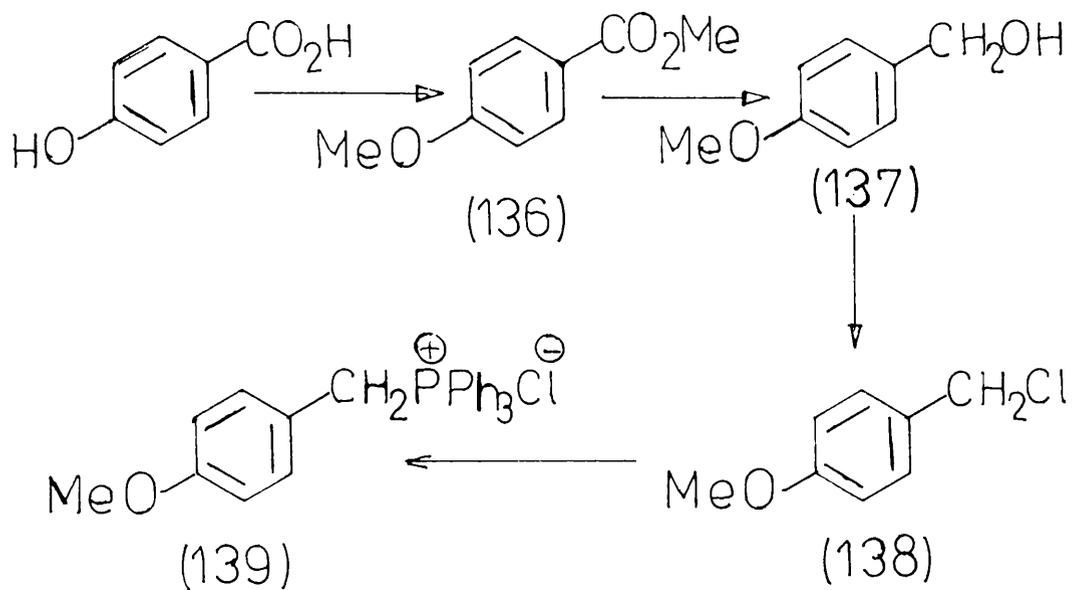
## DISCUSSION

Our aim was to obtain by synthesis 3,4,4'-trimethoxy-  
*bibenzyl*-2-carboxylic acid (131).

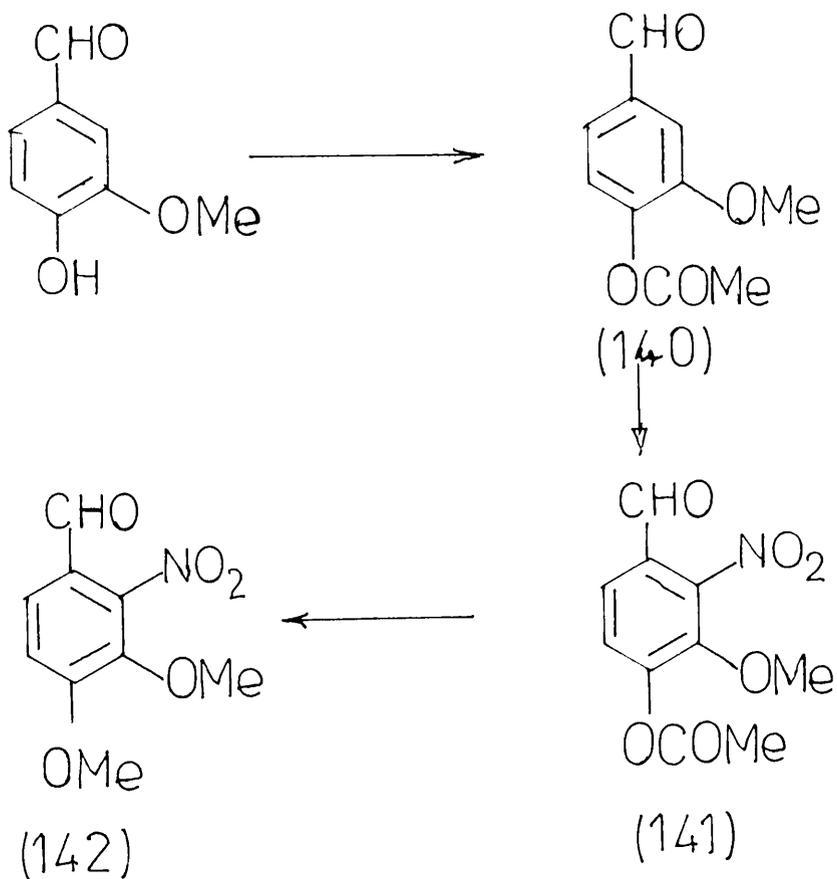
First the Huneck<sup>70</sup> approach was attempted. Reaction of guaiacol (132) with  $\beta$ -chloropropionyl chloride in presence of  $\text{POCl}_3$  afforded the chloroester (133) in good yield. Unfortunately Friedel-Crafts cyclisation of (133) to give the desired hydrindanone (134) failed despite several efforts - (scheme 8). Under similar conditions 4-methoxyphenyl-propionyl chloride (135) also failed to cyclise, (scheme 9). This approach was therefore abandoned.

The next approach we considered was construction of the bibenzyl system using a Wittig reaction. We intended introducing the carboxyl group via a cyano derivative. Thus we prepared 4-methoxybenzyl <sup>*triphenyl*</sup>phosphonium chloride (139), (scheme 10), and 2-nitroveratraldehyde (142), (scheme 11) as appropriate precursors.

The esterification of 4-methoxybenzoic acid was carried out using excess dimethyl sulphate in refluxing acetone and potassium carbonate for 5 hours. The  $^1\text{H}$  n.m.r. spectrum of the methoxy methyl ester (136) showed an AA' BB' quartet for four aromatic protons at  $\delta_{\text{H}}$  7.58 and 6.55 (J8.8Hz.), in addition to two methyl signals at  $\delta_{\text{H}}$  4.02 (OMe) and 3.95 ( $\text{CO}_2\text{Me}$ ).



Scheme 10



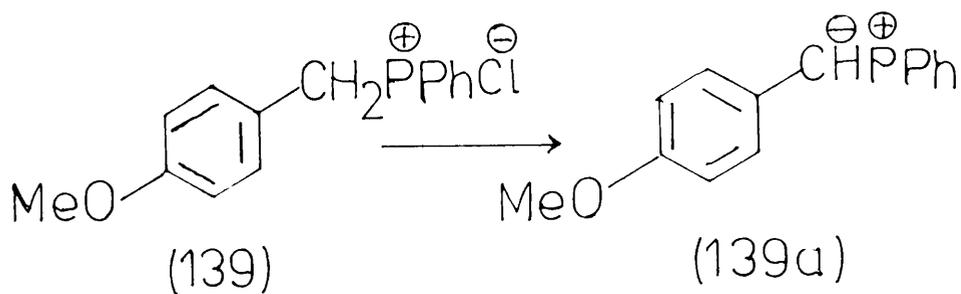
Scheme 11

Methyl 4-methoxybenzoate (136) was easily reduced to the corresponding alcohol (137) using lithium aluminium hydride in dry ether. The reduction was vigorous and the lithium aluminium hydride was added in small portions. The excess of lithium aluminium hydride was conveniently destroyed by addition of aqueous sodium sulphate. The  $^1\text{H}$  n.m.r. spectrum of the product (137) ( $\nu_{\text{max}}$  3390  $\text{cm}^{-1}$ ) showed loss of the methyl ester and the appearance of a two proton singlet at  $\delta_{\text{H}}$  4.45 due to the benzylic methylene group.

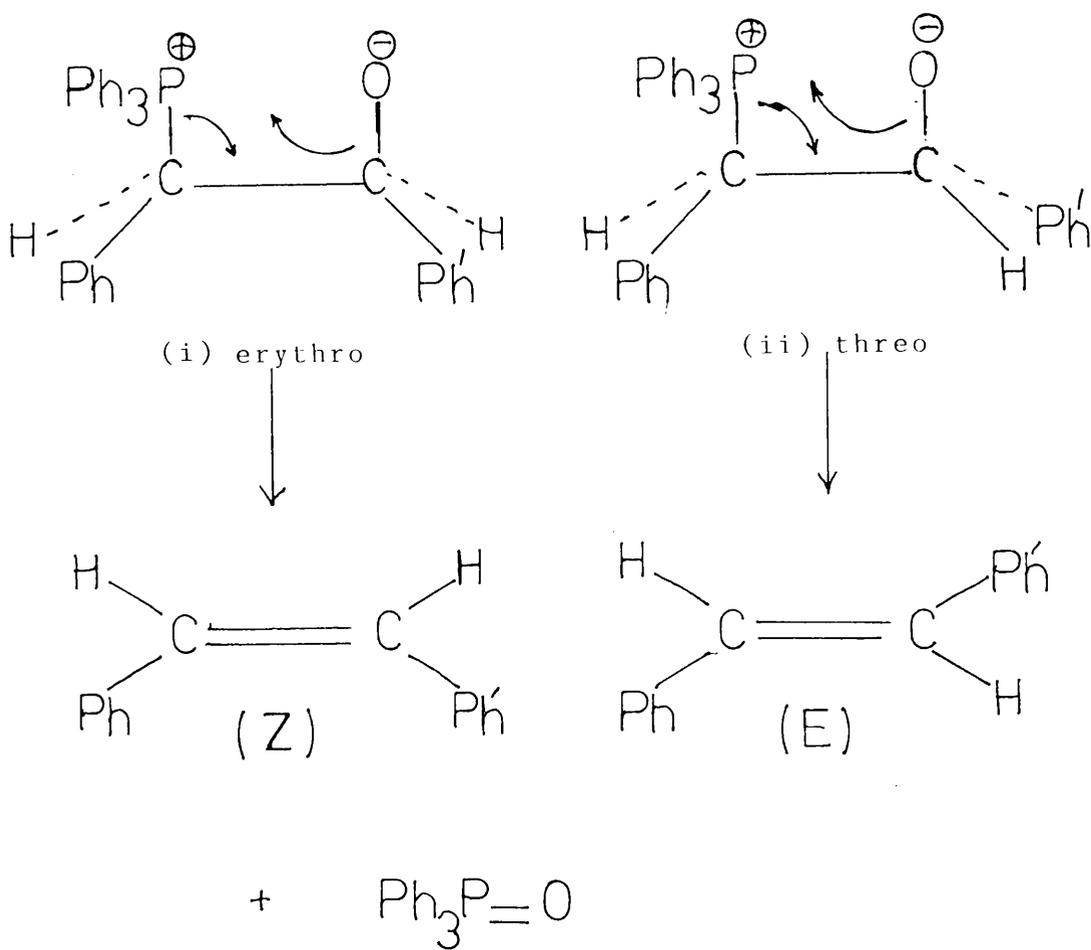
The benzyl chloride (138) was transformed into the Wittig reagent (139) by treatment with triphenyl phosphine under anhydrous conditions in toluene. The  $^1\text{H}$  n.m.r. spectrum of (139) showed the expected coupling of the benzylic methylene protons with phosphorous [ $\delta_{\text{H}}$  5.32(d,  $J_{\text{PH}}$  14Hz.)].

The preparation of 2-nitrovanillin (142) is described in scheme 11. Acetylation of vanillin with acetic anhydride in dry pyridine and stirring overnight led after crystallisation from water to vanillin acetate (140). [ $\nu_{\text{max}}$  ( $\text{CCl}_4$ ) 3020, 1760, 1700 and 1600  $\text{cm}^{-1}$ ]. Its  $^1\text{H}$  n.m.r. spectrum showed the loss of the hydroxyl group and the appearance of an acetate at  $\delta_{\text{H}}$  2.35(3H, S).

Vanillin acetate (140) was nitrated at  $-5^\circ$   $-0^\circ$  by refluxing with nitric acid (d. 1.50) with vigorous stirring. The yield is dependent on the strength of the acid (d. 1.50) and its age. It was found necessary to use carefully dried vanillin acetate for the nitration, otherwise it was difficult to remove all odour of nitric acid from the product.



Scheme 12

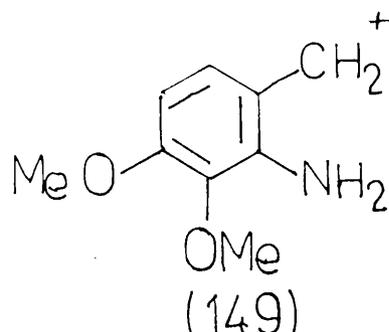
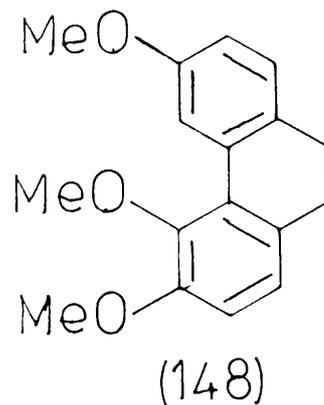
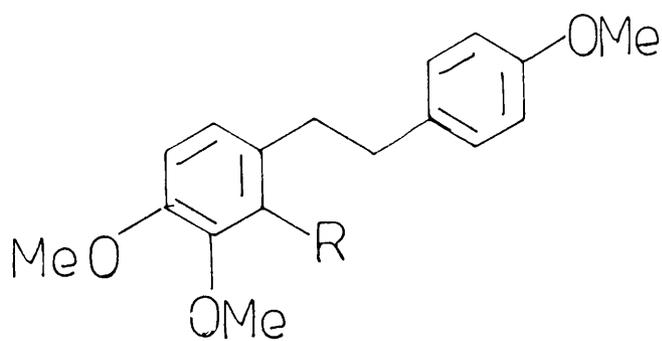
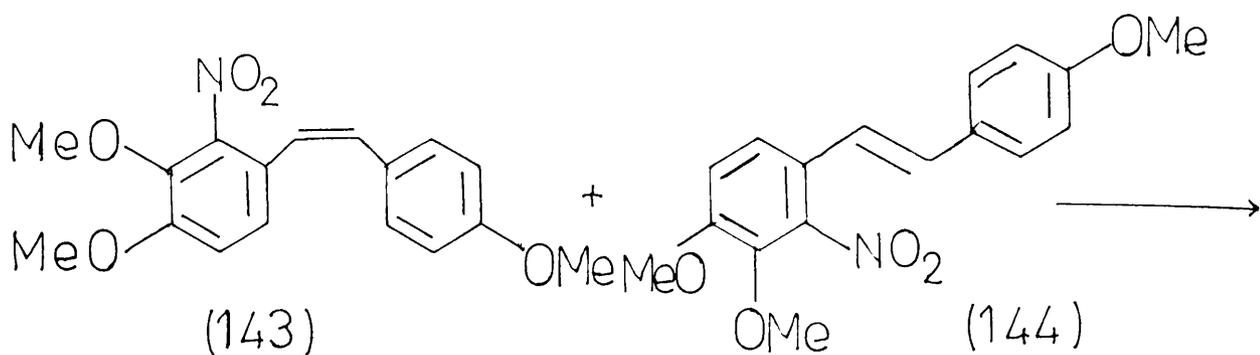
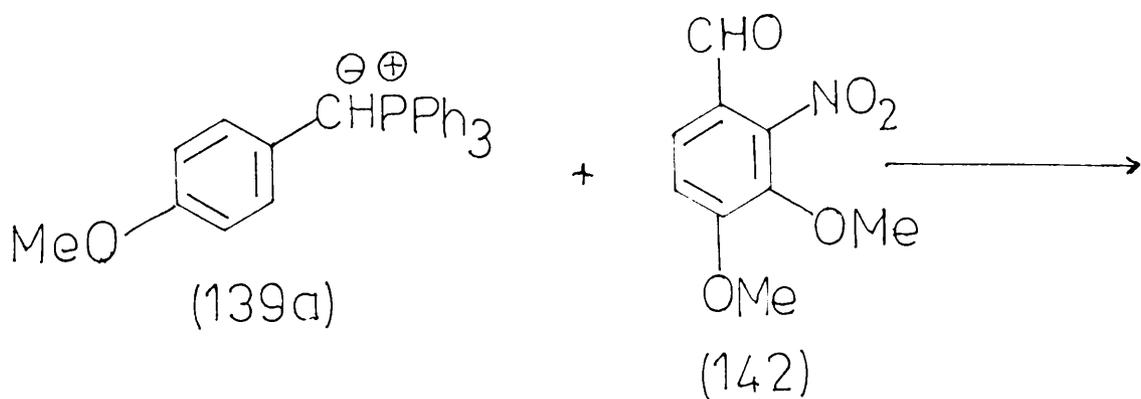


Scheme 13

Nitration of vanillin acetate (140) under these conditions gives mainly the 2- nitro derivative (141). A small portion of the 6- nitro derivative can be isolated from the mother liquor. The structure of the 2- nitro compound (141) was readily assigned on the basis of its  $^1\text{H}$  n.m.r. which shows two ortho- coupled aromatic protons as an AB quartet at  $\delta_{\text{H}}$  7.85 and 7.58 (J9.8Hz) together with the expected signals for the aldehyde at  $\delta_{\text{H}}$  10.15, the methoxy group at  $\delta_{\text{H}}$  3.95 and the acetate methyl at  $\delta_{\text{H}}$  2.32. Mass spectrometry confirmed its molecular formula ( $\text{C}_{10}\text{H}_9\text{NO}_6$ ; m/ 239).

The acetate (141) was converted to 2- nitrovanilaldehyde (142) following the method of MacDonald<sup>74</sup>, using dimethyl sulphate in the presence of strong base (50% sodium hydroxide). It was found necessary to use an excess of dimethyl sulphate and to maintain the pH at about 11. Ammonia was added to destroy the excess of the reagent. On cooling the solution to 0°C the product (142) precipitated. Its  $^1\text{H}$  n.m.r. spectrum confirmed the presence of two methoxy groups at  $\delta_{\text{H}}$  4.00 and 3.92, the aldehyde singlet at  $\delta_{\text{H}}$  9.77 and an AB quartet at  $\delta_{\text{H}}$  7.64 and 7.12 (J8.7Hz) for the aromatic protons.

The first step of the Wittig reaction involves the formation of a carbanion by deprotonation of the phosphonium chloride (139), (scheme 12). The ylid (139a) is a good nucleophile and should react easily with the aldehyde. There are two possible transition states: (i) erythro, and (ii) threo, (scheme 13), which lead to two isomeric products (143) and (144) as shown in scheme 13.



Scheme 14

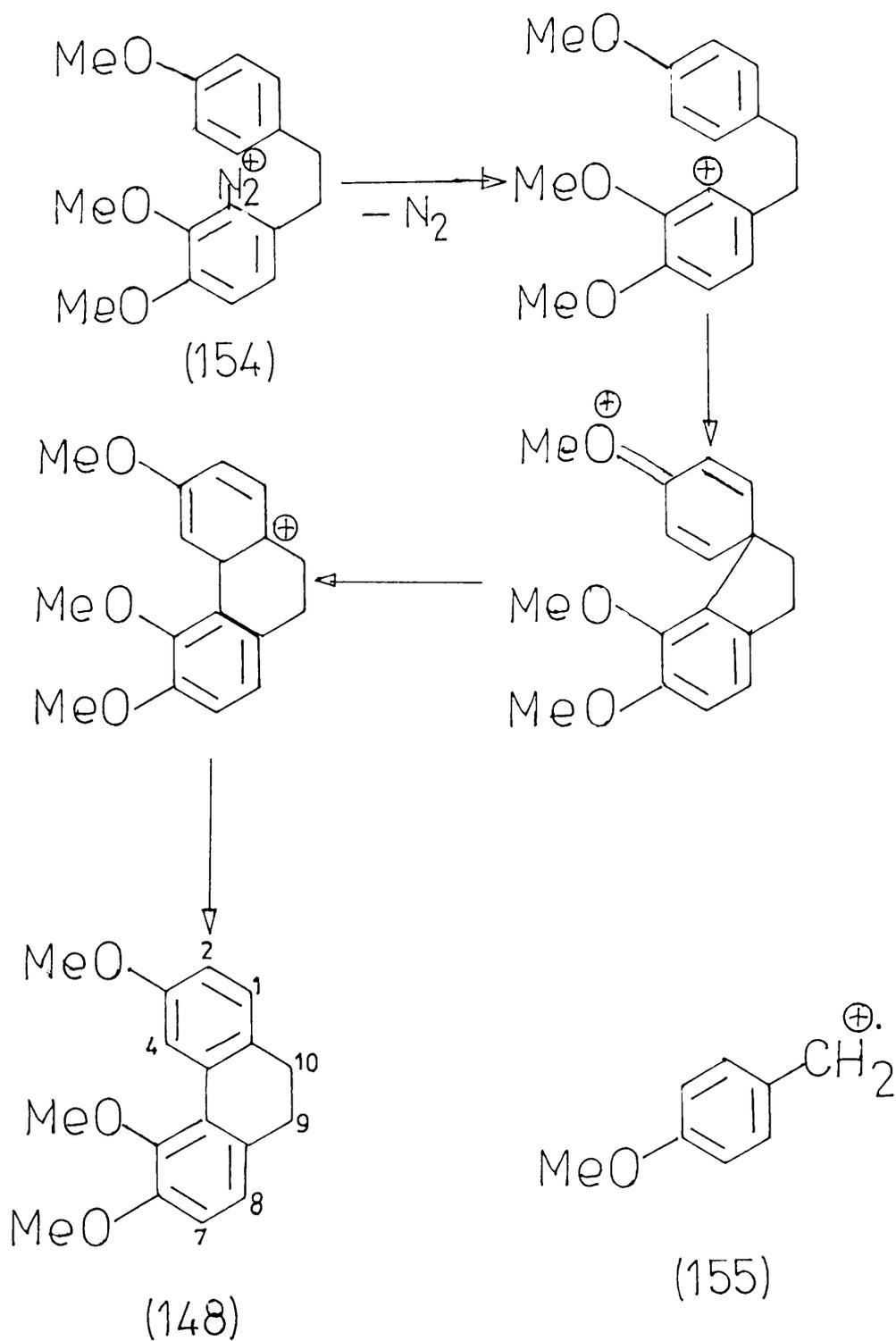
On addition of butyl lithium to the phosphonium chloride (139) a deep-red colour appeared, presumably indicating the formation of the ylid (139a). Reaction of the aldehyde (142) with the ylid (139a) gave the desired stilbene as a mixture of Z- (143) and E- (144) forms, the <sup>former</sup> predominating since studies<sup>75</sup> with non-stabilised ylides have shown that (Z-) alkene normally predominated. In the <sup>1</sup>H n.m.r. spectrum of the mixture the olefinic protons of the Z-isomer appeared as an AB quartet at  $\delta_{\text{H}}$  6.60 and 6.25 (J<sub>11</sub> 9.5 Hz). Its aromatic protons appeared at  $\delta_{\text{H}}$  7.09 (d, J<sub>8</sub> 4 Hz., H<sub>2,6</sub>), 6.94 (d, J<sub>8</sub> 6 Hz., H<sub>3,5</sub>); 6.79 (d, J<sub>8</sub> 7 Hz., H<sub>5</sub>), and 6.72 (d, J<sub>8</sub> 7 Hz., H<sub>6</sub>), and its three methoxy groups at  $\delta_{\text{H}}$  3.93, 3.86 and 3.75 (each 3H, s). The high resolution mass spectrum of the 2-nitrostilbene mixture confirmed the molecular formula C<sub>17</sub>H<sub>17</sub>NO<sub>5</sub> (m/z 315.2352). The <sup>1</sup>H n.m.r. spectrum of the stilbene mixture showed the formation of the (143) and (144) in a ratio of 3:1. The E-isomer appears similar to the Z-form except the large coupling constant of 16 Hz. of the olefinic protons.

The nitrostilbene mixture was converted to the corresponding bibenzyl (145), by hydrogenation in methanol over 5% Pd-C. Filtration of the catalyst and removal of solvent in vacuo afforded the 2-nitrobibenzyl (145) in high overall yield. The structure accorded with its <sup>1</sup>H n.m.r. spectrum which shows the loss of olefinic protons and the appearance of a broad multiplet at  $\delta_{\text{H}}$  2.77 arising from the four benzylic methylene protons.

The aromatic protons of (145) were observed as an AB quartet at  $\delta_{\text{H}}$  7.04 and 6.92 (J 8.7 Hz.) and an AA'BB' system at  $\delta_{\text{H}}$  6.8 and 6.83 (J 8.9 Hz.). The remaining signals of the three methoxy groups appeared at  $\delta_{\text{H}}$  3.92, 3.87 and 3.77. (145) showed a molecular ion at  $m/z$  317.2681 in the high resolution mass spectrum ( $\text{C}_{17}\text{H}_{19}\text{O}_5$  requires 317.1263)

The next step involved reduction of the nitrobibenzyl (145) to the corresponding amino compound (146). Attempts to achieve this by prolonged hydrogenation in EtOAc over 5% Pd/C were unsuccessful. Lithium aluminium hydride proved to be a much more convenient reagent for the reduction, which readily afforded the aminobibenzyl (146) as a crystalline solid. Its i.r. spectrum showed the presence of primary amino bands at  $\bar{\nu}_{\text{max}}$  3450  $\text{cm}^{-1}$ . Its  $^1\text{H}$  n.m.r. spectrum revealed the characteristic AA'BB' [ $\delta_{\text{H}}$  7.08(d, J 8.7 Hz.); 6.81(d, J 8.7 Hz.)] and AB [ $\delta_{\text{H}}$  6.78(d, J 8.6 Hz.), 6.35(d, J 8.5 Hz.)] systems for the aromatic protons, three methoxy groups at  $\delta_{\text{H}}$  3.81, 3.80 and 3.76; and a broad multiplet at  $\delta_{\text{H}}$  2.81 due to the benzylic methylenes. The signal for the amino protons appeared as a singlet (exchangeable with  $\text{D}_2\text{O}$ ) at  $\delta_{\text{H}}$  2.16. Accurate mass measurement of (146) confirmed its molecular formula as  $\text{C}_{17}\text{H}_{21}\text{NO}_3$  ( $m/z$  287.1521). The base peak  $m/z$  166 is due the fragment (149).

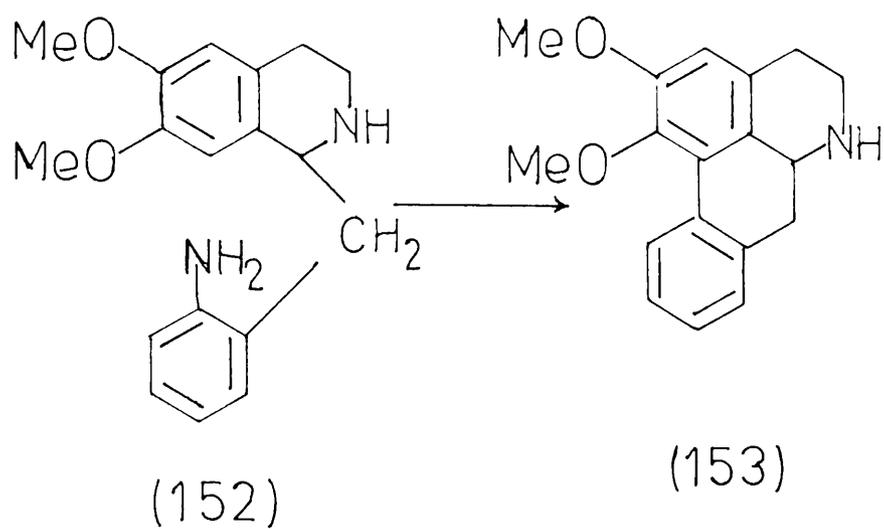
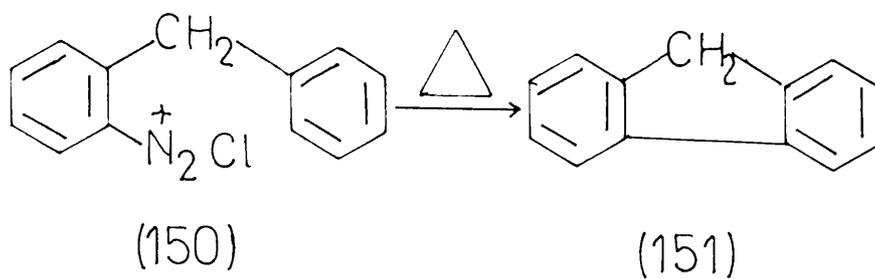
The amino compound (146) was subjected to typical Sandemeyer conditions in the hope of obtaining the nitrile (147). Thus a solution of the diazonium salt of (146) and potassium cyanide was added to a hot solution of CuCN at 85°C.



Scheme 15

Unfortunately the major product proved to be the dihydrophenanthrene (148), the result of a Pschorr ring closure. Intramolecular cyclisation of diazonium salts, best known as the Pschorr cyclisation, dates back to the work of Pschorr<sup>76</sup> in 1896. In fact the first example of this type of reaction (150) - (151) was reported by Fischer and Schmidt<sup>(77)</sup> in 1894. The reaction has found more modern application in a synthesis of aporphine<sup>78</sup> alkaloids. The alkaloid (153) was formed in good yield by diazotising the amine (152) in presence of copper salts. It has been reported that in the presence of copper salts<sup>79</sup> diazonium salts of cis-stilbene give phenanthrenes rather than nitriles in 60-80% yield.

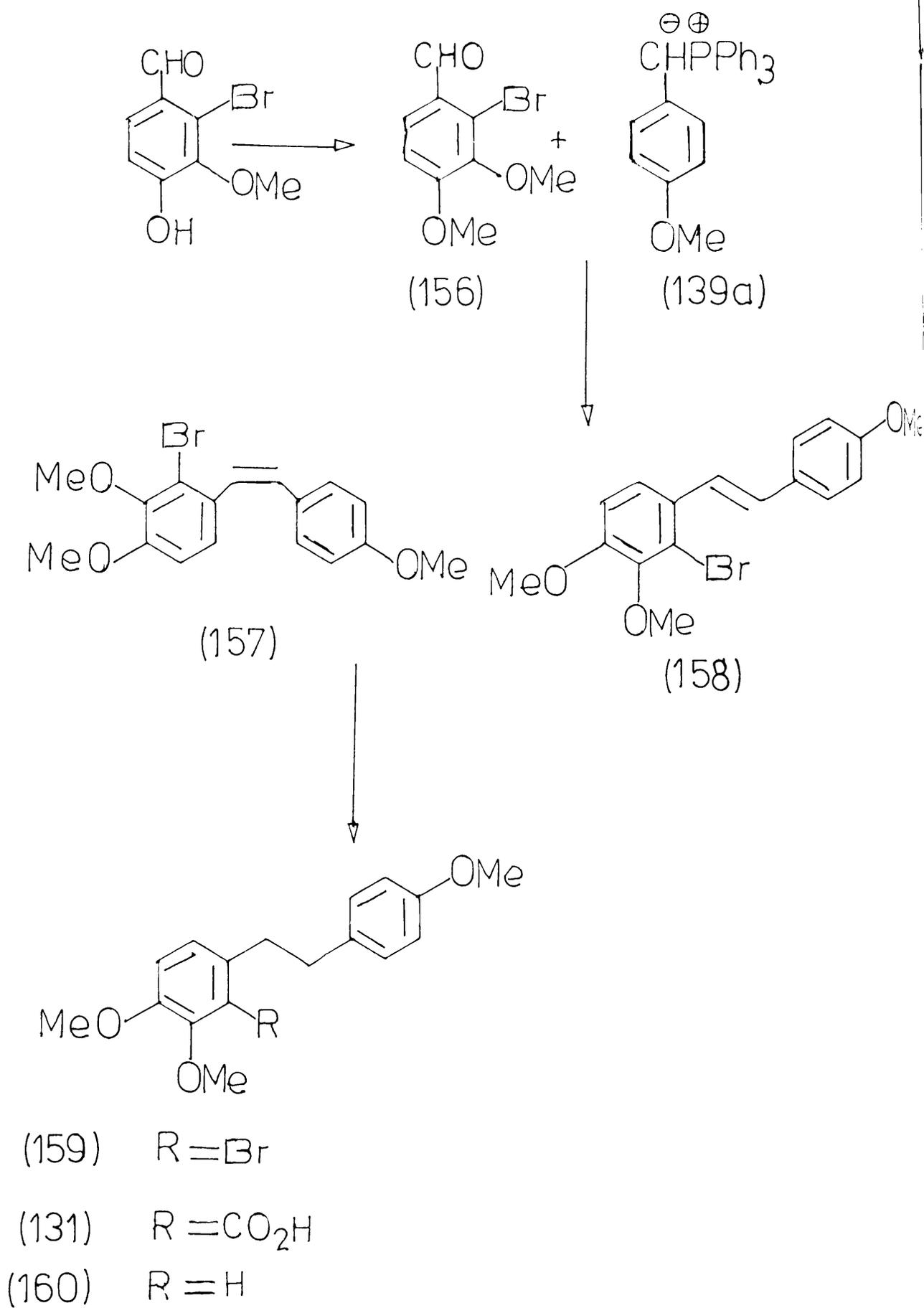
Only one possible dihydrophenanthrene product can arise from the diazonium salt (154) since the positions 2'- and 6'- are similar. A suggested mechanism is drawn in (scheme 15). Attack of the electrophile on C-3 or C-5 of the top ring i.e. meta to the methoxy group is unfavourable in electronic terms. Initial ipso attack followed by migration and aromatisation may be a more likely mechanism. The structure of the dihydrophenanthrene (148) was assigned on the basis of its <sup>1</sup>H n.m.r. The structure (148) has five aromatic protons [ $\delta_{\text{H}}$  8.12(d, J2.7 Hz.); 7.18(d, J8.2Hz.), 6.96(d, J8.2Hz.), 6.82(dd, J2.7, 8.2Hz) and 6.78(d, J8.2 Hz)]. The doublet at  $\delta_{\text{H}}$  8.12 is characteristic of a proton at C-4 in this system. It has a meta coupling (J2.72Hz) to the signal at  $\delta_{\text{H}}$  6.82 and the latter has a further ortho coupling (J2.7, 8.2Hz) to the proton of C-1 at



$\delta_{\text{H}}$  6.96(d, J8.2Hz.). There are also three methoxy groups [ $\delta_{\text{H}}$  3.90, 3.84, and 3.73 each (3H, s)] and an  $A_2B_2$  system [ $\delta_{\text{H}}$  2.73 (4H, m)]. Accurate mass measurement confirmed the molecular formula [ $C_{17}H_{18}O_3 = m/z$  270.3015 as the base peak].

In addition to cyclised product (148) the desired nitrile (147) was obtained in very low yield. Its structure was confirmed by the spectroscopic data, in particular an i.r. band at  $\bar{\nu}_{\text{max}}$  2220  $\text{cm}^{-1}$  and a  $^{13}\text{C}$  resonance at  $\delta_{\text{C}}$  115.2 arising from the cyano group. The  $^1\text{H}$  n.m.r. spectrum showed the expected AA' BB' [ $\delta_{\text{H}}$  7.10(d, J8.7Hz.), 6.99(d, 8.5Hz.) and AB [6.85(d, J6.21Hz.); 6.81(d, J6.5Hz.)] systems, three methoxyls at  $\delta_{\text{H}}$  4.00, 3.84 and 3.77 and a broad multiplet at  $\delta_{\text{H}}$  2.84 for the benzylic protons. The molecular formula  $C_{18}H_{19}NO_3$  was established by high resolution mass spectrometry. The parent ion is at  $m/z$  297.2362, ( $C_{18}H_{19}NO_3$  requires 297.3570) and the base peak at  $m/z$  121.0653 is due to the fragment (155) ( $C_8H_9O^+$ ).

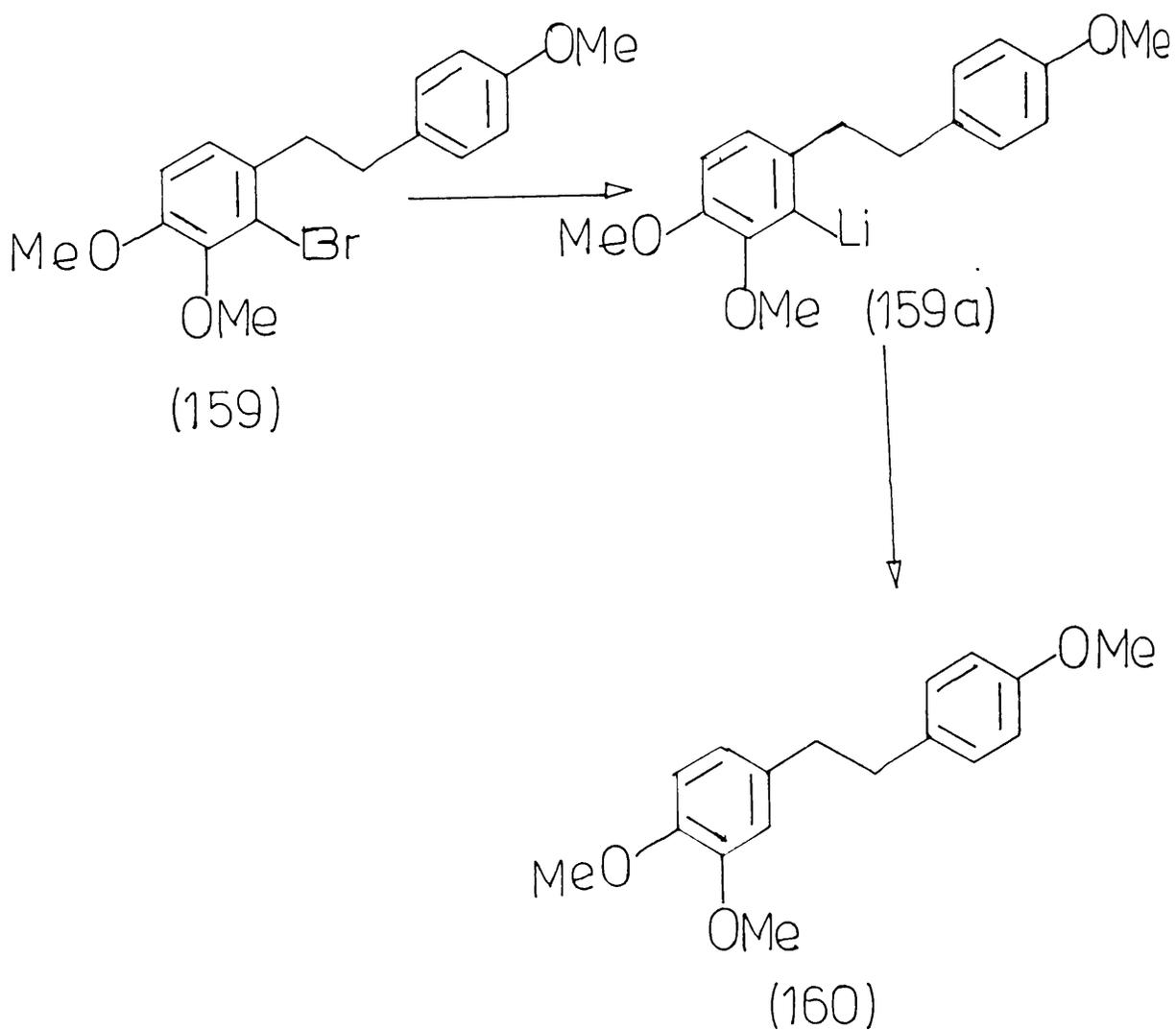
Attempted hydrolysis of the nitrile (147) in 40% sodium hydroxide at reflux for 3 hours failed to produce acidic product (131). The nitrile was recovered unchanged. The failure of this reaction and the problem of the low yield in the formation of the nitrile led us to abandon this route in favour of direct carboxylation of an aromatic bromo-precursor.



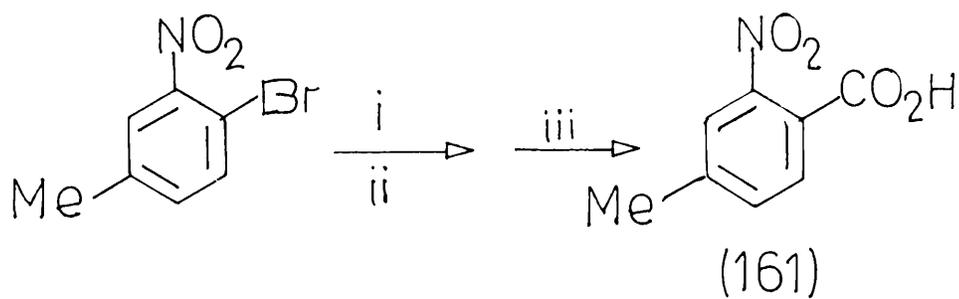
2-Bromovanillin was converted to bromoveratraldehyde (156) using methyl iodide and anhydrous potassium carbonate. The  $^1\text{H}$  n.m.r. spectrum of (156) confirmed the presence of two methoxy groups [ $\delta_{\text{H}}$  3.98 and 3.80]. The aldehyde ( $\delta_{\text{H}}$  10.75) and two ortho-coupled aromatic protons [ $\delta_{\text{H}}$  7.71 and 6.95, AB q, J9.8Hz.] The mass spectrum confirmed the molecular formula  $\text{C}_9\text{H}_9\text{BrO}_3$  [ $m/z$  246 as the base peak].

A Wittig reaction of the aldehyde (156) with the ylid (139a) gave the desired stilbene product as a mixture of Z (157) and E - (158) isomers. In this sequence the isomer (158) predominated, on the evidence of the  $^1\text{H}$  n.m.r spectrum of the mixture. It showed a pair of doublets (J16.6Hz.) at  $\delta_{\text{H}}$  7.35 and 6.68 for the olefinic protons. The aromatic protons appeared as AA' BB' [ $\delta_{\text{H}}$  7.44 and 7.08] and AB [ $\delta_{\text{H}}$  6.90 and 6.86] systems while the three methoxy groups resonated <sup>at</sup>  $\delta_{\text{H}}$  3.88, 3.86 and 3.79. Mass spectrometry gave the molecular formula as the base fragment ( $m/z$  348  $\text{C}_{17}\text{H}_{17}\text{BrO}_3$ ) (Scheme 16).

The 2-bromobibenzyl (159) was obtained from the stilbene mixture in good yield by hydrogenation for 8 hours. Its  $^1\text{H}$  n.m.r. spectrum confirmed its structure [ $\delta_{\text{H}}$  7.43 (d, J8.8Hz.,  $\text{H}_3, \text{H}_5$ ), 7.14 (d, J8.6Hz.,  $\text{H}_2, \text{H}_6$ ); 6.84 (d, J8.6Hz.,  $\text{H}_5$ ); 6.81 (d, 8.9Hz, H-6)]; and three singlets of methoxy groups at 3.85, 3.84 and 3.76 and a broad multiplet at  $\delta_{\text{H}}$  2.86. The mass spectrum confirmed its molecular formula ( $m/z$  350,  $\text{C}_{17}\text{H}_{19}\text{BrO}_3$  requires 350).



Scheme 17



(1)  $\text{C}_4\text{H}_9\text{Li}$ ; (ii)  $\text{THF}/-100^\circ$ ; (iii)  $\text{CO}_2$

Scheme 18

The bromobibenzyl (159) was subjected to the typical halogen metal exchange conditions in the hope of producing the desired carboxylic acid (131). Thus, n-butyl lithium was added to the bromo-compound (159) in dry THF and the solution stirred under nitrogen at  $-78^{\circ}\text{C}$  for 1 hour to form the lithio-derivative (159a) (Scheme 17). The reaction mixture was then quenched with carbon dioxide gas for 1 hour. Unfortunately the major product obtained proved to be the debromobibenzyl (160). None of the desired carboxylic acid (131) was detected. The debromobibenzyl (160) was obtained in reasonable yield and its structure followed readily from its spectroscopic properties. Its mass spectrum confirmed the molecular formula with the base peak at  $m/z$  272 ( $\text{C}_{17}\text{H}_{19}\text{O}_3$ ) and major fragments at  $m/z$  151 and 121. In the  $^1\text{H}$  n.m.r. spectrum the typical AA' BB' system of the top ring again appeared [ $\delta_{\text{H}}$  7.08(d, J8.6Hz.), 6.89(d, J8.7Hz.)]. The substitution pattern of the other ring [ $\delta_{\text{H}}$  6.79(d, J8.1Hz.), 6.72(dd, J8.0, 1.8Hz), 6.62(d, J1.8Hz)] clearly revealed the loss of the bromo substituent. The remaining features of the spectrum included three methoxyl groups [ $\delta_{\text{H}}$  3.85, 3.83 and 3.77] and a broad singlet ( $\delta_{\text{H}}$  2.86) arising from the four benzylic protons.

The failure of the carboxylation step is disappointing. Normally lithiation followed by quenching with carbon dioxide should give reasonable yields of carboxylic acid products.

4- Bromo- 3- nitro toluene<sup>80</sup> for example, afforded the corresponding acid (161) in 80% yield (scheme 18). Clearly moisture has been allowed to enter the system prior to the carbon dioxide. The formation of the debromobibenzyl (160) proves beyond doubt that lithio intermediate (159a) was formed.

## EXPERIMENTAL

## General Experimental

Melting-points (m.p.) which are uncorrected, were determined on a Kofler hot-stage apparatus. Infrared (i.r.) spectra were recorded in  $\text{CCl}_4$  solution on either Perkin Elmer 580 or 257 instruments. Mass spectra (m.s.) were recorded using an MS12 instrument (low resolution) and an MS902S instrument (high resolution). Unless otherwise stated, nuclear magnetic resonance (n.m.r.) spectra were recorded for  $\text{CDCl}_3$  solutions at 90 MHz, on a Perkin-Elmer R.32 instrument. Highfield n.m.r. spectra were recorded using a Bruker WP200SY instrument ( $^1\text{H}$ , 200 MHz,  $^{13}\text{C}$  50.32 MHz). Chemical shifts were measured using the  $\delta$  scale with tetramethylsilane as internal standard or relative to  $\text{CHCl}_3$  at  $\delta_{\text{H}}$  7.25 or  $\text{CDCl}_3$  at  $\delta_{\text{C}}$  77.0. Column chromatography was carried out on Merck silica HF<sub>254</sub>. Kieselgel GF<sub>254</sub> was used for preparative thin layer chromatography. Analytical t.l.c. plates were visualised using u.v. light (254 or 350nm) and by spraying with ceric sulphate  $\text{H}_2\text{SO}_4$ . Eluants for column chromatography were increasing percentages of Ethyl acetate in petroleum ether. All solvents and reagents used were of analytical grade except for column chromatography when bulk solvents were used. Solvents were removed using a Büchi rotary evaporator and water aspirator. Petroleum ether refers to the fraction boiling between 60°C and 80°C. Organic solutions were dried over anhydrous magnesium sulphate or sodium sulphate.

Methyl 4-methoxybenzoate (136)

A mixture of 4-hydroxybenzoic acid (20 g, 0.144 mole), anhydrous potassium carbonate  $K_2CO_3$  (50 g) and dimethyl sulphate (40 ml) in acetone (200 ml) was refluxed for 5 hr. The acetone was removed and the residue diluted with water (250 ml) and ether (250 ml). The organic layer was washed with concentrated ammonia solution (2 x 50 ml) and then water (50 ml). The ethereal layer was dried over anhydrous sodium sulphate and evaporated to give a white precipitate of methyl-4-methoxy benzoate (136) (20.9 g, 87%); m.p.  $51^{\circ}C-52^{\circ}C$ .

$\delta_H$ : 7.58(d, J8.2Hz.), 6.55(d, J8.2Hz.), 4.02(s,  $-CO_2CH_3$ ), 3.95(s,  $-OCH_3$ ).

M.S.: 166( $M^+$ ). 135( $M^+ - OCH_3$ , base peak), 107( $M^+ - CO_2Me$ ). 31( $-OCH_3$ ).

4-Methoxybenzyl alcohol (137)<sup>A</sup>

A solution of methyl 4-methoxybenzoate (136) (20.8 g, 0.125 mole) in dry ether (50 ml) was added over a period of 20 min to a solution of lithium aluminium hydride (6.5 g, 0.175 mole) in dry ether (20 ml). Gentle refluxing was continued for 3 hr. The chilled mixture was then decomposed by careful addition of aqueous sodium sulphate. The aqueous phase was extracted thoroughly with ether and the combined organic layers were washed with 5% sodium bicarbonate and dried over sodium sulphate. Distillation afforded (137)

[15.6 g, 90%] as a viscous oil, b.p. 80/0.2 mm)

$\delta_{\text{H}}$ : 7.15(d, J8.6 Hz.), 6.86(d, J8.5Hz.), 4.45(S,  $-\text{CH}_2\text{OH}$ ),  
3.71(S,  $-\text{OCH}_3$ ), 3.68(S, exchangeable with  $\text{D}_2\text{O}$ ).

$\bar{\nu}_{\text{max}}$  3450  $\text{cm}^{-1}$

#### 4-Methoxybenzyl chloride (138)<sup>8</sup>

A stirred solution of alcohol (137) (15.6 g, 0.118 mole) and dimethyl aniline (12 g, 0.099 mole) in anhydrous toluene (80 ml) was slowly treated with redistilled thionyl chloride (9.2 ml 0.126 mole), in toluene (15 ml). The mixture was warmed to room temperature, and then heated under reflux for 1 hr. The organic layer was separated, washed, dried and evaporated to give a brown oil which on distillation yielded 4-methoxybenzyl chloride (138) (15.3, 89%) as a colourless oil b.p. 95/0.2 mm.

$\delta_{\text{H}}$ : 7.42(d, J8.6Hz.), 6.88(d, J8.7Hz.), 4.54(S,  $-\text{CH}_2\text{Cl}$ )  
3.64(S,  $-\text{OCH}_3$ ).

#### 4-Methoxybenzyltriphenylphosphonium chloride (139)

The benzyl chloride (138) (15 g, 0.10 mole) and triphenylphosphine (29 g, 0.11 mole) were stirred and heated in anhydrous toluene under  $\text{N}_2$  at  $60^\circ\text{C}$  for 46 hr. The hygroscopic precipitated phosphonium chloride (139) (36 g, 84%) was separated by filtration, washed with light petroleum ether, then dried in vacuo. It had m.p.  $137^\circ\text{C}$ .

$\delta_{\text{H}}$ : 7.60(15H, br m,  $-\text{Ph}_3\text{P}$ ), 6.89(d, J9.0Hz.), 6.58(d, J8.9Hz.), 5.21(d, 14.0Hz.), 3.62(S,  $-\text{OCH}_3$ ).

Preparation of Acetyl vanillin (140)

Vanillin (10 g) in pyridine (10 ml) and acetic anhydride (12 ml) were stirred at room temperature overnight. Methanol (8 ml) was added, the temperature was increased to 50°C for 1 hr. Cooling and addition of water gave a white precipitate which was filtered and dried under pump pressure to yield the acetate (140) (11.8 g, 93.4% yield), m.p. 75°C-76°C.

$\delta_{\text{H}}$ : 10.11(S,  $-\text{CHO}$ ), 7.68(dd, J8.7, 2.7Hz.), 7.41(d, J8.9 Hz.), 7.31(d, J8.8Hz), 3.93(S,  $-\text{OCH}_3$ ), 2.35(S,  $-\text{OCO CH}_3$ ).

M.S.: 194( $\text{M}^+$ ), 151( $\text{M}^+ - \text{COCH}_3$ , base peak), 43( $\text{M}^+ - 152$ )

2-Nitroacetylvanillin (141)<sup>81</sup>

Dry acetyl vanillin (140) (11.2 g, 0.057 mole) was stirred in an ice bath. Red fuming nitric acid (15 ml, d 0.5) was slowly added in small portions. The temperature was maintained below 0°. The solid dissolved to give a dark-red solution. When the addition of nitric acid had been completed, the contents of the flask were poured on to ice. The crude product separated as a yellow precipitate. It was collected by filtration and well washed with water.

Crystallisation from aqueous ethanol afforded 2- nitroacetylvanillin (10.2 g, 73%) m.p. 70 - 72°C.

$\delta_{\text{H}}$ : 10.15(S, -CH $\underline{\text{O}}$ ), 7.85(d, 9.8Hz.), 7.58(d, 9.9Hz.),  
3.95(S, -OCH $\underline{\text{H}}_3$ ) 2.32(S, -COCH $\underline{\text{H}}_3$ )

M.S. 239(M $^+$ ), 196(M $^+$  - COCH $\underline{\text{H}}_3$ ), 43(M $^+$  - 196, base Peak)

2- Nitroveratraldehyde (142)<sup>74</sup>

2- Nitroacetylvanillin (141) (11 g) in water (50 ml) and dimethyl sulphate (30 ml) were stirred vigorously while 50% aqueous sodium hydroxide (18 ml) was added dropwise during one hour, the pH being kept at about 8 and the temperature at 35° - 42° C throughout. Further dimethyl sulphate (10 ml) was then added followed by more alkali until the pH was 11. After further stirring for 20 min., the mixture was cooled, conc. ammonia (10 ml) was added and the reaction left at 0°C. The product was filtered off, washed with water (50 ml) and crystallised from ether to give 2- nitroveratraldehyde (142) (8.2 g, 84% yield), m.p. 61 - 62.5 °C [Lit, 63.5 - 64°].

$\delta_{\text{H}}$ : 9.77(S, -CH $\underline{\text{O}}$ ), 7.64(d, J8.7Hz.), 7.12(d, J8.7Hz.)  
4.00(S, -OCH $\underline{\text{H}}_3$ ), 3.92(S, -OCH $\underline{\text{H}}_3$ )

M.S. 211(M $^+$ ), 165(M $^+$  - NO $_2$ ), 137(166-CO).

2-Nitro-3,4,4'-trimethoxystilbene (143)<sup>83</sup>

To 4-methoxybenzyl<sup>triphenyl</sup>phosphonium chloride (139) (9.92 g, 0.02 mole) in dry THF (50 ml) was added n. butyl lithium (12 ml, d, 0.693) and the mixture stirred under nitrogen at -50°C for 1 hr. 2-Nitroveratraldehyde (142), (5g, 0.023 mole) in THF (15 ml) was added and the stirring was continued for 2 hours at room temperature. The mixture was then acidified with dilute hydrochloric acid and extracted with ether (2 x 40 ml). Evaporation of the solvent afforded a pale yellow oil (7.8 g) which showed mainly 2 spots on t.l.c. of r.f. = 0.5 and 0.2. These spots were separated by flash column chromatography in EtOAc: Petrol (0 - 100%). Fractions 7, 8 and 9 were found to contain a single crystalline product (r.f. = 0.2) identical with triphenylphosphine oxide. Fractions (1 - 6) were due to the stilbene as a mixture of Z - (143) and E - (144) forms [3.8 g, 50.94%].

$\delta_{\text{H}}$ : 7.09(d, J8.39Hz.), 6.94(d, J8.51Hz.), 6.79(d, 8.7Hz.)  
6.72(d, J8.85), 6.60(d, J11.87Hz.), 6.25(d, J11.8Hz.)  
3.93(s, -OCH<sub>3</sub>), 3.86(s, -OCH<sub>3</sub>), 3.75(s, -OCH<sub>3</sub>)

$\delta_{\text{C}}$ : 158.99(s), 152.06(s), 140.74(s), 133.31(d), 130.23(d),  
128.41(s), 125.15(d), 122.73(s), 120.43(d), 114.10(s)  
113.61(d), 62.05(q), 56.16(q), 55.11(q).

M.S. m/z 315.1102 (C<sub>17</sub>H<sub>17</sub>NO<sub>5</sub> requires 315.1107)

2-Nitro- 3,4,4', trimethoxybibenzyl (145)

2-Nitro- 3,4,4'- trimethoxystilbene mixture (3.5 g, 0.011 mole) in methanol (100 ml) and 5% Pd-C (0.4 g) was stirred under hydrogen for 2 hr. Filtration of the catalyst and evaporation of the solvent afforded the bibenzyl (145) (3.2 g, 93%) as an oil.

$\delta_{\text{H}}$ : 7.04(d, J8.99Hz.), 6.92(d, J8.82Hz.), 6.89(d, J8.74Hz.), 6.83(d, J8.62Hz.), 3.92(S, -OCH<sub>3</sub>), 3.87(S, -OCH<sub>3</sub>), 3.77(S, -OCH<sub>2</sub>) and 2.77(4H, br m).

$\delta_{\text{C}}$ : 157.94(s), 151.35(s), 140.52(s), 132.72(s), 129.29(d), 125.23(d), 124.91(d), 123.84(s), 113.80(d), 113.73(d), 101.65(s), 61.94(q), 55.54(q), 55.13(q), 35.00(t), 32.93(t).

M.S. m/z 317.2681, (C<sub>17</sub>H<sub>19</sub>NO<sub>5</sub> requires 317.1263)

2- Amino- 3,4,4'- trimethoxybibenzyl (146)

Lithium aluminium hydride (6 g) was carefully added in small portions to a solution of <sup>the</sup> 2- nitrobibenzyl (145) (3 g, 9.46 x 10<sup>-3</sup> mole) in dry ether (50 ml). Gentle refluxing was continued for 2 hr. The reaction was worked up by adding aqueous sodium sulphate followed by hydrochloric acid (5 N, 10 ml). The aqueous phase was extracted thoroughly with ether, and the combined organic layers washed with 5% sodium bicarbonate and dried anhydrous magnesium sulphate. The product (146) was obtained as a yellow solid (1.9 g, 70% yield) m.p. 92°C.

$\delta_{\text{H}}$ : 7.08(d, J8.73Hz.), 6.81(d, J8.7Hz.), 6.78(d, J8.54Hz.),  
6.35(d, J8.48Hz.), 3.82(s, OCH<sub>3</sub>), 3.80(s, -OCH<sub>3</sub>),  
3.78(s, -OCH<sub>3</sub>), 2.95(br. m. 4H), 2.16(s, NH<sub>2</sub>, exchange-  
able with D<sub>2</sub>O).

$\delta_{\text{C}}$ : 157.84(s), 151.02(s), 138.53(s), 133.87(s), 129.29(d),  
129.26(d), 123.85(d), 119.87(s), 118.24(d), 113.74(d),  
113.57(d), 101.66(s), 59.76(q), 55.56(q), 55.16(q),  
34.60(t), 33.14(t).

M.S.  $m/z$  287.1513 (C<sub>17</sub>H<sub>21</sub>NO<sub>3</sub> requires 287.1521).

$\bar{\nu}_{\text{max}}$ : 3420 cm<sup>-1</sup>.

2- Cyano- 3,4,4'- trimethoxybibenzyl (147)<sup>82</sup>

2- Amino- 3,4,4'- trimethoxybibenzyl (146) (1 g) was diazotised in dilute HCl (10 ml, 2 N) with sodium nitrite (0.5 g, 7.2 x 10<sup>-3</sup> mole) at 0°C. The resulting diazonium salt solution was added over 75 min with stirring, simultaneously with potassium cyanide (0.6 g, 9.2 x 10<sup>-3</sup> mole) in H<sub>2</sub>O (8 ml) to a hot solution of CuCN-KCN at 80° - 5° [obtained by adding NaHSO<sub>4</sub> (0.3 g) and potassium hydroxide (0.5 g) in H<sub>2</sub>O (10 ml) to cupric sulphate CuSO<sub>4</sub>.5H<sub>2</sub>O (0.8 g) and NaCl (1 g) in H<sub>2</sub>O (7 ml) at 85°, cooling, decanting the supernatant liquid, diluting with H<sub>2</sub>O (8 ml) and adding potassium cyanide (0.5 g)] The mixture was stirred for 30 mn at 85°. Cooled and filtered, then the filtrate residue was extracted with EtOAC (2 x 30 ml) which yielded a crude product of (0.8 g) and showed mainly 2 spots on t.l.c. These spots were separated by

flash column chromatography in EtOAc; petroleum gave the nitrile (147) in very low yield (0.025 g, 2.5%) and the cyclised product (148) in good yield (0.5 g, 54%).

(i) the nitrile (147); n.m.r. signals at.

$\delta_{\text{H}}$ : 7.10(d, J8.7Hz.), 6.99(d, J8.53Hz.), 6.85(d, J6.34Hz.)  
6.81(d, J6.58Hz.), 4.00(s, -OCH<sub>3</sub>), 3.84(s, -OCH<sub>3</sub>),  
3.77(s, -OCH<sub>3</sub>), 2.84(br m, 4H)

$\delta_{\text{C}}$ : 158.01(s), 150.55(s), 137.72(s), 132.76(s), 129.43(d),  
129.36(d), 124.61(d), 116.73(d), 115.21(s), 113.77(d),  
61.54(q), 56.00(q), 55.11(q), 36.29(t)

M.S.  $m/z$  297.2362 (C<sub>18</sub>H<sub>19</sub>NO<sub>3</sub> requires 297.1365)

ir. : 2220 ; 1610 cm<sup>-1</sup>

(ii) The cyclised compound (148)

$\delta_{\text{H}}$ : 8.12(d, J2.72Hz.), 7.18(d, J8.26Hz.), 6.96(d, J8.20 Hz.)  
6.82(dd, J2.69, 8.24Hz.), 6.78(d, J8.2 Hz.),  
3.90(s, OCH<sub>3</sub>), 3.84(s, OCH<sub>3</sub>), 3.73(s, -OCH<sub>3</sub>),  
2.73(br, 4H).

$\delta_{\text{C}}$ : 158.34(s), 152.14(s), 147.00(s), 133.40(s), 132.33(s)  
130.96(s), 129.44(d), 129.41(s), 123.06(d) 113.62(d)  
110.99(d), 60.19(q), 55.93(q), 55.16(q), 29.75(t)  
29.24(t)

M.S.:  $m/z$  270.3015 (C<sub>17</sub>H<sub>18</sub>O<sub>3</sub> requires 270.1266)

Preparation of 2- bromoveratraldehyde (156)

2- Bromovanillin (7.8 g, 0.033 mole) was heated in acetone (50 ml) with iodomethane (30 ml) and anhydrous potassium carbonate (50 g, 0.36 mole) under reflux on water bath for 6 hr. The inorganic material was removed by filtration; the residue was washed 3 times with acetone and evaporated. To the crystalline residue 10% sulphuric acid (10 ml) was added and dried at room temperature. Chromatography over alumina in  $\text{CHCl}_3$  yielded prisms of 2- bromoveratraldehyde (156) (6 g, 74%); m.p.  $75 - 77^\circ$ .

$\delta_{\text{H}}$ : 10.75(S,  $-\text{CHO}$ ), 7.71(d, J9.8Hz.), 6.95(d, J9.8Hz.),  
3.98(S,  $-\text{OCH}_3$ ), 3.80(S,  $-\text{OCH}_3$ )

M.S.: 244( $\text{M}^+$  base peak), ( $\text{C}_9\text{H}_9 \text{ BrO}_3$  requires 244).

2- Bromo- 3,4,4'- trimethoxystilbene

A mixture of 4- methoxy <sup>benzyl</sup>triphenylphosphonium chloride (139) (8 g, 0.019 mole) and n-butyl lithium (10 ml, d 0.693) was stirred at  $-50^\circ$  in dry THF (50 ml) under  $\text{N}_2$  for 1 hr. To this a solution of 2- bromoveratraldehyde (156) (5 g, 0.020 mole) in THF (10 ml) was added, and the mixture stirred at room temperature for 18 hr. The mixture was then acidified with dilute HCl and extracted with ether (50 ml). The organic layer on evaporation gave a yellow oil which showed mainly 3 spots on t.l.c. Column chromatography in petrol: EtOAc afforded a mixture of Z- (157) and E- (158) of

2-bromo-3,4,4'-trimethoxystilbene (6 g, 84%), and triphenylphosphine oxide (1.5 g) together with the unchanged phosphonium chloride (139) (1 g).

$\delta_{\text{H}}$ : 7.44(d, J8.67Hz.), 7.35(d, J16.65Hz.),  
7.08(d, J8.67Hz.), 6.90(d, J8.87Hz.),  
6.86(d, J8.77Hz.), 6.68(d, J8.88Hz.),  
3.88(s, -OCH<sub>3</sub>), 3.86(s, -OCH<sub>3</sub>), 3.79(s, -OCH<sub>3</sub>)

$\delta_{\text{C}}$ : 159.56(s), 152.42(s), 146.63(s), 131.54(d),  
132.28(d), 129.91(s), 127.63(d), 125.31(s),  
121.62(d), 119.72(s), 114.25(d), 113.61(d),  
113.32(d), 111.7(d), 56.32(q), 55.67(q),  
55.43(q)

M.S.  $m/z$  348(M<sup>+</sup>, base peak); (C<sub>17</sub>H<sub>17</sub>BrO<sub>3</sub> requires 348)

2-Bromo-3,4,4'-trimethoxybibenzyl (159)

2-Bromo-3,4,4'-trimethoxystilbene mixture (0.6 g, 0.017 mole) in EtOAc (200 ml) and 10% Pd-C (0.3 g) was stirred under hydrogen for 8 hr. Filtration of the catalyst, and evaporation of the solvent afforded the bibenzyl (5.8 g, 96%). Recrystallisation from hexane yielded (159) (4 g) as prisms m.p. 88 - 90<sup>o</sup>.

$\delta_{\text{H}}$ : 7.43(d, J8.79Hz.), 7.14(d, J8.62Hz.),  
6.84(d, J8.65Hz.), 6.81(d, J8.98Hz.)  
3.85(s, -OCH<sub>3</sub>), 3.84(s, -OCH<sub>3</sub>), 3.76(s, -OCH<sub>3</sub>),  
2.86(br m, 4H)

M.S. 350(M<sup>+</sup>), 229 (M<sup>+</sup> - C<sub>8</sub>H<sub>10</sub>O), 121(M<sup>+</sup> - 229, base peak)

3,4,4'- Trimethoxybibenzyl- 2- carboxylic acid (131)

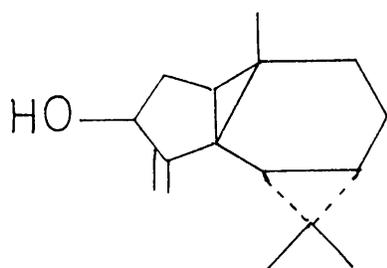
2- Bromo- 3,4,4'- trimethoxybibenzyl (159) (1 g,  $2.85 \times 10^{-3}$  mole) in dry THF and *n*-butyl lithium (15 ml, d, 0.647) were stirred at  $-78^{\circ}\text{C}$  under  $\text{N}_2$  for 1 hr. Dry carbon dioxide gas was bubbled into this solution for 1 hr. Water (30 ml) was added, and the THF removed by evaporation. The residue was acidified with dil HCl and extracted with EtOAc (2 x 50 ml). The organic layer was evaporated to yield a solid material (0.88 g). Crystallization from ether afforded the debromobibenzyl (160) (0.7 g ) m.p.  $90 - 91^{\circ}$ . No trace of the carboxylic acid (131) was found.

$\delta_{\text{H}}$ : 7.08(d, J8.65Hz.,  $\text{H}_{-3',5'}$ ), 6.82(d, J8.75Hz.,  $\text{H}_{2',6'}$ ),  
6.79(d, J8.07Hz., H-5), 6.72(dd, J806, 1.8Hz.,  $\text{H}_6$ ),  
6.62(d, J1.85Hz., H-2), 3.84, 3.81, 3.77(each 3 H, s)  
and  $\text{A}_2\text{B}_2$  at  $\delta_{\text{H}}$  2.82(4H, br)

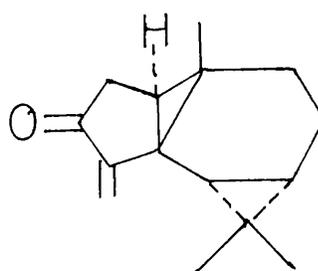
$\delta_{\text{C}}$ : 159.24(s), 157.24(s), 149.21(s), 147.31(s)  
134.66(d), 133.91(d), 130.90(d), 129.53(d)  
127.62(d), 120.43(d), 119.61(d), 113.82(s)  
59.74(q), 55.97(q), 55.36(q), 37.80(t)  
37.28(t)

M.S. 272( $\text{M}^+$ ), 151( $\text{M}^+ - 121$ , base peak), 121( $\text{M}^+ - 151$ ).

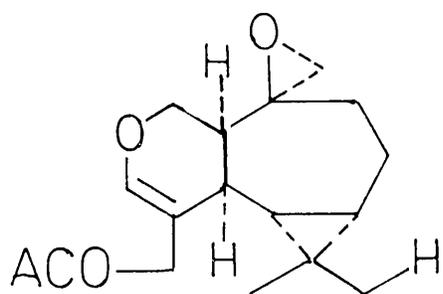
C H A P T E R 3



(162)



(163)



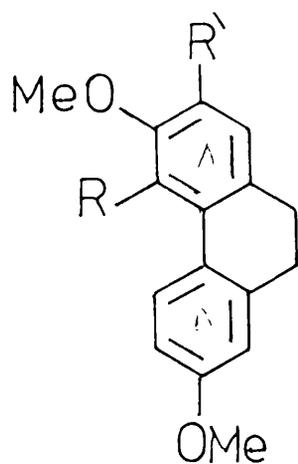
(164)

## INTRODUCTION

Plagiochilaea consists of two main groups Mylia and Plagiochila, although some authorities place Mylia in a separate subfamily of the Jungermanniales. Plagiochila belongs to Jungermanniales and more than 1500 species are known in the world. From Japan 20 species have been reported<sup>17</sup>.

Mylia species have proved to be a very rich source of terpenoids. Mylia taylorii, for example, contains sesquiterpenoids of the aromadendrane class, myliol (162), dehydromylione A (163) and taylorine (21). These might suggest that there is a similarity between M. taylorii and Plagiochila species which also contain aromadendranes and secoaromadendranes. Asakawa et al<sup>17</sup> reported the investigation of fourteen Plagiochila species and showed that the characteristic constituents are ent-secoaromadendranes e.g. plagiochilide (18). It is known that there are two chemical types, one with the sharp, pungent substance plagiochiline A (164), e.g. P. asplenioides, P. hattoriana, P. pulcherrima and the other type which lacks this pungent substance e.g. P. acanthophyle subsp. Japonica and P. arbuscula.

Phenanthrene and dihydrophenanthrene derivatives are rare amongst the Hepaticae. 3, 4-Dimethoxy-5-hydroxy-9, 10-dihydrophenanthrene (100) was the first example isolated from the liverwort Riccardia jackii<sup>38</sup>. Connolly<sup>73</sup> isolated the dihydrophenanthrene derivatives (165) - (170)



(165)  $R = \text{OMe}, R' = \text{OH}$

(165a)  $R = \text{OMe}, R' = \text{OAC}$

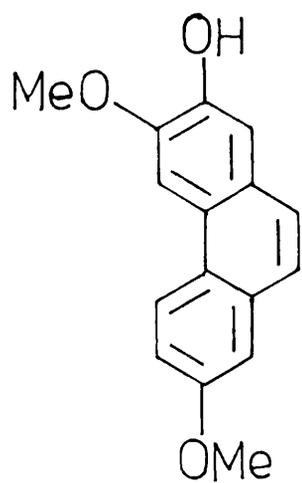
(166)  $R = R' = \text{OMe}$

(167)  $R = \text{OMe}, R' = \text{H}$

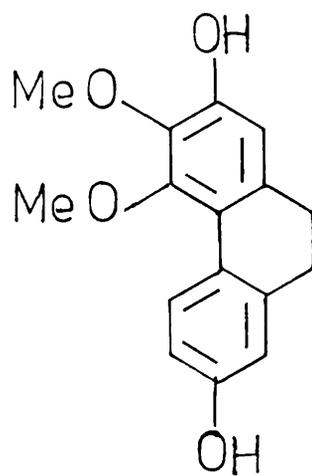
(168)  $R = \text{OH}, R' = \text{H}$

(169)  $R = \text{H}, R' = \text{OH}$

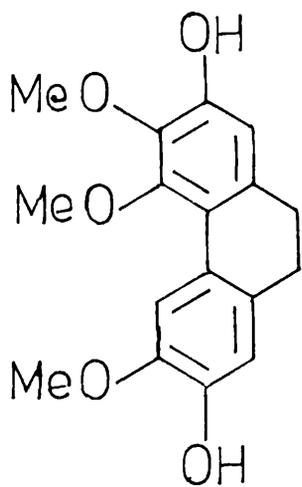
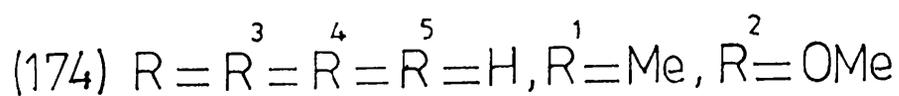
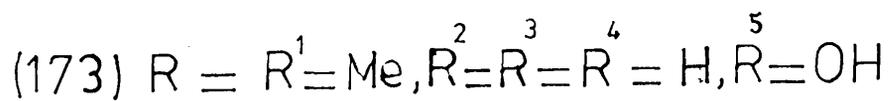
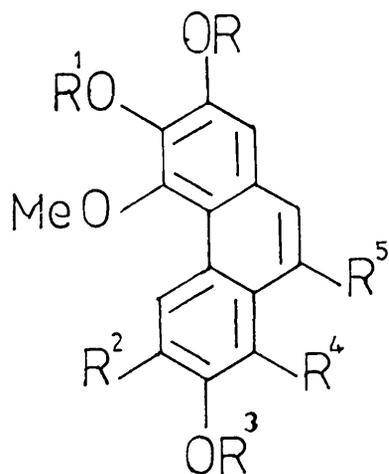
(170)  $R = \text{H}, R' = \text{OMe}$



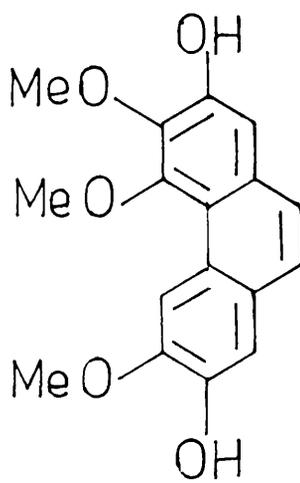
(171)



(172)



(175)

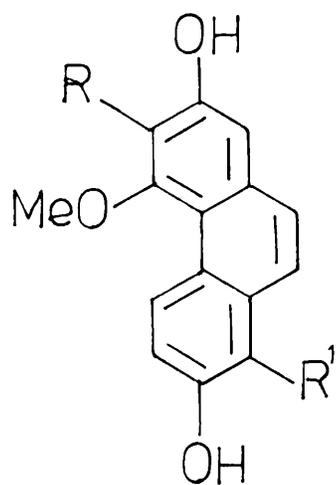


(176)

from the Scottish liverwort Plagiochila spinulosa (vide infra). Recently Asakawa<sup>84</sup> isolated a new phenanthrene derivative (171) from the Indian liverwort Marchantia polymorpha.

A large number of papers deal with the isolation of phenanthrene and dihydrophenanthrene derivatives from higher plant families. Majumder<sup>85</sup> reported the isolation of erianthridin (172) a new 9, 10- dihydro- phenanthrene derivative from the orchids Eria carinata and Eria stricta. From the orchid Bulbophyllum gymnopus, Majumder<sup>86</sup> also isolated another new phenanthrene derivative, gymnopusin (173) together with 2,7- dihydroxy- 3,4,6- trimethoxyphenanthrene (174) and the 9, 10- dihydro derivative (175). Stermitz<sup>87</sup> found a large number of phenanthrenes and the dihydrophenanthrene derivatives (175) and (176) - (178) in Oncidium cebollata, a peyote- replacement plant.

A reinvestigation of Plagiochila spinulosa was undertaken in order to characterise fully the aromatic constituents and to obtain sufficient quantities for biological testing. This chapter describes the isolation and spectroscopic analysis of several dihydrophenanthrenes and other aromatic compounds from P. spinulosa. The synthesis of 2- hydroxy- 3,7- dimethoxy- 9,10,- dihydrophenanthrene (169) is also described.



(177)  $R = \text{OMe}, R' = \text{H}$

(178)  $R = \text{H}, R' = \text{OMe}$

## DISCUSSION

### Constituents of *Plagiochila spinulosa*

Preliminary studies of the constituents of *P. spinulosa* by Harrison<sup>89</sup> afforded 2- hydroxy- 3,4,7- trimethoxy- 9,10- dihydrophenanthrene (165) as the major component accompanied by 3,4,7- trimethoxy- 9,10- dihydrophenanthrene (167) and the pentasubstituted benzene derivative methyl 6-methoxy- 3,4- methylenedioxy- 2- methylbenzoate (189). In the present work these compounds were again obtained.

It was clear from the <sup>1</sup>H n.m.r. spectrum of the major compound (165), C<sub>17</sub>H<sub>18</sub>O<sub>4</sub>, m/z 286, that it is a dihydrophe- nanthrene with three methoxyl groups ( $\delta_{\text{H}}$  3.96, 3.83, and 3.73) and a phenolic hydroxyl ( $\delta_{\text{H}}$  5.67, s, exchangeable with D<sub>2</sub>O). The protons arising from the two methylene groups appear as a complex multiplet at  $\delta_{\text{H}}$  2.69. A highly deshielded aromatic proton [ $\delta_{\text{H}}$  8.17 (d, J8.4Hz)] must be attached to C-5 (or C-4). It has an ortho- neighbour [ $\delta_{\text{H}}$  6.80 (dd, J8.5, 2.9Hz)] which is in turn coupled to a meta- proton [ $\delta_{\text{H}}$  6.76 (d, J2.9Hz)]. These data readily established the substitution pattern of ring B which must have an oxygen substituent at C-7. Ring A must have three oxygen substituents since the remaining aroma- tic proton is present as a singlet [ $\delta_{\text{H}}$  6.11]. This proton is probably H-1 since it couples with one of the methylene groups at 360 MHz. Thus the compound is a trimethyl ether (165) of 2,3,4,7- tetrahydroxy- 9, 10- dihydrophenanthrene.

The  $^{13}\text{C}$  n.m.r. shifts of (165) reveal that two of the methoxyl groups [ $\delta_{\text{C}}$  61.1 and 60.0] have no neighbouring aromatic protons and are situated at C-3 and C-4. Small shifts of H-1 in the acetate (165a) [ $\delta_{\text{H}}$  6.71] and methyl ether (166) [ $\delta_{\text{H}}$  6.58] suggested that the phenolic hydroxyl group is situated at C-2 and that the natural compound is 2-hydroxy-3,4,7-trimethoxy-9,10-dihydrophenanthrene (165). Confirmation of this was obtained from the fully coupled  $^{13}\text{C}$  n.m.r. spectrum of (165) which also provided evidence for the assignment of all the  $^{13}\text{C}$  resonances. C-1, C-2, and C-3 all lost long-range couplings with the phenolic hydroxyl proton following  $\text{D}_2\text{O}$  exchange.

The second compound, the trimethoxy dihydrophenanthrene (167),  $\text{C}_{17}\text{H}_{18}\text{O}_3$ ,  $m/z$  270.1259, has the same substitution pattern in ring B as evidenced by the presence of the spin system H-5 [ $\delta_{\text{H}}$  8.37 (d,  $J_{8.6}\text{Hz}$ )], H-6 [ $\delta_{\text{H}}$  6.83 (dd,  $J_{8.6}, 2.8\text{Hz}$ )] and H-8 [ $\delta_{\text{H}}$  6.77 (d,  $J_{2.8}\text{Hz}$ )]. Ring(A) contains an AB system [ $\delta_{\text{H}}$  6.93 (d,  $J_{8.1}\text{Hz}$ , H-1) and  $\delta_{\text{H}}$  6.75 (d,  $J_{8.1}\text{Hz}$ , H-2)] which must involve H-1 and H-2 since the former is coupled with a methylene group and neither proton is strongly deshielded. Furthermore one methoxyl signal ( $\delta_{\text{C}}$  60.0 attached to C-4) has no proton neighbour. The second compound is therefore, 3,4,7-trimethoxy-9,10-dihydrophenanthrene (167)

An interesting pentasubstituted benzene derivative (189),  $C_{11}H_{12}O_5$ ,  $m/z$  224,  $\nu_{\max}$  1733  $cm^{-1}$  was also isolated. Its spectroscopic properties revealed the presence of an aromatic methyl group [ $\delta_H$  2.14 (d,  $J_{0.5Hz}$ ),  $\delta_C$  12.5], a methoxyl group [ $\delta_H$  3.75 (d,  $J_{0.3Hz}$ ),  $\delta_C$  57.0] a carbomethoxyl [ $\delta_H$  3.87 (s),  $\delta_C$  52.1], a methylenedioxy group [ $\delta_H$  5.92 (2H, s),  $\delta_C$  92.9 (t)] and an aromatic proton [ $\delta_H$  6.29 (s);  $\delta_C$  101.3 (d)]. The  $^{13}C$  chemical shift of the methoxyl indicating that it has a proton neighbour. This is also confirmed by the observation of a small long-range coupling ( $J_{0.3Hz}$ ) between the methoxyl and the aromatic proton. The  $^1H$  chemical shift of the aromatic proton is consistent with the presence of two ortho- oxygen functions. Finally a long-range coupling ( $J_{0.5Hz}$ ) between the aromatic proton and the methyl group favours a para relationship and leads to the structure methyl 2- methyl- 3,4- methylenedioxy- 6- methoxybenzoate (189) for the natural product. This compound is clearly related to orsellenic acid (190) a common polyketide secondary metabolite.

Further work on P. spinulosa by Ifeadike and Singh<sup>73</sup> resulted in the isolation of the lunularic acid derivative (131), whose attempted synthesis has been described earlier, together with the most interesting and unusual constituent spinuloplugin A (192), whose structure was determined by x-ray analysis.<sup>73</sup> Several other compounds were obtained.

These appeared to be dihydrophenanthrenes. In our re-investigation we have isolated and characterised the dihydrophenanthrenes (165) and (167) - (170) and in addition, have obtained spinuloplagin B (191), an isomer of (192), whose structure will be described below.

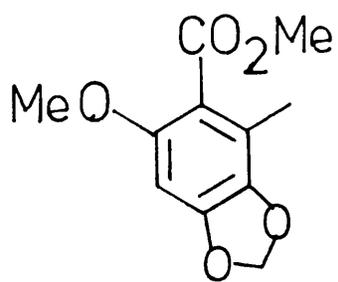
The dihydrophenanthrenes (167) - (170), readily identifiable by the presence of a four proton multiplet for the methylene protons, have the same substitution pattern in ring B as the previous compounds (165) and (170) i.e. a methoxyl group attached to C-7 (see experimental section). The changes in ring A are easily deduced by considering the  $^1\text{H}$  aromatic proton signals and the  $^{13}\text{C}$  chemical shifts of the methoxyl signals.

Compound (169),  $\text{C}_{16}\text{H}_{16}\text{O}_3$ ,  $m/z$  256.1088, has a phenolic hydroxyl [ $\delta_{\text{H}}$  5.42 (s, exchangeable with  $\text{D}_2\text{O}$ )], a methoxyl and two aromatic proton signals [ $\delta_{\text{H}}$  7.45 and 6.66]. The deshielded proton must be attached to C-4. The singlet nature of the aromatic proton <sup>signal</sup> indicates that the oxygen functions are at C-2 and C-3. A decision between a 2- hydroxy- 3- methoxy and a 2- methoxy- 3- hydroxy arrangement was reached by synthesis (see later). The compound is therefore 2- hydroxy- 3,7- dimethoxy- 9, 10- dihydrophenanthrene (169).

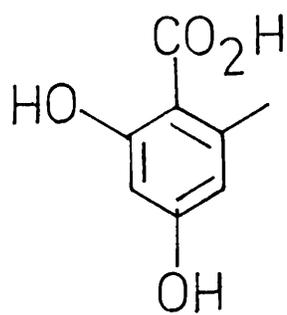
The next compound (170),  $C_{17}H_{18}O_3$   $m/z$  270, is a trimethoxy dihydrophenanthrene. None of the methoxyl groups has a  $^{13}C$  chemical shift in the region of 60 ppm and thus all have at least one ortho proton neighbour. The aromatic protons of ring A again appear as singlets [ $\delta_H$  7.19 (H-4) and 6.73 (H-1)]. The compound is therefore 2,3,7-trimethoxy-9, 10-dihydrophenanthrene (170) and is the methyl ether of (169) above.

The final dihydrophenanthrene (168),  $C_{16}H_{16}O_3$  is isomeric with (169). Ring A again has two oxygen substituents, a methoxyl and a phenolic hydroxyl [ $\delta_H$  3.56 (s, exchangeable with  $D_2O$ )]. The strongly deshielded nature of H-5 [ $\delta_H$  8.37 (d,  $J_{8.7}Hz.$ )] reveals that there is an oxygen substituent on C-4. The fact that it is the hydroxyl group follows from the absence of a methoxyl with a deshielded  $^{13}C$  chemical shift. Thus the compound is 4-hydroxy-3,7-dimethoxy-9, 10-dihydrophenanthrene (168). In agreement with this proposal the ring A protons resonate as an ABq ( $J_{8.1}Hz$ ) at  $\delta_H$  6.74 and 6.69. The  $^{13}C$  chemical shifts of most of the above compounds have been measured and are reported in the experimental section. Complete assignments have been made only for certain compounds.

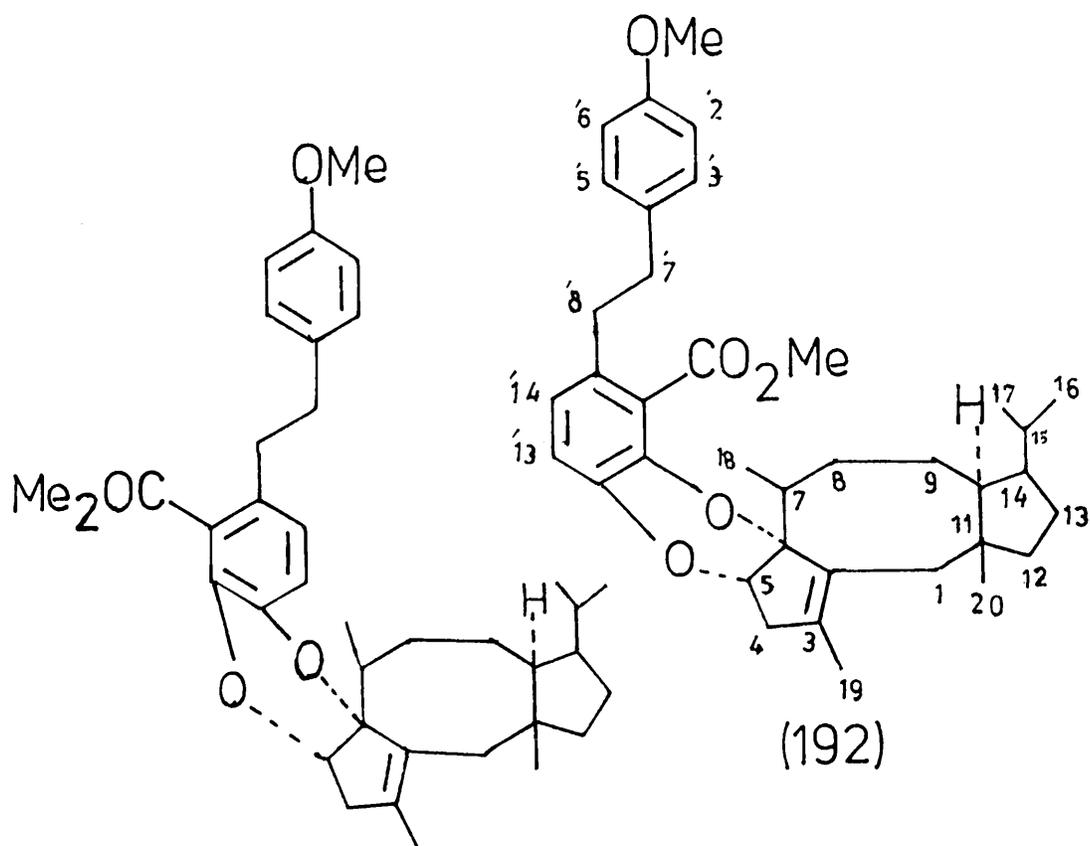
Careful chromatography of the mother liquors of spinuloplogin A (192) afforded, in minor amount, an isomeric compound which we name spinuloplogin B (191)  $C_{37}H_{48}O_5$ ,  $m/z$  572.3527. It is clearly a bibenzyl-diterpenoid conjugate



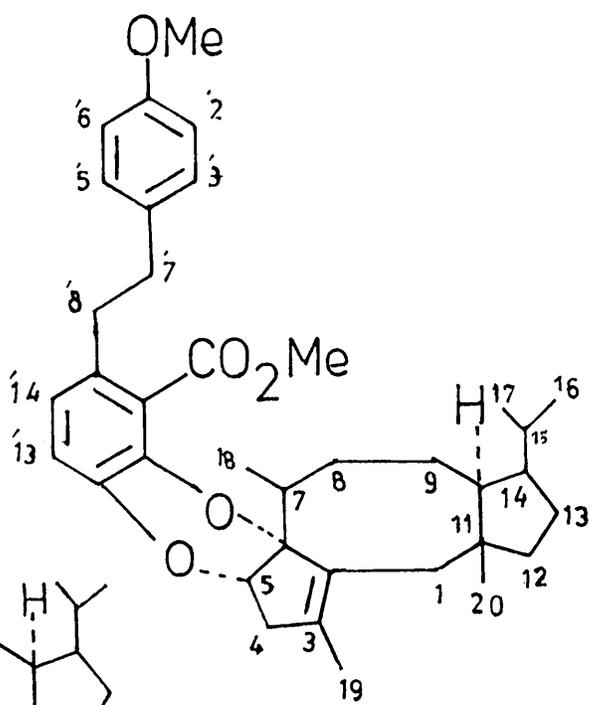
(189)



(190)



(191)



(192)

and its spectroscopic properties are very similar to those of (192). The  $^1\text{H}$  and  $^{13}\text{C}$  chemical shifts of the two compounds are listed in table for comparison. The shifts of the diterpenoid portion are virtually identical in the two cases. Only in the case of the lower ring of the bibenzyl ring are differences apparent. Thus the protons of the AB system shift from  $\delta_{\text{H}}$  6.76 and 6.56 in A to  $\delta_{\text{H}}$  6.72 and 6.60 in B. Comparison of the  $^{13}\text{C}$  spectrum reveals small shift differences in all of the carbon of the lower aromatic ring. The absence of  $^{13}\text{C}$  shift in the diterpenoid portions of (191) and (192) excludes the possibility of configurational differences. The only remaining possibility is regioisomerism i.e. in the formation of the dioxane ring oxygen-3 has become bonded to C-5 of the diterpenoid and oxygen-4 to C-6. Thus spinuloplagin B has structure (191).

Spinuloplagin A and B are unusual natural products. It is not immediately apparent how the dioxan ring is formed. A similar type of system has been reported<sup>90</sup> in nilgherron A (193) which is a diterpenoid dimer. The formation of these compounds is perhaps similar to the formation of the coumarinolignan<sup>91</sup> (194) which has been synthesised by both chemical and enzymatic oxidation of a dihydroxycoumarin in the presence of a phenylpropene.

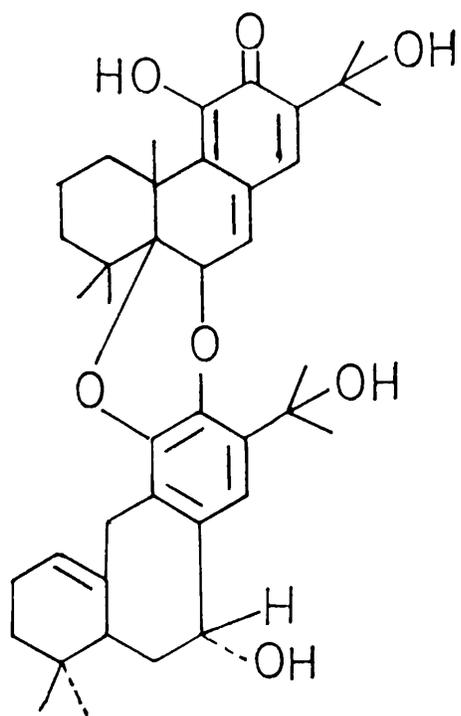
$^1\text{H}$  n.m.r. chemical shifts of spinuloplagins A (192) and B (191)

H	(191)	(192)
3', 5'	7.05 (d, J8.7 Hz.)	7.07 (d, J8.7Hz.)
2', 6'	6.80 (d, J8.7Hz.)	6.80 (d, J8.7Hz.)
13'	6.72 (d, J8.3 Hz.)	6.76 (d, J8.3Hz.)
14'	6.60 (d, J8.3Hz.)	6.56 (d, J8.3Hz.)
5	4.89 (t, J7.75Hz.)	4.87 (t, J7.75Hz.)
OCH <sub>3</sub>	3.89 (s)	3.87 (s)
CO <sub>2</sub> CH <sub>3</sub>	3.77 (s)	3.78 (s)
7', 8'	2.75 (m, 4 H)	2.76 (m, 4 H)
19	1.60 (s)	1.61 (s)
18	1.11 (d, J7.0Hz.)	1.11 (d, J6.9Hz.)
16 } 17 }	0.89 (d, J6.6Hz.) 0.83 (d, J6.6Hz.)	0.90 (d, J6.7Hz.) 0.84 (d, J6.6Hz.)
20	0.72 (s)	0.73 (s)

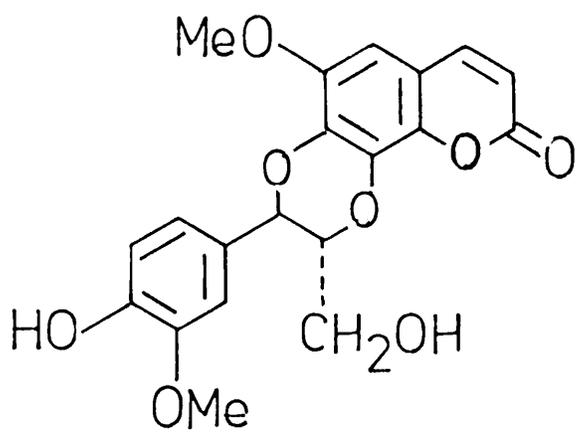
$^{13}\text{C}$  n.m.r. chemical shifts of spinuloplagins A (192) and B (191)

C	(191)	(192)
$\text{CO}_2\text{Me}$	168.3	168.3
1'	157.8	157.8
11'	145.1	144.6
12'	139.4	140.2
2	137.9	137.5
3	136.2	136.2
9'	134.0	134.0
4'	131.1	132.6
3', 5'	129.3 (2d)	129.3 (2d)
14'	122.2 (d)	120.7 (d)
10'	119.1	122.3
13'	117.5 (d)	118.8 (d)
2', 6'	113.7 (2d)	113.7 (2d)
6	93.2	94.1
5	74.9	74.7
$\text{OCH}_3$	55.2	55.2
$\text{CO}_2\text{CH}_3$	52.1	51.9
d	48.6	48.6
d	47.0	46.9
s	45.7	45.7
t	44.1	44.1

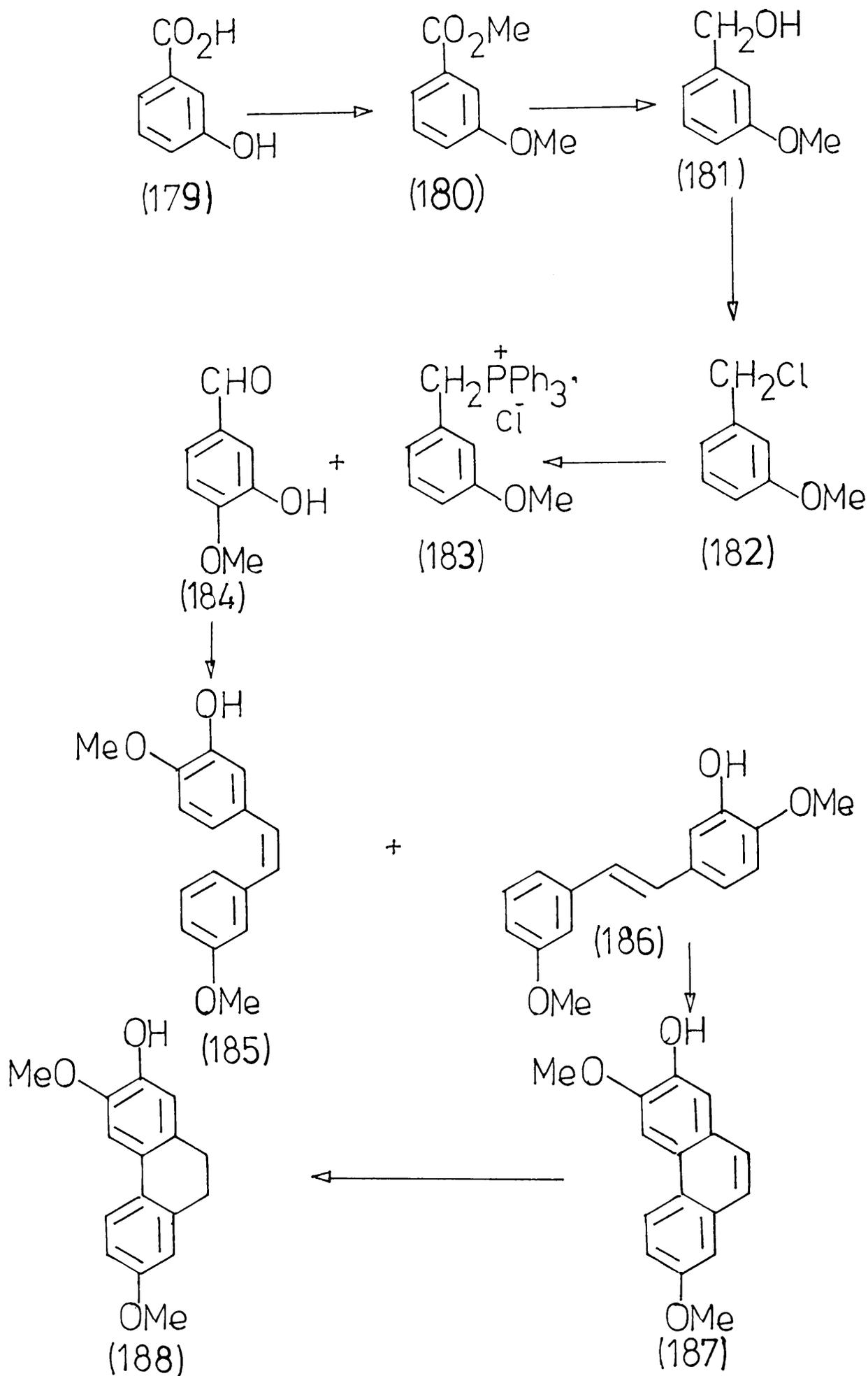
C	(191)	(192)
t	38.6	38.5
7' }	37.0	37.0
8' }	35.6	35.7
d	33.8	33.7
t	30.2	30.1
d	28.0	28.0
t + q	24.6	24.6
t	23.0	23.0
q	20.0	20.1
q	19.3	19.3
q	16.7	16.8
q	15.4	15.4



(193)



(194)



Synthesis of 2-hydroxy-3,7-dimethoxy-9,10-dihydro-phenanthrene

The liverwort Plagiochila spinulosa, a less common species in Scotland, has yielded 2-hydroxy-3,4,7-trimethoxy-9,10-dihydrophenanthrene (165) and the related dihydrophenanthrenes (166) - (170). Since 2-hydroxy-3,7-dimethoxy-9,10-dihydrophenanthrene (169) showed some activity in antifungal tests, we decided to synthesise it in order to obtain sufficient quantities for further biological evaluation.

3-Hydroxybenzoic acid (179) was methylated using dimethyl sulphate and anhydrous potassium carbonate in acetone to give methyl 3-methoxybenzoate (180) [ $\delta_{\text{H}}$  3.88 and 3.80 (each 3H, s, oMe)]. Reduction of (180) with lithium aluminium hydride yielded the desired 3-methoxybenzyl alcohol (181) [ $\delta_{\text{H}}$  4.45 (2H, s)] which was readily converted to the corresponding benzyl chloride (182) [ $\delta_{\text{H}}$  4.46 (2H, s)] by reaction with re-distilled thionyl chloride. The benzyl chloride (182) was transformed into the phosphonium chloride (183) by treatment with triphenylphosphine under anhydrous conditions in toluene for 46 hours. Its  $^1\text{H}$  n.m.r. spectrum showed the expected coupling of the benzylic methylene protons with phosphorous [ $\delta_{\text{H}}$  5.42 (d, J14Hz.)]. A Wittig reaction of isovanillin (184) with the phosphonium chloride (183) afforded the expected stilbene as a mixture of Z- (185) and E- (186) isomers. The  $^1\text{H}$  n.m.r. spectrum showed overlapping aromatic

protons at [ $\delta_{\text{H}}$  7.35 - 6.75] and a broad signal at  $\delta_{\text{H}}$  5.66 exchangeable with  $\text{D}_2\text{O}$ , together with two methoxyl groups [ $\delta_{\text{H}}$  3.88 and 3.86]. The mass spectrum showed a molecular ion at  $m/z$  256 corresponding to the molecular formula  $\text{C}_{16}\text{H}_{16}\text{O}_3$ .

The stilbene mixture was transformed into the corresponding phenanthrene (187) by photocyclisation with a 450 w u.v. lamp in an immersion well type quartz reactor under dry conditions in methanol<sup>+I<sub>2</sub></sup>. The mixture was then washed with sodium thiosulphate to remove excess iodine. Flash column chromatography followed by prep. t.l.c. afforded the desired phenanthrene (187) in low yield. Its spectroscopic properties were consistent with structure (187). Thus, the i.r. spectrum showed a broad band at  $\bar{\nu}_{\text{max}}$  3410  $\text{cm}^{-1}$ . In the  $^1\text{H}$  n.m.r. spectrum, the aromatic protons [ $\delta_{\text{H}}$  7.45 - 6.80] were not resolved, but a singlet at  $\delta_{\text{H}}$  5.42, exchangeable with  $\text{D}_2\text{O}$ , and two methoxy groups [ $\delta_{\text{H}}$  3.84 and 3.78] were apparent. The mass spectrum showed the expected molecular ion at  $m/z$  254 ( $\text{C}_{16}\text{H}_{14}\text{O}_3$  requires 254).

Hydrogenation of phenanthrenes should occur at the 9, 10- double bond<sup>88</sup>. The phenanthrene (187) was converted to the corresponding dihydrophenanthrene (188) by hydrogenation in presence of 10% palladised charcoal in acetic acid. The reaction was allowed to continue for three days. After separation by prep. t.l.c., the 9,10- dihydrophenanthrene (188) was obtained in 10% overall yield. Its structure was confirmed by its  $^1\text{H}$  n.m.r. spectrum which showed the appearance

of a new broad multiplet signal at  $\delta_{\text{H}}$  2.46 due to four protons of C-9 and C-10 and which was identical with that of the natural product.

EXPERIMENTAL

Plagochila spinulosa, collected near Aberfoyle (Scotland) was dried, ground and extracted in a Soxhlet apparatus with hexane, Chromatography of the crude extract over alumina (grade H deactivated) afforded an initial separation. Subsequent purification by preparative t.l.c. afforded seven natural products.

( i) 2- Hydroxy- 3,4,7- trimethoxy- 9,10- dihydrophenanthrene (165), (120 mg) was the major component. It was purified by crystallisation from chloroform: petroleum followed by prep. t.l.c. and sublimation at ~~92~~<sup>92</sup>/0.1 mm. It had m.p.  $132^{\circ} - 35^{\circ}$  [lit.  $135 - 139^{\circ}$ ]<sup>89</sup>.

$\delta_{\text{H}}$ : 8.17 (d, J8.4Hz., H-5), 6.84 (dd, J2.7, 8.8Hz., H-6), 6.76 (d, J2.9Hz., H-8), 6.62 (s, H-1), 5.67 (s, exchangeable with D<sub>2</sub>O), 3.96 (s, -OCH<sub>3</sub>), 3.73 (s, -OCH<sub>3</sub>), 2.69 (m., 4H).

M.S.: m/z 286 (M<sup>+</sup> base peak), (C<sub>17</sub>H<sub>18</sub>O<sub>4</sub> requires 286).

( ii) 2- Hydroxy- 3,7- dimethoxy- 9,10- dihydrophenanthrene (169), (14 mg) was purified by prep. t.l.c. followed by sublimation at 105<sup>o</sup>/0.3 mm as an oil.

$\delta_{\text{H}}$ : ~~8.18 (d, J8.7Hz., H-5), 7.45 (s, 1H), 7.367(d, J2.7Hz., H-8), 7.08 (dd, J2.6, 8.5Hz., H-6), 6.66 (s, 1H), 5.42 (s, exchangeable with D<sub>2</sub>O), 3.82 (s, -OCH<sub>3</sub>), 3.71 (s, -OCH<sub>3</sub>), 2.69 m, 4H).~~

≡ (188) h61

M.S.:  $m/z$  256.1088 ( $C_{16}H_{16}O_3$  requires 256.2994).

(iii) 2,3,7-Trimethoxy-9,10-dihydrophenanthrene (170), (9 mg.) was purified by prep. t.l.c., followed by sublimation at  $85^\circ/0.3$  mm as an oil.

$\delta_H$ : 7.58 (d, J8.8Hz., H-5), 7.28 (s, 1H), 7.22 (s, 1H), 6.92 (d, J2.7Hz., H-8), 6.78 (dd, J2.6, 7.6Hz., H-6), 3.88 (s,  $-OCH_3$ ), 3.86 (s,  $-OCH_3$ ), 3.76 (s,  $-OCH_3$ ), 2.84 (m, 4H).

M.S.:  $m/z$  270 ( $M^+$ , base peak), ( $C_{17}H_{18}O_3$  requires 270).

(iv) 3,4,7-Trimethoxy-9,10-dihydrophenanthrene (167), (12 mg.), m.p.  $91 - 92^\circ$ , was purified by prep. t.l.c. followed by sublimation (twice) at  $90^\circ/0.2$  mm, then crystallisation from ethyl acetate:ether.

$\delta_H$ : 8.37 (d, J8.6Hz., H-5), 6.94 (d, J8.5Hz.), 6.83 (dd, J8.6, 2.8Hz., H-6), 6.76 (d, J2.7Hz., H-8), 6.68 (d, J8.6 Hz.), 3.88 (s,  $-OCH_3$ ), 3.84 (s,  $-OCH_3$ ), 3.78 (s,  $-OCH_3$ ), 2.76 (m, 4H)

M.S.:  $m/z$  270.1259 ( $C_{17}H_{18}O_3$  requires 270.1256).

( v) 4- Hydroxy- 3,7- dimethoxy- 9,10- dihydrophenanthrene (168), (8 mg.) was purified by prep. t.l.c. It could not be crystallised.

$\delta$   
H: 8.37 (d, J8.7Hz., H-5), 7.46 (d, J8.5Hz.), 6.86 (d, J2.6Hz., H-8), 6.74 (d, J8.2Hz.), 6.69 (d, J8.0Hz.), 6.65 (dd, J2.4, 8.6Hz., H-6), 3.84 (s,  $-\text{OCH}_3$ ), 3.65 (s,  $-\text{OCH}_3$ ), 3.56 (s, exchangeable with  $\text{D}_2\text{O}$ ).

( vi) Methyl 2- methyl- 3,4- methylenedioxy- 6- methoxy- benzoate (189), (8 mg.) was isolated as an amorphous solid. It was purified by prep. t.l.c. followed by sublimation at  $86^\circ/0.3$  mm, and had m.p.  $80^\circ$ .

$\delta$   
H: 6.29 (s, 1H), 5.92 (s, 2H), 3.87 (s,  $-\text{CO}_2\text{CH}_3$ )  
3.75 (s,  $-\text{OCH}_3$ ), 2.02 (s, 3H).

M.S.:  $m/z$  224 ( $\text{M}^+$ , base peak), ( $\text{C}_{11}\text{H}_{12}\text{O}_5$  requires 224)

(vii) Spinuloplugin B (191), (8 mg.) was isolated and purified by prep. t.l.c. as an oil [see table for spectroscopic properties].

#### Methyl 3- methoxybenzoate (180)

A mixture of 3- hydroxybenzoic acid (20 g., 0.144 mole), anhydrous potassium carbonate (50 g., 0.5 mole) and dimethyl sulphate (40 ml.) in dry acetone (200 ml.) was refluxed for 5 hr. The acetone was removed and the residue diluted with

H<sub>2</sub>O (250 ml) and ether (250 ml). The organic layer was washed with concentrated ammonia solution (2 x 40 ml), H<sub>2</sub>O (50 ml) and then dried over anhydrous magnesium sulphate. The solvent was evaporated to yield a yellow oil which on distillation at 75°/0.2 mm gave methyl 3-methoxybenzoate (180) (21 g, 87%).

$\delta_{\text{H}}$ : 7.64 (dd, J2.7Hz., 2.8Hz. H-2), 7.41 (dd, J8.1, 8.9Hz. H-5)  
7.16 (ddd, J2.4, 3.1, 8.4 Hz. H-4 or H-6), 7.12 (ddd, J2.5, 2.7, 8.4 Hz. H-4 or H-6), 3.88 (s, -OCH<sub>3</sub>), 3.80 (s, -OCH<sub>3</sub>).

### 3-Methoxybenzyl alcohol (181)<sup>c</sup>

A solution of methyl 3-methoxybenzoate (180) (21 g, 0.126 mole) in dry ether (50 ml) was added over a period of 20 min to a solution of lithium aluminium hydride (5g, 0.14 mole) in dry ether (10 ml). Gentle reflux was continued for 3 hr. The chilled mixture was then decomposed by careful addition of aqueous sodium sulphate. The aqueous phase was extracted thoroughly with ether and the combined organic layers were washed with 5% sodium bicarbonate and dried over sodium sulphate. Distillation afforded (181) (13 g, 75%) as a viscous oil. b.p. 93° - 95°/0.3 mm.

$\delta_{\text{H}}$ : 7.26 - 6.62 (4H), 4.98 (br, exchangeable with D<sub>2</sub>O),  
4.45 (s, -CH<sub>2</sub>OH), 3.52 (s, -OCH<sub>3</sub>)

3-Methoxybenzyl Chloride (182):<sup>y</sup>

A stirred solution of 3-methoxybenzyl alcohol (181) (13 g, 0.094 mole) and dimethylaniline (15 g, 0.124 mole) in anhydrous toluene (100 ml) was slowly treated with redistilled thionyl chloride ( <sup>0.100</sup> mole) in toluene (10 ml). The mixture was warmed to room temperature, and then heated under reflux for 1 hr; cooled and acidified with dilute hydrochloric acid. The organic layer was separated, washed, dried and evaporated to give a brown oil which on distillation afforded 3-methoxybenzyl chloride (182) (11.8 g, 80%) as a colourless oil b.p. 95°.

$\delta_{\text{H}}$ : 7.32 - 6.84 (4H), 4.46 (s,  $-\text{CH}_2\text{Cl}$ ), 3.72 (s,  $-\text{OCH}_3$ )

3-Methoxybenzyl triphenylphosphonium chloride (183):

The benzyl chloride (182) (11 g, 0.07 mole) and triphenylphosphine (20 g, 0.076 mole) were stirred and heated in anhydrous toluene under  $\text{N}_2$  at 60° for 46 hr. The precipitated hygroscopic phosphonium chloride (183) (16 g, 54%) was separated by filtration, washed with light petroleum ether, then dried in vacuo. It had m.p. 254° - 56°.

$\delta_{\text{H}}$ : 7.68 (brm, 15H), 7.28 - 6.74 (4H), 5.42 (d, J14Hz.  $-\text{CH}_2$  ), 3.52 (s,  $-\text{OCH}_3$ ).

3- Hydroxy- 4, 3'- dimethoxystilbene:

3- Methoxybenzylphosphonium chloride (183) (1.7 g,  $4.06 \times 10^{-3}$  mole) and sodium hydride NaH (6 g, 0.25 mole) were stirred as a suspension in dry THF (30 ml) under nitrogen for 2 hours. A solution of 3- hydroxy- 4- methoxybenzaldehyde (isovanillin) (184) (0.9 g,  $5.92 \times 10^{-3}$  mole) was added in dry THF (8 ml) and the mixture was stirred for 18 hours at ambient temperature. The mixture was acidified with dil. HCl and extracted with ether (2 x 50 ml). The ether extract on evaporation gave a pale yellow oil (2 g, 84%) which showed three main spots [r.f. = 0.5, 0.3 and 0.2] on an analytical t.l.c. These spots were separated by flash column chromatography, eluting with light petroleum containing increasing proportions of ethyl acetate. 10 Fractions were collected. Fractions 2 - 6 were combined and crystallised from  $\text{CHCl}_3$  - light petroleum to give a crystalline product m.p.  $123^\circ - 124^\circ\text{C}$  (R.f. 0.5) corresponding to the stilbene mixture.

$\delta_{\text{H}}$ : 7.36 - 6.72 (9H), 5.66 (s, exchangeable with  $\text{D}_2\text{O}$ ),  
3.88 (s,  $-\text{OCH}_3$ ), 3.86 (s,  $-\text{OCH}_3$ ).

M.S.:  $m/z$  256 (base peak,  $\text{C}_{16}\text{H}_{16}\text{O}_3$  requires 256)

Fractions 7 - 9 contained unchanged isovanillin (R.f. 0.3) and  $\text{Ph}_3\text{P} = \text{O}$  (R.F. 0.2).

2-Hydroxy-3,7-dimethoxyphenanthrene (187)

The stilbene mixture (2 g, 0.023 mole) was dissolved in methanol (600 ml) containing iodine (1 g) and irradiated for 24 hr with a 450w u.v. lamp in an immersion well type quartz reactor under dry N<sub>2</sub>. The methanol was evaporated and the brown residual oil dissolved in EtoAC (100 ml) and washed sodium thiosulphate (2 x 50 ml). The organic layer was then washed with brine, dried over magnesium sulphate and evaporated to give a brown solid (1.5 g) which showed 2 main spots on t.l.c. These spots were separated by flash column chromatography in petroleum: EtoAC followed by prep. t.l.c. The more polar spot proved to be the phenanthrene (187) (0.4 g, 20%) m.p. 125°C.

$\delta_{\text{H}}$ : 7.45 - 6.80 (aromatic protons, **7H**), 5.42 (s, exchangeable with D<sub>2</sub>O), 3.84 (s, -OCH<sub>3</sub>), 3.78 (s, -OCH<sub>3</sub>).

$\delta_{\text{C}}$ : 145.30 (s), 144.93 (s), 129.15 (s), 128.78 (s), 126.66 (d), 124.95 (s), 121.73 (s), 119.84 (d), 117.03 (s), 114.73 (d), 113.95 (d), 111.62 (d), 110.92 (d), 110.43 (d), 55.96 (q), 55.21 (q).

M.S.: m/z 254 (C<sub>16</sub>H<sub>14</sub>O<sub>3</sub> requires 254).

2-Hydroxy- 3,7- dimethoxy- 9,10- dihydrophenanthrene (188)

The crude phenanthrene (187) (0.4 g,  $1.5 \times 10^{-3}$  mole) and 10% palladised charcoal (200 mg) in acetic acid were stirred under hydrogen at room temperature for 3 days. The reaction mixture was filtered and the charcoal washed with dilute acetic acid. The solution was made alkaline with sodium bicarbonate, extracted with Et<sub>2</sub>O, dried and evaporated to leave a yellow oil. Preparative t.l.c. gave 3 main spots.

Bands 1 and 3 were not investigated further.

Band 2: 2- hydroxy- 3,7- dimethoxy- 9,10- dihydrophenanthrene (188) (0.04 g, 10%).

$\delta_{\text{H}}$ : 7.52 - 7.12 (5 H), 4.30 (s, exchangeable with D<sub>2</sub>O)  
3.84 (s, -OCH<sub>3</sub>), 3.80 (s, -OCH<sub>3</sub>), 2.46 (4H, m).

M.S.:  $m/z$  256 (C<sub>16</sub>H<sub>16</sub>O<sub>3</sub> requires 256).

C H A P T E R 4

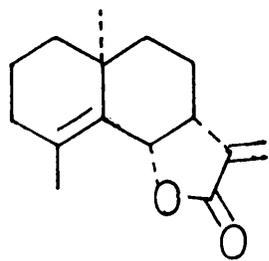
## INTRODUCTION

The Frullaniaceae consists mainly of the larger genus Frullania and the smaller Jubula.

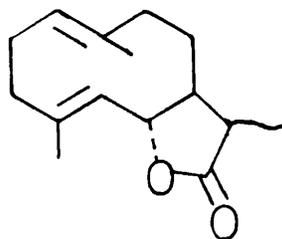
### Frullania

Frullania species are very rich sources of sesquiterpene lactones and bibenzyls, and there are more than 1000 species known in the world. Frullania species are very interesting from the viewpoint of medicinal chemistry since they produce sesquiterpene lactones which cause intense allergic contact dermatitis<sup>24</sup> and have antitumoral and plant growth regulatory activity. Of the many species known, eleven have been reported to be contact sensitising<sup>93</sup>. Ourisson and co-workers<sup>94</sup> isolated the active component (-)- frullanolide (32) from Frullania tamarisci. The enantiomeric compound (+)- frullanolide (195) was also isolated from Frullania dilatata<sup>24</sup>. Since then, a large number of other compounds have been reported from Frullania species. Connolly and Thornton<sup>26</sup> published the isolation of the compounds (34) - (36) together with (32) from the Scottish liverwort F. tamarisci.

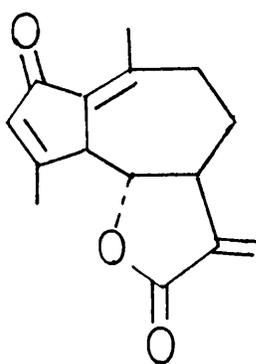
Asakawa et al<sup>52</sup> have investigated twenty-five Frullania species and concluded that they can be divided into five chemotypes depending on the classes of metabolites found.



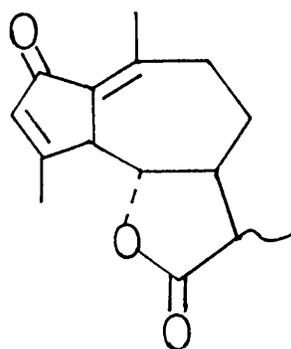
(195)



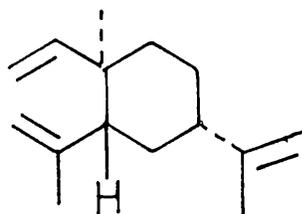
(196)



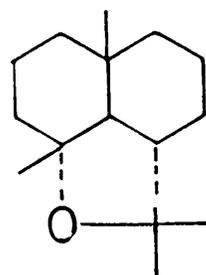
(197)



(198)



(199)



(200)

( i) Sesquiterpene Lactone- Bibenzyl type:

This type is typified by F. tamarisci and F. dilatata. Sesquiterpene lactones such as frullanolide (195),  $\gamma$ -cyclo-costunolide (34) and costunolide (35) are the major components together with bibenzyls and other sesquiterpenoids as minor components.

( ii) Sesquiterpene Lactone type:

This category produces a large quantity of sesquiterpene lactones but no bibenzyls. These sesquiterpenoids are usually germacranolides e.g. (35), (196) and guaianolides e.g. (197) - (198) rather than eudesmanolides.

( iii) Bibenzyl type:

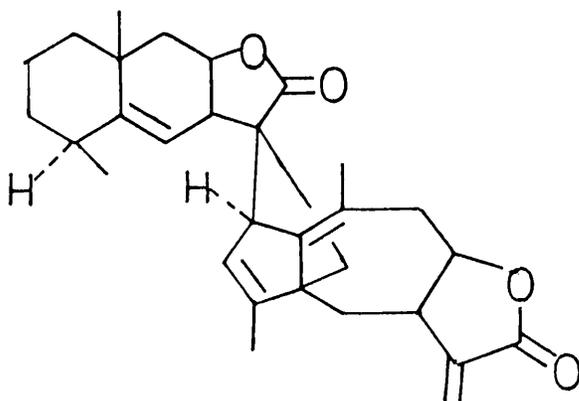
The Frullania species of this type contain bibenzyls e.g. (87) as major constituents and lack sesquiterpenoid lactones.

( iv) Monoterpene type:

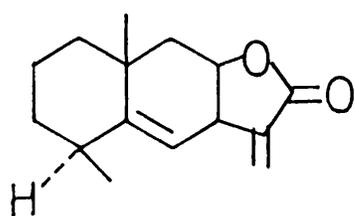
Monoterpene hydrocarbons are present in thirteen of twenty-five Frullania species. Most of the species contain  $\alpha$ - and  $\beta$ - pinenes (7) - (8) and camphene (9), but they do not contain any bibenzyls or sesquiterpenoid lactones.

( v) Cyclocolorenone type:

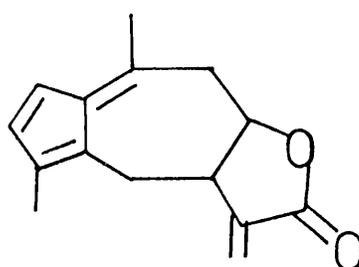
This category contains only one member, Frullania diversitexta. It produces mainly cyclocolorenone (13) and a few diterpenoid acetates.



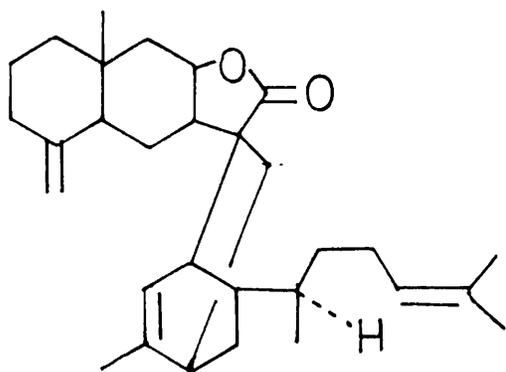
(201)



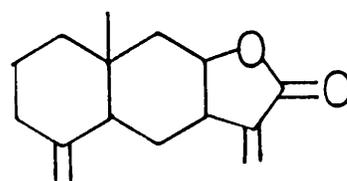
(202)



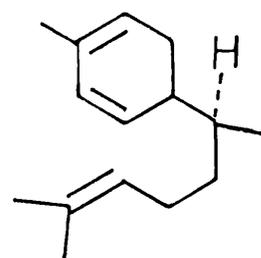
(203)



(204)



(205)



(206)

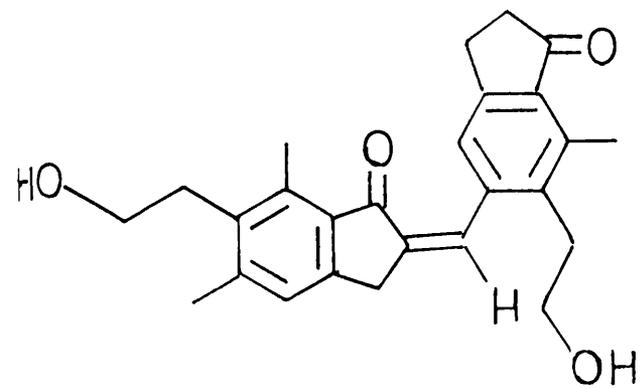
## Jubula

Jubula japonica contains sesquiterpenoids, cyclocolorenone (13),  $\beta$ -elemene (199) and maalioxide (200) all of which have been detected by gcms<sup>95</sup>.

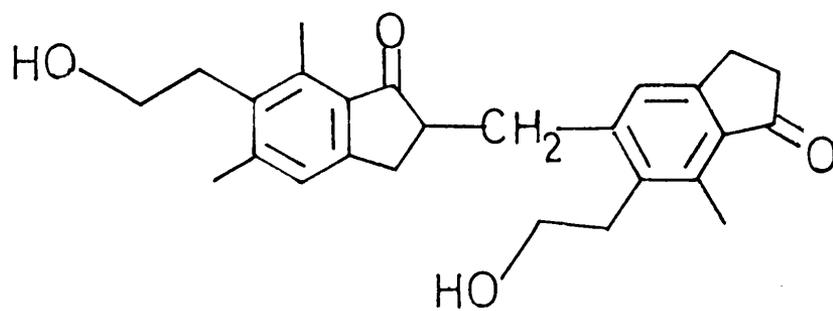
Dimeric sesquiterpenoids are not uncommon in the plant kingdom. The most common mode of dimerisation involves the Diels Alder reaction. The dimer (201) from the roots of Rudbeckia laciniata, recently published by Jakupovic<sup>96</sup>, illustrates this point. It clearly arises by Diels Alder reaction of the precursors (202) and (203). The dimer (204) from Helenium autumnale<sup>97</sup> provides another example. It has been synthesised by reaction of isoalantolactone (205) with zingiberene (206).

Aldol condensation provides another possible mechanism of dimerisation<sup>98</sup>. The dinorsesquiterpenoid dimers monachosorins A (207), B (208) and C (209) can be formally derived by aldol condensation of 6-(2-hydroxyethyl)-5,7-dimethylindan-1-one (210) and the 5-formyl compound (211). They occur in the fronds of Monachosorum arakii along with mukagolactone (212).

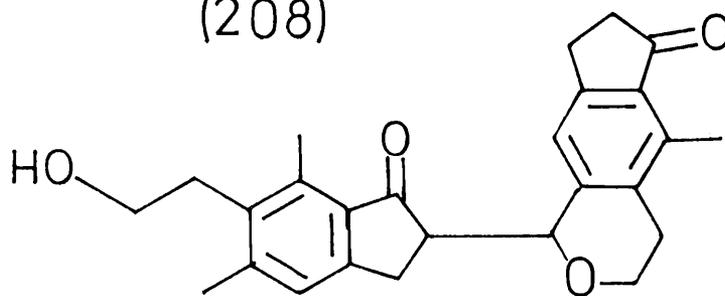
During our work on Frullania tamarisci we isolated a new dimeric sesquiterpenoid\* formed by a different kind of mechanism. Its structure forms the basis of this section.



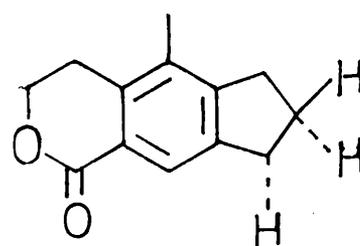
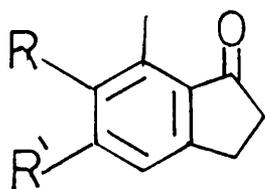
(207)



(208)



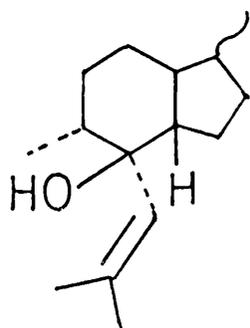
(209)



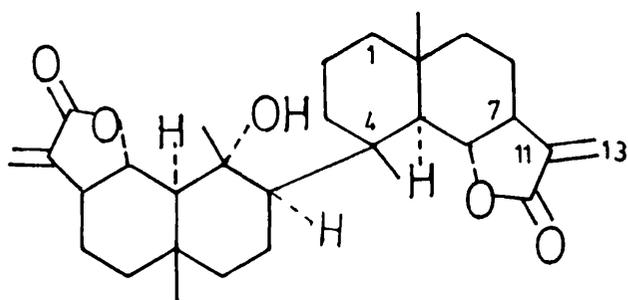
(212)

(210)  $R = \text{CH}_2\text{CH}_2\text{OH}$ ,  $R' = \text{CH}_3$

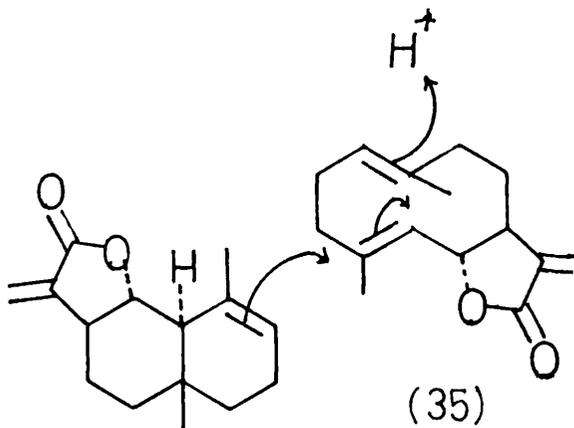
(211)  $R = \text{CH}_2\text{CH}_2\text{OH}$ ,  $R' = \text{CHO}$



(213)



(214)



(36)

(35)

## DISCUSSION

The Scottish liverwort Frullania tamarisci was previously investigated by Connolly and Thornton<sup>26</sup>. Further work by Connolly et al<sup>99</sup> yielded, additionally to the previous known compounds, the unusual sesquiterpenoid tamariscol (213) as the major constituent. In the present work, both (-)- frullanolide (32) and tamariscol (213) were obtained. The alcohol (213), a pungent oil, C<sub>15</sub>H<sub>16</sub>O (m/z 222), [ $\bar{\nu}_{\max}$  3610 cm<sup>-1</sup> (free hydroxyl)] has in its <sup>1</sup>H n.m.r. spectrum signals for a trisubstituted double bond [ $\delta_{\text{H}}$  5.12 (m, J1.4Hz.), two vinyl methyls [1.92 (d, J1.2Hz.) and 1.79 (d, J1.6Hz.)] and two secondary methyls [ $\delta_{\text{H}}$  0.96 (d, J6.6Hz.) and 0.89 (d, J6.6Hz.)]. These together with four methine and four methylene groups constitute a bicarbocyclic system. The structure of (213) was eventually solved using 2 D INADEQUATE.

An interesting dimeric sesquiterpenoid lactone (214), C<sub>30</sub>H<sub>42</sub>O<sub>5</sub> ( $\bar{\nu}_{\max}$  3580 and 1765 cm<sup>-1</sup>) was also isolated. The <sup>1</sup>H n.m.r. spectrum of (214) is similar to that of the sesquiterpenoid lactone (36) especially with respect to the protons associated with the trans-fused exomethylene  $\gamma$ - lactone. Thus the exomethylene protons appear as doublets at  $\delta_{\text{H}}$  6.65 (J3.2Hz.), 6.48 (J3.2Hz.), 5.42 (J3.0Hz.) and 5.34 (J3.0Hz.) consistent with the presence of typical conjugated lactone systems. The lactone termini, H-6 and H-'6, are overlapping triplets at  $\delta_{\text{H}}$  3.96 and 4.04 (each t, J11.2Hz.).

A triplet of quartets (J11.2, 3.2Hz.) can be assigned to the protons attached to C-7 and C-7'. Finally there are two proton doublets at  $\delta_{\text{H}}$  1.94 and 1.90 (J11.2Hz.) due to H-5 and H-5'. Double irradiation experiments readily confirmed the assignments made above. The other features of the  $^1\text{H}$  n.m.r. spectrum include a hydroxyl proton [ $\delta_{\text{H}}$  3.2 (br, exchangeable with  $\text{D}_2\text{O}$ )] and four tertiary methyl groups at  $\delta_{\text{H}}$  1.26, 1.25, 1.06 and 1.05. These data, in conjunction with the molecular formula and the  $^{13}\text{C}$  spectrum, clearly indicate that the dimer consists of two eudesmanolide systems with the same trans-fused lactone as in  $\gamma$ -cyclocostunolide (36) and with C-4 fully substituted in both monomeric units. The  $^{13}\text{C}$  spectrum [see experimental] reveals signals for thirty carbons, confirming that the dimer is not symmetrical. There are five doublets, nine triplets and three singlets among the non-oxygenated carbons together with one tertiary oxygenated carbon. This tally suggests that the two units are joined together as in (214). The formation of (214) is perhaps best envisaged as involving costunolide and  $\gamma$ -cyclocostunolide, both known constituents of F. tamarisci. The electrophile resulting from cyclisation of costunolide may attack the double bond of  $\alpha$ -cyclocostunolide instead of losing a proton. Subsequent attack by water then gives the dimer.

Determination of the relative stereochemistry of the dimer poses some problems. It is not possible to observe H-3' and to measure its couplings. It seems reasonable, however, to assume that the sesquiterpenoid substituent will be equatorial while H-3' is axial. NoE difference experiments failed to provide a definitive answer. Significant NoEs were observed between H-7, 7' and H-5, H5' and between H-6, 6' and the highfield methyl signals (3H-14, 14'). Irradiation of H-6, 6' also apparently gave NoEs at 3H-15, 15' although it is not possible to be certain that both methyls were affected. The reverse experiment is difficult to do without irradiating other protons. It seems likely, however, that the stereochemistry of dimer (214) is as shown.

This dimer is the first to be reported from the *Hepaticae*.

EXPERIMENTAL

Plant material (1 kg) was collected in the West of Scotland. After drying, it was finely ground and extracted in a Soxhlet apparatus first with ether and then methanol to give a combined crude extract (35 g). A large scale chromatography over alumina (grade H deactivated) in a solvent system of Et<sub>2</sub>O:EtOAc (100% - 0% ether), followed by a sephadex column of (i) CH<sub>2</sub>Cl<sub>3</sub>:hexane (4:1) and (ii) CHCl<sub>3</sub>:MeOH (1:1) and followed also by prep. t.l.c. afforded the following compounds.

(-)- Frullanolide (32) : It was recrystallised from CHCl<sub>3</sub>-light petroleum as needles m.p. 73° - 74° [lit. 74° - 76°]<sup>26</sup> (450 mg).

$\delta_{\text{H}}$ : 6.17 (d, J1Hz., H-13), 5.46 (d, J1Hz., H-13'),  
5.12 (d, J5Hz., H-6), 1.88 (brm., H-7),  
1.28 (s, 3H-15), 1.13 (s, 3H-14).

Tamariscol (213): It was obtained as a pungent oil (130 mg),  
 $\bar{\nu}_{\text{max}}(\text{CCl}_4)$  3610 cm<sup>-1</sup>.

$\delta_{\text{H}}$ : 5.12 (m, J1.4Hz., H-10), 1.92 (d, J1.2Hz., 3H-12),  
1.79 (d, J1.6Hz., 3H-13), 0.96 (d, J6.6Hz.,  
secondary methyl), 0.89 (d, J6.6Hz., secondary  
methyl).

Dimeric lactone (214): This was the most polar. It was recrystallised from MeOH and had m.p.  $207^{\circ} - 208^{\circ}$  (25 mg).

$\nu_{\max}^{(\text{CCl}_4)}$   $3580 \text{ cm}^{-1}$ ,  $1765 \text{ cm}^{-1}$  ( $\gamma$ -lactone).

$\delta_{\text{H}}$ : 6.65 (d,  $J_{3.2\text{Hz.}}$ , H-13,  $13'$ ), 6.48 (d,  $J_{3.2\text{Hz.}}$ , H-13<sup>1</sup>,  $13^1$ ), 5.42 (d,  $J_{3.2\text{Hz.}}$ , 2H, exomethylene protons)  
 5.34 (d,  $J_{2.9\text{Hz.}}$ , 2H, exomethylene protons),  
 4.04 (t,  $J_{11.19\text{Hz.}}$ , H-6), 3.96 (t,  $J_{11.24\text{Hz.}}$ , H-6<sup>'</sup>),  
 2.59(2H, tq,  $J_{11.0, 3.3\text{Hz.}}$ ), 3.2 (br, exchangeable with  $\text{D}_2\text{O}$ ), 1.94 (d,  $J_{11.3\text{Hz.}}$ , H-5), 1.90 (d,  $J_{10.71\text{Hz.}}$ , H-5), 1.27 (s,  $-\text{CH}_3$ ), 1.26 (s,  $-\text{CH}_3$ ), 1.06 (s,  $-\text{CH}_3$ )  
 1.05 (s,  $-\text{CH}_3$ ).

$\delta_{\text{C}}$ : 170.1 (s), 169.6 (s), 139.4 (s), 138.5 (s), 117.8 (t),  
 116.9 (t), 82.1 (d), 81.9 (d), 71.6 (s), 59.7 (d),  
 55.7 (d), 53.1 (d), 51.1 (d), 49.3 (d), 45.1 (st),  
 45.0 (t), 42.3 (s), 41.8 (t), 40.8 (t), 40.5 (t),  
 38.0 (s), 34.7 (t), 24.4 (q), 23.8 (q), 22.8 (t),  
 22.5 (t), 22.2 (t), 21.4 (q), 19.0 (q), 18.1 (t)

References

1. W.H. Perkin J. Chem. Soc., 1904, 654.
2. G. Garnier, L. Bezaniger-Beauguesne and G. Debraux  
"Resources Medicinal de la flore française" Vigot  
frères Editeurs, Paris 1969, Vol. 1, p. 78.
3. G. Lopes, Dr. Pharm. Thesis, University of Bordeaux  
1955.
4. Z. Pavletic and B. Stilonovic, Acta Bot. Croat., 1963,  
22, 133.
5. K. Muller, Hoppe-Seyler's Z. Physiol. Chem., 1905, 45,  
299.
6. Y. Fujita, T. Ueda and T. Ono Nippon Kagaku Zasshi,  
1956, 77, 400.
7. K. Markham and L. Porter, Prog. Phytochem., 1978, 5,  
181.
8. J. Connolly, Rev. Latinoam. Quim., 1981, 12, 121.
9. Y. Asakawa, Fortschr. Chem. Org. Naturst., 1982, 42, 1.
10. S. Huneck, Manual of Bryology, 1983, in the press.
11. C. Suire, Y. Asakawa, M. Toyota and T. Takemoto,  
Phytochemistry, 1982, 21, 349.
12. W. Karrer, E. Cherbuliez and C.H. Eugster in "Konstitu-  
tion und Vorkommen der Organischen Pflanzenstoffe"  
Birkhauser, Basel, 1977, Chemische Reihe Band 17, p. 30.
13. Y. Asakawa, M. Toyota, T. Aratani, Proc. Bryol. Soc.  
Jpn., 1976, 1, 155
14. S. Huneck and E. Klein, Phytochemistry, 1976, 6, 383.
15. A. Matsuo, M. Nakayamo, S. Sato, T. Takemoto, S. Uto and  
S. Hattori Experientia, 1974, 30, 321.

16. Y. Asakawa, M. Toyota and T. Takemoto Phytochemistry, 1978, 17, 457.
17. Y. Asakawa, H. Inoue, M. Toyota and T. Takemoto, Phytochemistry, 1980, 19, 3623
18. Y. Asakawa, M. Toyota, T. Takemoto, I. Kubo and K. Nakanishi, Phytochemistry, 1980, 19, 2147.
19. A. Matsuo, S. Sato, M. Nakayama and S. Hayashi, J. Chem. Soc. Perkin Trans. I, 1979, 2652.
20. M. Nakayama, S. Ohira, S. Shinke, Y. Matsushita, A. Matsuo and S. Hayashi, Chem. Letters. 1979, 1245.
21. J. Connolly, A. Harding and I. Thornton, J. Chem. Soc., Perkin Trans. 1, 1974, 2487.
22. Matsuo, T. Maeda, M. Nakayama and S. Hattori, Tetrahedron Lett. 1973, 4131.
23. S. Welsh, S. Chayabunjonglerd and A.S.C.P. Rao, J. Org. Chem., 1980, 45, 4086.
24. G. Perold, J. Muller and G. Ourisson, Tetrahedron, 1972, 28, 5797.
25. A. Greene, J. Muller and G. Ourisson, Tetrahedron Lett., 1972, 2489.
26. J. Connolly and I. Thornton, Phytochemistry, 1973, 12, 631.
27. Y. Asakawa, G. Ourisson and T. Arantani, Tetrahedron Lett., 1975, 3957.
28. Y. Asakawa, M. Toyota, T. Takemoto and C. Suire, Phytochemistry, 1979, 18, 1007.

29. Y. Asakawa, R. Matsuda, M. Toyota, T. Takemoto,  
J. Connolly and W. Phillips, Phytochemistry, 1983, 22,  
961.
30. V. Benesova, Z. Samek, V. Herout and F. Sorm, Collect.  
Czech. Chem. Commun., 1969, 34, 582.
31. A. Corbella, P. Gariboldi, G. Jommi, F. Orsini,  
A. DeMaro and A. Immirzi, J. Chem. Soc. Perkin Trans. 1,  
1974, 1875.
32. Y. Asakawa, M. Toyota and T. Aratani, Tetrahedron Lett.,  
1976, 3619.
33. Y. Asakawa, M. Toyota, T. Takemoto and C. Suire,  
Phytochemistry, 1979, 18, 1349.
34. Y. Asakawa, J. Connolly, C. Fakunle, D. Rycroft and  
M. Toyota, J. Chem. Research, 1987, 82.
35. M. Toyota, F. Nagashima and Y. Asakawa, Phytochemistry,  
1988, 27, 1789.
36. J. Connolly and I. Thornton, J. Chem. Soc. Perkin Trans. 1,  
1973, 736.
37. M. Przybylska and F. Ahmed Acta Crystallogr. Sect. B.  
1977, 33, 366.
38. I. Benes, T. Vanek and M. Budesinky, Collect, Czech.  
Chem. Commun., 1982, 47, 1873.
39. Y. Asakawa, C. Suire, M. Toyota, N. Tokunaga, T. Takemoto,  
S. Hattori and M. Mizutani, J. Hattori Bot. lab., 1980,  
48, 285.

40. Y. Asakawa, T. Takemoto, M. Toyota and T. Aratani, Tetrahedron Lett., 1977, 1407.
41. S. Huneck and K. Overton, Phytochemistry, 1971, 10, 3279.
42. J. Connolly, Rev. Latinaom. Quim., 1981, 12, 121.
43. V. Amico, R. Currenti, G. Oriente, M. Piattelli and C. Tringali, Phytochemistry, 1981, 20, 848.
44. S. Huneck, G. Baxter, A. Cameron, J. Connolly and D. Rycroft, Tetrahedron Lett., 1983, 3787.
45. D. Barrow, D. Barton, E. Chain, U. Ohnorge and R. Sharma, J. Chem. Soc. Perkin Trans. 1, 1973, 1590.
46. T. Sassa and M. Togashi, Agric. Biol. Chem., 1973, 37, 1505.
47. A. Matsuo, H. Nozaki, M. Nakayama, S. Hayashi and D. Takaoka, J. Chem. Soc., Chem. Commun., 1978, 198.
48. Y. Fukuyama, T. Masuya, M. Tori, M. Kido, M. Wakamatsu and Y. Asakawa, Phytochemistry, 1988, 27, 1797.
49. A. Caldicott and G. Eglinton, Phytochemistry, 1976, 15, 1139.
50. Y. Asakawa, M. Toyota and T. Takemoto, Experientia, 1978, 34, 155.
51. A. Matsuo, M. Nakayama and S. Hattori, Z. Naturforschung, 1971, 26B, 1023.
52. Y. Asakawa, R. Matsuda, M. Toyota, S. Hattori and G. Ourisson, Phytochemistry, 1981, 20, 2187.

53. Y. Asakawa, T. Takikawa and M. Tori, Phytochemistry, 1987, 26, 1023.
54. V. Benesova and V. Harout, Collect. Czech. Chem. Commun., 1972, 37, 1764.
55. Y. Asakawa, M. Toyota and T. Takemoto, Phytochemistry, 1978, 17, 2005 ..
56. M. Nakayama, A. Matsuo, T. Kami and S. Hayashi, Phytochemistry, 1979, 18, 328.
57. M. Gleizes, G. Pauly and C. Suire, Botaniste, 1972, 55, 339.
58. A. Matsuo, O. Ishii, M. Suzuki, M. Nakayama and S. Hayashi; 24th Symposium on The Chemistry of Terpenes, Essential Oils and Aromatics. Koriyama, Japan, 1980, symposium papers, p. 224.
59. Y. Asakawa, R. Matsuda, M. Toyota, C. Suire, T. Takemoto and S. Hattori: 24th Symposium on the Chemistry of Terpenes, Essential Oils and Aromatics. Yamaguchi, Japan 1981, symposium paper, p. 92.
60. Y. Asakawa, M. Toyota and L. Harrison, Phytochemistry, 1985, 24, 1505.
61. I.F.M. Valio, R.S. Burden and W.W. Schwabe, Nature 1969, 223, 1176.
62. K. Fries, Beit. Biol. Pflanz., 1964, 40, 177.
63. J. Graham, Phytochemistry, 1977, 16, 249.
64. Y. Asakawa, N. Takunaga, M. Toyota, T. Takemoto and C. Suire, J. Hattori Bot. Lab., 1979, 45, 395.

65. Y. Asakawa, N. Takunaga, M. Toyota, T. Takemoto, S. Hattori, M. Mizutani and C. Suire, J. Hattori Bot. Lab. 1979, 46, 67.
66. S. Huneck, Tetrahedron, 1976, 32, 109.
67. Y. Asakawa, R. Matsuda and A. Cheminat, Phytochemistry 1987, 26, 1117.
68. A. Setsuko and Y. Ohta, Phytochemistry, 1984, 23, 1379.
69. Y. Arai, T. Kamikawa and T. Kubota, Tetrahedron Letters, 1972, 1615.
70. S. Huneck and K. Schreiber, Phytochemistry 1977, 16, 1013.
71. R.J. Pryce, Phytochemistry, 1971, 10, 2679.
72. R.J. Pryce, Phytochemistry, 1972, 11, 1355.
73. J.D. Connolly, P. Ifeadike, S. Singh, <sup>and A.C. Free,</sup> unpublished work.
74. S.F. MacDonald, J. Chem. Soc., 1948, 376.
75. G.R. Pettit, S. Singh, M. Niven, E. Hamel and J. Schmidt, J. Nat. Prods., 1987, 50(1), 119.
76. R. Pschorr, Chem. Ber., 1896, 29, 496.
77. Fisher and Schmidt, Chem. Ber., 1894, 27, 2786.
78. D.R. Dalton, and A. Annama, Syn. Comm., 1972, 2(5), 303.
79. D.F. DeTar and Y. Wen Chu, J. Amer. Chem. Soc., 1954, 76, 1686.
80. E. Perham and Y. Seyad, J. Org. Chem., 1974, 39, 2051.
81. R. Pschorr and C. Sumuleanu, Chem. Ber., 1899, 32, 3405
82. F.W. Hoffman, Chem. Abst., 1952, 46, 5617.
83. S.R. Bhandari, A. Kapadi, P. Mujumder, M. Joardar and J. Shoolery, Phytochemistry, 1985, 24, 801.

84. Y. Asakawa, M. Tori, K. Takikawa, H. Krishnamurty and S.Kantikar, Phytochemistry, 1987, 26, 1811.
85. P. Majumder and M. Joardar, Indian J. Chem., 1985, 24B, 1192.
86. P. Majumder and S. Banarje, Phytochemistry, 1988, 27, 245.
87. F. Stermitz, T. Suess, C. Schauer and D. Anderson, J. Nat. Prod., 1983, 46, 417.
88. M. Sargent, J. Chem. Soc. Perkin Trans. 1, 1984, 1919.
89. L. Harrison, Ph.D. Thesis, University of Glasgow, 1983.
90. T. Miyase, P. Ruedi and C. Eugster, Helv. Chim. Acta., 1977, 60, 2789.
91. L. -J. Lin and G. Cordell, J. Chem. Soc., Chem. Commun., 1984, 160.
92. Y. Asakawa, M. Toyota, T. Takemoto, H. Fujiki and T. Sugimura, Planta Med., 39, 233.
93. J. Mitchell, B. Fritig, B. Singh and G. Towers, J. Invest. Dermatol., 1970, 54, 223.
94. H. Knoche, G. Ourisson, G. Perold, J. Fousterreau and J. Maleville, Science, 1969, 166, 239.
95. Y. Asakawa, N. Takunaya, T. Takemoto, S. Hattori, M. Mizutani and C. Suire, J. Hattori Bot. Lab., 1980, 47, 153.
96. J. Jakupovic, Y. Jia, R.King and F. Bohlmann, Liebigs Ann. Chem., 1986, 1474.
97. R. Matusch and H. Haberlein, Liebigs Ann. Chem., 1987, 455.

98. T. Satake, T. Murakami, N. Yokote, Y. Seiki and C. Chen, Chem. Pharm. Bull., 1985, 33, 4175.
99. J. Connolly, L. Harrison and D. Rycroft, Tetrahedron Lett., 1984, 25, 1401.

Supplementary References

- A. Merck Index, Tenth Edition, 1983, 687.
- B. R.L. Shriner and C.J. Hull, J. Org. Chem., 1945, 10, 228.
- C. C. Mettler, Berichte, 1906, 39, 2939.
- D. J.W. Cornforth and R. Robinson, J. Chem. Soc., 1942, 684.

